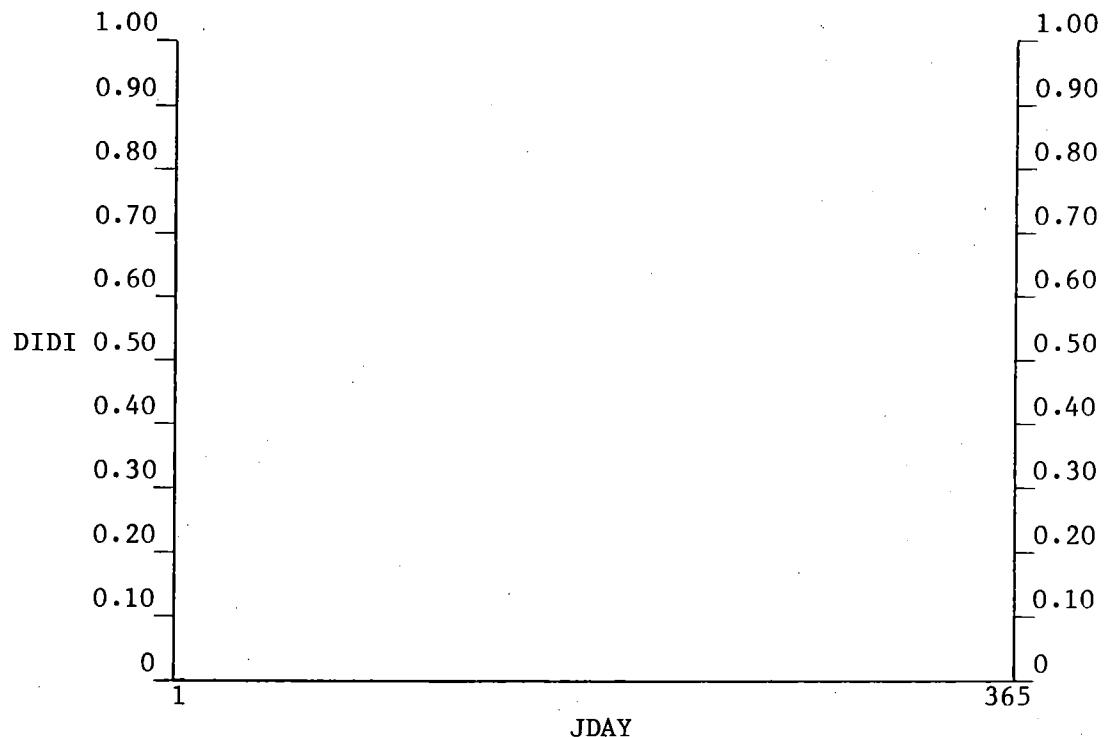


TOPIC 3. DIET DIGESTIBILITIES

General trends in diet digestibilities follow the general trends in the cell structures of the plants. The stages and parts of plant growth that have thinner and less lignified cell walls are, for the most part, more digestible than those stages and parts with more lignified cell walls. Cell chemistry also affects digestibility, however. Tannins, for example, act as inhibitors of digestion. Changes in cell structure occur as plant phenology changes over the growing season. Emerging, growing tissue cannot have rigid cell walls, for new tissue is being added as cells increase in both number and size. When the numbers and sizes of cells in plant tissue have both reached maximum, cell maturation occurs and cell walls increase in thickness and rigidity. The cells in stems become very rigid and serve as supporting tissue. Cells in leaf tissue mature, become decadent, and the leaf falls to the ground. Flower petals mature, wither, and fall. Functional changes in different plant parts are accompanied by structural changes in the cells, and these changes affect nutritive relationships between animal and range.

The concepts underlying relationships between cell structure and digestibility permit one to generalize on seasonal variations in diet digestibility. Consumption of decadent lignified dormant forage results in stable diet digestibilities. As the growing season progresses, diet digestibilities increase as new growth makes up an increasing proportion of the diet. As the growing season progresses and plants mature, diet digestibilities begin to drop until they reach the annual low when only decadent lignified forage is available again. Patterns of seasonal change are illustrated below.



Free-ranging ruminants are generally quite selective feeders, but there are times and locations where preferred species and parts are limited. In the spring, for example, new growth becomes available at a rate that is dependent on species phenology and growing conditions. As the snow melts, animals can supplement the winter diet of dormant forage with new growth as it becomes available, and overall diet digestibilities increase slowly. As plant growth continues, larger amounts of new growth become available and diet digestibilities increase more rapidly. It is important to realize that this pattern of digestibility coefficients applies to the diet rather than individual plants. Digestibilities change in relation to the phenology of the plant, and new plant tissue makes up increasingly larger proportions of the diet in the first part of the growing season and smaller proportions in the last part.

Digestibilities are measured by in vivo or in vitro methods. The former involves live animals and the latter, laboratory procedures. In vivo measurements were conducted in the early years of nutrition experimentation, and in vitro has become an accepted technique in later years. These two kinds of measurements are discussed and references given in UNITS 3.1 and 3.2. When direct measurements of particular diets are not available, then calculated diet digestibilities may be determined (UNIT 3.3). Reasonable estimates of digestibilities should be possible for all wild ruminants if general knowledge of nutritive processes is used to make the estimate.

UNIT 3.1: IN VIVO MEASUREMENTS OF DIGESTIBILITIES

There have been many experimental feeding trials of wild ruminants on different forages, with concomitant collection of feces and urine in order to determine at least apparent digestibilities. Results of these are published in the long lists of references that follow this UNIT.

Some of the empirical measurements are made in live animals consuming prescribed, often single-species diets. These measurements are not only expensive because collecting the current annual growth of browse plants is a very slow process. One very convincing exercise illustrating foraging conditions in the winter is the collection of a daily supply of dry weight current annual growth on a cold day with a clippers and bag, especially on an overbrowsed range. The cost of clipping just the CAG is prohibitive if wages must be paid. Volunteers have helped collect on some research projects. Ruminants must be on the test forages for several days before measurements are actually made to allow previous diet residues to pass through the gastrointestinal tract. Thus a five-day trial must last about two weeks total.

Another way to conduct in vivo trials is by collecting the entire plant and letting the test animals browse the parts and amounts desired. It is harder to determine the amounts eaten using this procedure, but less expensive to collect and feed the browse.

In vivo measurements are difficult to complete because wild ruminants do not consume single-species diets very readily, especially of lower quality forage. Single species tests eliminate the beneficial effects of associative digestibilities, resulting in even lower estimates of digestibilities of the lower-quality forages. A mix of forages provides a more suitable substrate for rumen microflora than a single-species substrate.

In vivo measurements of digestibilities have been tried using nylon bag techniques which permit tests of selected forages in a bag suspended in the rumen of a fistulated animal. This technique reduces the expense of large-scale collections, and alleviates the problem of consumption of low-palatability forages and single-species diets. The bag does impede rumen fluid circulation and sorting, so the results are not exactly as they would be if the forage were free in the rumen.

In vivo digestibilities give the impression of biological reality, but all test results must be considered as estimates that vary in relation to a large number of test conditions. Results should be evaluated as patterns in an ecological context.

REFERENCES, UNIT 3.1

IN VIVO MEASUREMENTS OF DIGESTIBILITIES

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JWMAA 41--4 667 676 od-- seas nutr yield, dig, pine blair,rm; short,/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CJZOA 52-10 1201 1205 odvi dig, prox comp, wint brows mautz,ww; silver/ 1974

JANSA 21--4 1017 1018 odvi diges, brwse, cedar, aspen youatt,wg; ullre/ 1961
JANSA 32--5 999 1002 odvi chrm-51, totl collctn tech mautz,ww 1971

JRMGA 29--1 82 83 odvi comparis in vivo, in vitro ruggiero,lf; whel 1976

JWMAA 5---1 108 114 odvi digestiv capacit of wt dee forbes,eb; marcy/ 1941
 JWMAA 28--4 791 797 odvi diges, cedar, aspen browse ullrey,de; youat/ 1964
 JWMAA 31--3 448 454 odvi dig, cedar, jack pine brow ullrey,de; youat/ 1967
 JWMAA 32--1 162 171 odvi dig, cedar, basalm fir brw ullrey,de; youat/ 1968
 JWMAA 33--3 482 490 odvi digest ener req, mich does ullrey,de; youatt 1969
 JWMAA 35--2 366 368 odvi confin eff, dry mattr dige mautz,ww 1971
 JWMAA 35--4 732 743 odvi limita, winter aspen brows ullrey,de, youat/ 1971
 JWMAA 36--3 885 891 odvi dig, metabol, aspen browse ullrey,de; youat/ 1972
 JWMAA 36--4 1052 1060 odvi var, determ diges capacity mothershead,cl; / 1972
 JWMAA 37--2 195 201 odvi dry matter, energ intak,dig ammann,ap; cowan/ 1973
 JWMAA 39--1 67 79 odvi feed analyses, digest, w-t robbins,ct; van / 1975
 JWMAA 40--4 630 638 odvi dige, 7 northern browse sp mautz,ww, silver/ 1976
 JWMAA 43--3 798 801 odvi dosh, compar digest capaci palmer,wl; cowan, 1979

UASPA 51--2 89 89 odvi compar in viv, in vitr dig ruggiero,l 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

TPCWD*14--- 1 89 odhe digestibil, forag sp, colo dietz,dr; udall,/ 1962

CAFGA 39--2 163 175 odhe nutr val, forag plnts,calif hagen,h1 1953
 CAFGA 41--1 57 78 odhe dig, naturl, artific foods bissell,hd; harr/ 1955

CWSPA 43--- 1 44 odhe vivo/vit rela, forag, colo milchunas,dg; dy/ 1978

JANSA 16--2 476 480 odhe dosh, live oak, chamis, ca bissell,hd; weir, 1957

odhe continued on the next page

*TPCWD is thought to be the correct CODEN for: State of Colorado - Dept. of Game and Fish Technical Publication.

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JRMGA 10--4 162 164 odhe nutr val, wntr brows plnts smith,ad 1957
JRMGA 12--1 8 13 odhe browse adequa, overwinteri smith,ad 1959
JRMGA 30--2 119 121 odhe comp in viv, in vitr diges urness,pj; smith/ 1977
JRMGA 30--2 122 127 odhe/evaluat deer habitat, nutr wallmo,oc; carpe/ 1977

JWIDA 10--2 166 169 odhe invest tansy ragwort poisn dean,re; winward, 1974

JWMAA 14--3 285 289 odhe sagebrush as a winter feed smisth,ad 1950
JWMAA 16--3 309 312 odhe digestibil, native forages smith,ad 1952
JWMAA 28--4 785 790 odhe effect essenti oils, rumen nagy,jg; steinho/ 1964
JWMAA 31--3 443 447 odhe prev diet, digest alfal ha nagy,jg; vidacs,/ 1967
JWMAA 34--4 964 967 odhe cell wall dig, forag value short,h1; reagor, 1970
JWMAA 38--4 823 829 odhe capabil, utiliz fibr alfal schoonveld,gg; n/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

BJNUA 40--2 347 358 ceel dosh, compar digest forags milne,ja; macrae/ 1978
HOECD 4---1 59 65 ceel caca, seas diff, dig brows cederlund,g; nyst 1981
NATUA 263-- 763 764 ceel intk, diges, vetega, scotl milne,ja; macrae/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CJBOA 54--9 966 970 alal qual, lowbsh crnbry, alask oldemeyer,jl; sem 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

BPURD 1---- 95 107 rata feeding experimnts, lichns jacobsen,e; skjen 1975
JWMAA 44--3 613 622 rata digestib, rangifer forages person,sj; pegau/ 1980

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

anam

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ATRLA 22-14 225 230 bibi doca, yak, fora intak, dig richmond,rj; hud/ 1977
IZYBA 16--- 54 57 bibi diges, pelleted diet, rumi hintz,hf; sedgew/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovca

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JRMGA 24--1 73 75 doca est dig ener, dry, org mat rittenhouse,lr; / 1971

JONUA 15--4 383 395 doca cellul, ligni, nutri value crampton,ew; mayn 1938

NEXAA 133-- 1 38 doca yucca, chamiza, rang suppl brown,ls 1922

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

AGJOA 51--3 226 234 dosh intak, dig tech, suppl fee harris,le; cook,/ 1959

JANSA 23--3 700 710 dosh nutritnl qual, blue grass reid,rl; jung,ga/ 1964

JBRGA 32--3 141 147 dosh comparsn meth predic diges milne,ja 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JWMAA 40--2 283 289 wld1 nutr qual, dig, seed, frui epps,ea,jr 1976

XAMPA 1147- 1 220 wld1 range, wild1 habit evaluat paulson,ha,ed; r/ 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ATRLA 18--3 81 91 caca natu feed, roe de, ingesti drozdz,a; osiecki 1973

ATRLA 24-13 137 170 caca seas intak, dig, nat foods drozdz,a; osiecki 1979

UNIT 3.2: IN VITRO MEASUREMENTS OF DIGESTIBILITIES

In vitro digestibility measurements have become common-place since the early 1960's. Tilley and Terry (1963) used in vitro fermentation followed by pepsin digestion, and Oh et al. (1966) found the two-step technique of Tilley and Terry to be an accurate predictor ($r = 0.88$) of in vivo ruminant digestion of legumes and grasses. Van Soest (1965) related in vitro results to cell wall components of the feed. These early studies have been followed by a fair number of measurements on white-tailed and mule deer, but few studies have been done on the other wild ruminants.

In vitro measurements of digestibilities involve the use of very small samples of forage (a gram or less) in a temperature-controlled fermentation bath that has been inoculated with rumen fluid. The fermentation bath is maintained at body temperature for about 48 hours while digestion takes place. In vitro equipment used at Cornell's Wildlife Ecology Laboratory is pictured in Moen (1973:150). Up to 12 samples may be fermented at one time with this equipment. Duplicate samples of each forage are used, so six species may be evaluated during each run.

The cost and biological problems associated with in vivo measurements are eliminated with in vitro studies. New problems arise, however, as in vitro systems are more or less closed systems without biological absorption and feedback. Thus, the single sample present as a substrate is different from the real situation in the rumen where periodic feeding during each 24-hour period results in additions of new material to the rumen. The more or less continuous rumination and fermentation in the live animal results in a steady flow of both metabolites through the gastrointestinal walls and undigested waste products through the gastrointestinal canal.

Differences in the results from in vivo and properly-completed in vitro studies are not great. The economic advantages to in vitro studies are great, however. Further, the source of inocula need not be a major consideration; results with inocula from a captive deer on an alfalfa diet were within 2% of in vivo determinations, from a wild deer, within 1/2%, and from a cow, within 3% (Robbins et al. 1975). These differences may not satisfy a nutrition specialist, but they are all well within the range of accuracy when estimating diets, populations, and other ecological parameters of free-ranging animals. The use of cow inocula alleviates the need to hold wild ruminants for inocula. A fistulated white-tailed deer was available for rumen fluid for several years (See Moen 1973:149), but the much smaller fistula opening and the lower fluid component of rumen material in the deer compared to the cow made it much more convenient to use the readily-available and easily-collected rumen fluid from a cow at Cornell's Department of Animal Science. If docile wild ruminants are available for fistulation, especially the larger ones such as elk, then they should be used of course.

The results of a large number of in vitro tests of different plant parts, of forages collected at different times of the year, of associative digestibilities, and of the effects of inhibitors make it possible to get a much more complete picture of nutritive processes and patterns through the year under a wide variety of range conditions than with in vivo measurements. The accuracy of in vitro determinations of digestibilities by wild ruminants are fully as great as the accuracies in determining food habits and other characteristics of free-ranging animals. Hence in vitro measurements offer particular advantages for work with wild ruminants that far outweigh the disadvantages. Simply stated, many important and revealing nutritive evaluations could not be made without in vitro digestion techniques.

I wish to thank Dr. Peter Van Soest, Department of Animal Science, Cornell University, for the assistance given students and staff at the Wildlife Ecology Laboratory over the years as we have measured forage unique to white-tailed deer in New York State.

LITERATURE CITED

- Moen, A. N. 1973. Wildlife ecology. W. H. Freeman and Company, San Francisco. 458 pp.
- Oh, H. K., B. R. Baumgardt, and J. M. Scholl. 1966. Evaluation of forages in the laboratory. V. Comparison of chemical analyses, solubility tests, and in vitro fermentation. J. Dairy Sci. 49:850-855.
- Robbins, C. T., P. J. Van Soest, W. W. Mautz, and A. N. Moen. 1975. Feed analyses and digestion with reference to white-tailed deer. J. Wildl. Manage. 39(1):67-79.
- Tilley, J. M. A. and R. A. Terry. 1963. A two-stage technique for the in vitro digestion of forage crops. J. Brit. Grassl. Soc. 18:104-111.
- Van Soest, P. J. 1965. Non-nutritive residues: A system of analysis for the replacement of crude fiber. J. Assoc. Off. Agric. Chem. 49:546-551.

REFERENCES, UNIT 3.2

IN VITRO MEASUREMENTS OF DIGESTIBILITIES

SERIALS

CODEN	VO-NU BEPA ENPA ANIM KEY WORDS-----	AUTHORS-----	YEAR
JANSA 39--1 248	248 odvi cow vs deer, inoculum sourc palmer,wl; cowan,	1974	
JRMGA 28--5 419	421 odvi in vitr solu, dry mttr dig uresk,dw; dietz,/	1975	
JRMGA 29--1 82	83 odvi comparis in vitro, in vivo ruggiero,l; whela	1976	
JWMAA 27--2 184	195 odvi rumen fermentation, energ short,hl	1963	
JWMAA 35--2 221	231 odvi cellulose dig, chem, missour torgerson,o; pfa/	1971	
JWMAA 35--3 469	475 odvi odhe, nutr, chappral, ariz urness,pj; green/	1971	
JWMAA 35--4 698	706 odvi forag dig, diet, uplnd rng short,hl	1971	
JWMAA 38--1 20	31 odvi in vitr dig, foods, missou snider,cc; asplun	1974	
JWMAA 39--1 67	79 odvi/feed analyses, digest, w-t robbins,ct; van /	1975	
JWMAA 39--2 337	341 odvi comp, digest, decid br, ne robbins,ct; moen,	1975	
JWMAA 40--2 301	307 odvi eff innoc source, in vitro palmer,wl; cowan/	1976	
JWMAA 43--3 650	656 odvi carbohyd, urea influ diges mccullough,y	1979	
JWMAA 43--3 788	790 odvi pH influen, in vitro diges burbank,rk; wool/	1979	
SWNAA 24--2 297	310 odvi botan comp, nutr cont diet everitt,jh; gonza	1979	
UASPA 51--2 89	89 odvi compar in viv/in vitr dige ruggiero,l	1974	

CODEN	VO-NU BEPA ENPA ANIM KEY WORDS-----	AUTHORS-----	YEAR
CJFRA 2---3 250	255 odhe doug fir genot, brows pref radwan,ma	1972	
CWSPA 43--- 1	44 odhe viv/vitr rela, forag, colo milchunas,dg; dy/	1978	
FOSCA 16--1 21	27 odhe dougl-fir, microb ferm, ca oh,jh; jones,mb;/	1970	
JRMGA 30--2 119	121 odhe comparis in vitro, in vivo urness,pj; smith/	1977	
JRMGA 30--2 122	127 odhe evaluat deer habitat, nutr wallmo,oc; carpe/	1977	
JRMGA 30--3 206	209 odhe fo hab, semi-des grass-shr short,hl	1977	
JWMAA 28--4 785	790 odhe eff essen oils,rumn micro nagy,jg; steinhof/	1964	
JWMAA 31--3 443	447 odhe diet and dig, alfalfa hay nagy,jg; vidacs,g/	1967	
JWMAA 34--4 964	967 odhe cell wall dig, forag value short,hl; reagor,	1970	
JWMAA 36--4 1341	1343 odhe maint rumen fluid, in vitr schwartz,cc; nagy	1972	
JWMAA 38--3 531	534 odhe pestici eff, in vitro dige schwartz,cc; nagy	1974	
JWMAA 39--4 670	673 odhe nutr, diet, pondr pine ran urness,pj; neff,/	1975	

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
JWMAA 35--4 681 688 ceel in vitr dig, wint for, wyo ward,al 1971
ZTTFA 24--4 200 204 ceel inhib rum cellulolys, bark prins,ra; geelen, 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
CJBOA 54--9 966 970 alal nutr qual,lo bush cran, al oldemeyer,jl; see 1976
JWMAA 41--3 533 542 alal browse qual, popula, kenai oldemeyer,jl; fr/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
BPURD 1---- 251 256 rata in vitro digest of forages person,sj; white/ 1975
JWMAA 44--3 613 622 rata in vitr, nylon-bag digesti person,sj; pegau/ 1980

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
JWMAA 41--2 161 168 anam/bibi, diet qual, for avail schwartz,cc; nag/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
JAPEA 11--2 489 497 bibi trophi ecol, shrtgras plai peden,dg; van dy/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
JWMAA 32--4 773 777 ovca dig, alpine tundra plants johnston,; bezau,/ 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JBRGA 18--2 104 111 doca dosh,2-stage tech, in vitr tilley,jma; terry 1963

JDSCA 44-12 2242 2249 doca eff partic size, cellu dig dehority,ba; john 1961
JDSCA 49--7 850 855 doca chem anal, solubili, fermn oh,hk; baumgardt, 19

JRMGA 21--1 5 7 doca in vitr dig, cattle, range hoehne,oe; clant/ 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JANSA 23--3 700 710 dosh nutrit qual, in viv, vitro reid,r1; jung ga/ 1964

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CPLSA 42--4 692 697 in vitro diges, festu asso bezeau,lm; johnst 1962

JRMGA 24--2 134 136 in vitr dige, native hay karn,jf; clanton/ 1971
JRMGA 29--1 63 65 many effect of 2,4-d on digestn thilenius,jf; br/ 1976

XAMPA 1147- 1 220 wldl range, wildl habit evaluat paulson,ha,ed; r/ 1970

CHAPTER 11, WORKSHEET 3.2a

In vitro browse digestibilities

The SERIALS lists include many references with data on digestibilities. The tabulation below includes results of in vitro measurements on dormant browse plants of New York State that have been evaluated at Cornell's Wildlife Ecology Laboratory. They are included here to provide digestibility data for a relatively large number of white-tailed deer browses in Northeastern United States. Additional species should be added from published papers.

<u>Common name</u>	<u>Scientific name</u>	<u>GENS</u> <u>SPEC</u>	Dry matter Digestibility Coefficient
Alternate leaved dogwood	<i>Cornus alternifolia</i>	corn alte	0.64 *
Apple	<i>Pyrus malus</i>	pyru malu	0.62 **
Arrowwood	<i>Viburnum dentatum</i>	vibu dent	0.45 **
Aspen	<i>Populus tremuloides</i>	popu trem	0.61 **
Balsam fir	<i>Abies balsamea</i>	abie bals	0.62 *
Basswood	<i>Tilia americana</i>	tili amer	0.57 **
Beech	<i>Fagus grandifolia</i>	fagu gran	0.40 **
Blackberry	<i>Rubus alleghaniensis</i>	rubu alle	0.48 *
Black cherry	<i>Prunus serotina</i>	prun sero	0.37 *
Boxelder	<i>Acer negundo</i>	acer negu	0.57 **
Bur oak	<i>Quercus macrocarpa</i>	quer macr	0.43 **
Cucumber tree	<i>Magnolia acuminata</i>	magn acum	0.53 **
Elderberry	<i>Sambucus canadensis</i>	samb cana	0.37 *
Elm	<i>Ulmus americana</i>	ulmu amer	0.38 *
Fire cherry	<i>Prunus pensylvanica</i>	prun pens	0.40 *
Gray birch	<i>Betula populifolia</i>	betu popu	0.47 *
Gray dogwood	<i>Cornus racemosa</i>	corn race	0.55 ***
Hawthorn	<i>Crataegus sp.</i>	crat ----	0.43 **
Hazelnut	<i>Corylus cornuta</i>	cory corn	0.51 **
Hemlock	<i>Tsuga canadensis</i>	tsug cana	0.70 **
Hop hornbeam	<i>Ostrya virginiana</i>	ostr virg	0.37 *
Lilac	<i>Syringa vulgaris</i>	syri vulg	0.46 *
Meadow sweet	<i>Spiraea sp.</i>	spir ----	0.35 *
Mountain maple	<i>Acer spicatum</i>	acer spic	0.49 *
Nannyberry	<i>Viburnum lentago</i>	vibu lent	0.54 **

Continued on the next page

Paper birch	<i>Betula papyrifera</i>	betu papy	0.56 **
Red maple	<i>Acer rubrum</i>	acer rubr	0.57 **
Red oak	<i>Quercus rubra</i>	quer rubr	0.42 **
Red osier dogwood	<i>Cornus stolonifera</i>	corn stol	0.51 **
Red raspberry	<i>Rubus idaeus</i>	rubu idae	0.42 *
Red spruce	<i>Picea rubens</i>	pice rube	0.55 *
Scotch pine	<i>Pinus sylvestris</i>	pins sylv	0.64 *
Shagbark hickory	<i>Carya ovata</i>	cary ovat	0.48 **
Speckled alder	<i>Alnus rugosa</i>	alnu ovat	0.30 *
Staghorn sumac	<i>Rhus typhina</i>	alnu rugo	0.55 **
Striped maple	<i>Acer pensylvanicum</i>	rhus typh	0.54 *
Sugar maple	<i>Acer saccharum</i>	acer pens	0.46 **
White ash	<i>Fraxinus americana</i>	acer sach	0.48 **
White cedar	<i>Thuja occidentalis</i>	frax amer	0.67 **
White pine	<i>Pinus strobus</i>	thuj occi	0.60 *
Wild grape	<i>Vitis sp.</i>	pins stro	0.38 *
Willow	<i>Salix sp.</i>	viti ----	0.53 *
Witch hazel	<i>Hamamelis virginiana</i>	sali ----	0.49 **
Yellow birch	<i>Betula alleghaniensis</i>	betu alle	0.33 *

* - Wildlife Ecology Laboratory, New York measurements for Delmar.

** - Robbins, C. T. 1973. The biological basis for the determination of carrying capacity. PhD Thesis, Cornell University, Ithaca, N.Y.
239 pp.

*** - WEL Job # 74-01.

UNIT 3.3: CALCULATED DIET DIGESTIBILITIES

Wild ruminants have mixtures of species in their rumens at any one time, resulting in overall diet digestibilities that represents the effects of all of the individual forage species combined. How can diet digestibilities be estimated when the digestibilities of each of the forages in the diet at any one time have not been measured?

The dynamic characteristics of wild ruminant diets can never be duplicated under controlled conditions. It is, therefore, inevitable that diet digestibilities must be calculated since there is no place to "look up" the digestibilities of selected natural diets.

How can diet digestibilities be calculated? First, diet components are tabulated from food habits studies. Second, the fractions of the total diet made up of the different components are determined from preference and rumen content studies. Third, estimates of the digestibilities of each forage and plant part for the time of year being analyzed are made based on published data, patterns useful for estimation and interpolation, and first approximations where necessary. Fourth, a weighted mean procedure is used to calculate overall diet digestibility.

Weighted means are determined by multiplying the forage digestibility coefficient (FDIC) of each forage and plant part by its fraction in the diet (FRDI). The sum of the forage digestibilities is the weighted mean diet digestibility (DIDI). A tabular format for this is illustrated below.

Forage

1. _____ FDIC x FRDI = _____

2. _____ FDIC x FRDI = _____

.

n. _____ FDIC x FRDI = _____

A sample calculation:

1. -----	$0.62 \times 0.15 = 0.09$
2. -----	$0.60 \times 0.20 = 0.12$
3. -----	$0.54 \times 0.25 = 0.14$
4. -----	$0.50 \times 0.40 = 0.20$

SUMS 1.00 0.55 = DIDI

This weighted mean procedure is useful for estimating the contributions of different forages based on animal preferences, for estimating the contributions of different plant parts in relation to their abundance in the rumen and digestibilities, and other effects of selective grazing or browsing. The procedure does not take interactions between different components of a diet, or associative digestibility effects, however. Highly digestible species may provide a suitable substrate that will increase the digestibilities of lower quality species. Another associative digestibility effect to consider is that of chemical inhibition. One species may contain inhibitors that not only result in a low digestibility of that species, but also reduce digestibilities of other species present in the rumen at the same time.

The weighted mean procedure is a good place to start evaluating diets. More research on actual diet mixtures taken by free-ranging wild ruminants would result in better understanding of nutritive processes, however. Even though we do not yet have ultimate knowledge of these processes, ecological processes continue and all components of the total picture should be considered in context. Thus we go on to new considerations in the remaining CHAPTERS of this book.

REFERENCES, UNIT 3.3

CALCULATED DIET DIGESTIBILITIES

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odvi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ceel

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

rata

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

anam

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovca

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CHAPTER 11, WORKSHEET 3.3a

Weighted mean diet digestibilities

This WORKSHEET presents a format for calculating weighted mean diet digestibilities. Ten blanks are provided: most diets are composed of as few as three or four most abundant forages. Use blanks 1 - 9 for the forage species that should be considered separately. Combine all the rest in the last blank.

Animal _____ Location _____ JDAY _____

FDIC x FRDI

1. _____ x _____ = _____

2. _____ x _____ = _____

3. _____ x _____ = _____

4. _____ x _____ = _____

5. _____ x _____ = _____

6. _____ x _____ = _____

7. _____ x _____ = _____

8. _____ x _____ = _____

9. _____ x _____ = _____

10. _____ x _____ = _____

SUMS _____ = DIDI

CLOSING COMMENTS

CHAPTER ELEVEN: FORAGE CHARACTERISTICS AND THE DIGESTIBILITY OF PLANT TISSUE, contains descriptions of some of the basic cell characteristics that affect the digestibilities of different species and plant parts through time. The relatively recent cellular approach to digestibilities was preceded by a century of chemical analyses of plant material. The relatively recent in vitro digestibility measurements have been preceded by thousands of in vivo trials on domestic ruminants, and a fair number on wild ruminants. Extensive literature lists covering both the earlier chemical composition and the later cell characteristics work, the earlier in vivo measurements and the later in vitro ones, have been included in this CHAPTER. The data in these references are needed when calculating forage consumption in CHAPTER 12.

GLOSSARY OF SYMBOLS USED - CHAPTER ELEVEN

ABUP = Absorbed but unused protein

ACTI = Activity

ADF = Acid detergent fiber

APDE = Apparent digestible energy

APDP = Apparent digestible protein

AUUE = Absorbed but unused urinary energy

CRPR = Crude protein

CSCP = Cell soluble content in percent of forage

CSDP = Cell soluble digestibility in percent

CSFF = Cell soluble fraction of the forage

CWCP = Cell wall content in percent of forage

CWDG = In vivo cell wall digestibility

CWDP = Cell wall digestibility in percent

CWFF = Cell wall fraction of the forage

DIDI = Diet digestibility

DMDP = Dry matter digestibility in percent

DORM = Dormant

EURN = Endogenous urinary nitrogen

FDIC = Forage digestibility coefficient

FEEN = Fecal energy

FEPR = Fecal protein

FLBU = Floral buds

FLOP = Flowers open

FRDI = Fraction of the diet digestibility

GREN = Gross energy

HEFE = Heat of fermentation

HNUM = Heat of nutrient metabolism

JDAY = Julian day

KCAL = Kilocalories

LCUC = Lignin-cutin content expressed as percent of the acid-detergent fiber

LEBU = Leaf buds

LEEM = Leaves emerging

LEFA = Leaves fallen

LEWI = Leaves withering

LGCC = Lignin-cutin content of the acid-detergent fiber

LGCC = Lignin-cutin content as a percent of the acid-detergent fiber

LGNC = Lignin content of the acid-detergent fiber

MAIN = Maintenance
MEEN = Metabolizable energy
MFEN = Metabolic fecal energy
MFNT = Metabolic fecal nitrogen
MNIC = Metabolizable nitrogenous compounds
MUEN = Metabolic urinary energy

NEEN = Net energy
NENS = Net nitrogen synthesized

PIUF = Protein in undigested forage
PLPA = Plant Part
PROD = Production
PTCW = Percent cell wall

REGU = Regulation

SFDI = Seeds and fruits dispersed
SFMT = Seeds and fruits maturing

TDIP = True digestible protein
TDMD = True dry matter digestibility in percent
TRDE = True digestible energy

UFOR = Undigested forage residue
UREN = Urinary energy
URNI = Urinary nitrogen

GLOSSARY OF CODENS - CHAPTER ELEVEN

AAAHA	Australian Journal of Experimental Agriculture and Animal Husbandry
ABSZA	Annales Botanici Societatis Zoologicae Botanicae Fenniae Vanamo
ADAGA	Advances in Agronomy
ADCSCA	Advances in Chemistry Series
AGJOA	Agronomy Journal(US)
AGNSA	New South Wales Agricultural Gazette
AJBOA	American Journal of Botany
AJCNA	American Journal of Clinical Nutrition
AMEBA	Annales Medicinae Experimentalis et Biologiae Fenniae
AMNAA	American Midland Naturalist
AMNTA	American Naturalist
AMSCA	American Scientist
APMBA	Applied Microbiology (US)
ATICA	Arctic (Canada)
ATRLA	Acta Theriologica (Poland)
AZFOA	Annales Zoologici Fennici (Finland)
AZWBA	Arizona Game and Fish Department Wildlife Bulletin
BIJOA	Biochemical Journal (England)
BJNUA	British Journal of Nutrition (England)
BOGAA	Botanical Gazette (US)
BOREA	Botanical Review (US)
BPURD	Biological Papers of the University of Alaska Special Report
BSECB	Biochemical Systematics and Ecology
BTBCA	Bulletin of the Torrey Botanical Club
BZSSA	Botanicheskii Zhurnal SSSR (USSR)
CAEBA	California Agricultural Experiment Station Bulletin
CAFGA	California Fish and Game
CFGGA	California Department of Fish and Game, Game Bulletin
CGFPA	Colorado Division of Game, Fish, and Parks Special Report
CJBOA	Canadian Journal of Botany
CJFRA	Canadian Journal of Forest Research (Canada)
CJZOA	Canadian Journal of Zoology
CNAPA	Canada Department of Agriculture Publication
CNDRA	Condor
CNJNA	Canadian Journal of Animal Science
CNRDA	Canadian Journal of Research, Section D, Zoological Sciences
COVEA	Cornell Veterinarian
CPLSA	Canadian Journal of Plant Science
CRPSA	Crop Science
CWRSB	Canadian Wildlife Service Report and Management Bulletin Series
CWSPA	Colorado Division of Wildlife Special Report
ECMOA	Ecological Monographs
ECOLA	Ecology
ELPLB	Ekologia Polska
ENDEA	Endeavour

FOSCA	Forest Science
FRCRA	Forestry Chronicle
FRSTA	Forestry
GRBNA	Great Basin Naturalist
HLTPA	Health Physics
HOECD	Holarctic Ecology
IZYBA	International Zoo Year Book
JACSA	Journal of the American Chemical Society
JAFCA	Journal of Agricultural Food and Chemistry
JAGRA	Journal of Agricultural Research
JANCA	Journal of the Association of Official Analytic Chemists
JANSA	Journal of Animal Science
JAPEA	Journal of Applied Ecology
JASIA	Journal of Agricultural Science
JBRGA	Journal of the British Grassland Society
JCECD	Journal of Chemical Ecology
JDSCA	Journal of Dairy Science
JEKOA	Journal of Ecology
JFUSA	Journal of Forestry
JOMAA	Journal of Mammalogy
JONUA	Journal of Nutrition
JPHAA	Journal of the American Pharmaceutical Association
JRMGA	Journal of Range Management
JSFAA	Journal of the Science of Food and Agriculture
JWIDA	Journal of Wildlife Diseases
JWMAA	Journal of Wildlife Management
LATBA	Louisiana Agricultural Experiment Station Bulletin
MDCBA	Minnesota Department of Conservation Technical Bulletin
MDCRA	Michigan Department of Conservation Game Division Report
MGLHA	Mitteilungen aus dem Gebiete derr Lebensmitteluntersuchung und hygiene
MLTBB	Maine Life Sciences and Agricultural Experiment Station Technical Bulletin
MUATA	Minnesota Agricultural Experiment Station Technical Bulletin
NAREA	Nutrition Abstracts and Reviews
NASRA	National Academy of Sciences--National Research Council, Publication
NATUA	Nature (England)
NAWTA	North American Wildlife and Natural Resources Conference, Transactions of the, NCANA Naturaliste Canadien, Le
NDFRA	North Dakota Farm Research
NETMA	Netherlands Journal of Agricultural Science
NEXAA	New Mexico Agricultural Experiment Station Bulletin
NEZFA	New Zealand Journal of Agricultural Research
NFGJA	New York Fish and Game Journal

- NOSCA Northwest Science
 NUABA Nevada Agricultural Experiment Station Bulletin

 OIKSA Oikos (Denmark)
 OJSCA Ohio Journal of Science

 PASHA Proceedings of the American Society for Horticultural Science
 PCGFA Proceedings of the Southeastern Association of Game and Fish Commissioners
 PLPHA Plant Physiology
 PLSOA Plant and Soil
 PMSCA Proceedings of the Minnesota Academy of Science
 PNASA Proceedings of the National Academy of Sciences of the United States
 PNUSA Proceedings of the Nutrition Society
 PSAFA Proceedings of the Society of American Foresters
 PYTCA Phytochemistry
 PZSLA Proceedings of the Zoological Society of London

 RAPHB Recent Advances in Phytochemistry

 SCIEA Science
 SJECA Soviet Journal of Ecology (English translation of Ekologiya)
 SOSCA Soil Science
 SSSAA Soil Science Society of America, Proceedings
 SSSJD Soil science Society of America Journal
 SWNAA Southwestern Naturalist
 SZSLA Symposia of the Zoological Society of London

 TAEBA Texas Agricultural Experiment Station Bulletin
 TAEMA Texas Agricultural Experiment Station Miscellaneous Publication
 TNWSD Transactions of the Northeast Section, The Wildlife Society
 TPCWD Colorado Division of Wildlife Technical Publication

 UABPA Biological Papers of the University of Alaska
 UASPA Proceedings of the Utah Academy of Sciences, Arts and Letters
 UAXBA Utah Agricultural Experiment Station Bulletin
 UCPZA University of California Publications in Zoology

 WAEBA Wyoming Agricultural Experiment Station Bulletin
 WCDBA Wisconsin Department of Natural Resources Technical Bulletin
 WGFBA Wyoming Game and Fish Commission Bulletin
 WLMOA Wildlife Monographs
 WMBAW Wildlife Management Bulletin (Ottawa) Series 1 (Canada)
 WUAPA Wisconsin Agricultural Experiment Station, Research Report
 WVAFA West Virginia Agriculture and Forestry
 WZMNA Wissenschaftliche Zeitschrift Karl-Marx Universitaet Leipzig
 Mathematisch-Naturwissenschaftliche Reihe

 XAAHA U S D A Agricultural Handbook
 XAGCA U S D A Circular
 XAMPA U S D A Miscellaneous Publication

XARRA U S Forest Service Research Note RM
XATBA U S D A Technical Bulletin
XFINA U S Forest Service Research Note INT
XFIPA U S Forest Service Research Paper INT
XFNSA U S Forest Service Research Note SO
XFPNA U S Forest Service Research Paper PNW
XFPSA U S Forest Service Research Paper SO
XFRMA U S Forest Service Research Paper RM
XPNWA U S Forest Service Research Note PNW

ZEJAA Zeitschrift fuer Jagdwissenschaft
ZTTFA Zeitschrift fuer Tierphysiologie Tierer naehrung und
Futtermittellkunde

LIST OF PUBLISHERS - CHAPTER ELEVEN

acpr	Academic Press	New York	nyny
agrc	Agricultural Research Council	London	loen
butt	Butterworth	Washington, D. C.	wadc
cdch	C. D. Church	Corvallis, OR	coor
dvnc	D. Van Nostrand Co.	New York	nyny
esli	E. and S. Livingstone, Publishers	Edinburgh, Great Britain	edgb
isup	Iowa State University Press	Ames, IO	amia
long	Longman	London	loen
mhbc	McGraw-Hill Book Company, Inc.	New York	nyny
mopc	Morrison Publishing Company	Ithaca, NY	itny
nasc	National Academy of Science	Washington, D. C.	wadc
nhfg	New Hampshire Fish & Game Dept.	Concord, NH	conh
olbo	Oliver and Boyd	Edinburgh, Scotland	edsc
prha	Prentice-Hall, Inc.	Englewood Cliffs, NJ	ecnj
spve	Springer-Verlaug Inc.	New York	nyny
stmp	St. Martin's Press	New York	nyny
whfr	W. H. Freeman Co.	San Francisco, CA	sfca

LIST OF WORKSHEETS - CHAPTER ELEVEN

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GLOSSARY OF PLANT CODE NAMES

GENS SPEC SCIENTIFIC NAME

abie amab	<i>Abies amabilis</i>
abie bals	<i>Abies balsamea</i>
abie conc	<i>Abies concolor</i>
abie gran	<i>Abies grandis</i>
abie lasi	<i>Abies lasiocarpa</i>
abie ----	<i>Abies</i>
abut inca	<i>Abutilon incanum</i>
abut theo	<i>Abutilon theophrasti</i>
acac angu	<i>Acacia angustissima</i>
acac farn	<i>Acacia farnesiana</i>
acac greg	<i>Acacia greggii</i>
acac roem	<i>Acacia roemeriana</i>
acac ----	<i>Acacia</i>
acal grac	<i>Acalypha gracilens</i>
acer circ	<i>Acer circinatum</i>
acer glau	<i>Acer glabrum</i>
acer macr	<i>Acer macrophyllum</i>
acer negu	<i>Acer negundo</i>
acer pens	<i>Acer pensylvanicum</i>
acer pseu	<i>Acer pseudoplatanus</i>
acer rubr	<i>Acer rubrum</i>
acer sacc	<i>Acer saccharinum</i>
acer sach	<i>Acer saccharum</i>
acer spic	<i>Acer spicatum</i>
acer ----	<i>Acer</i>
achi lanu	<i>Achillea lanulosa</i>
achi mill	<i>Achillea millefolium</i>
aden fasc	<i>Adenostoma fasciculatum</i>
aesc cali	<i>Aesculus californica</i>
aesc hipp	<i>Aesculus hippocastanum</i>
aesc pavi	<i>Aesculus pavia</i>
agar arve	<i>Agaricus arvensis</i>
agar bisp	<i>Agaricus bisporus</i>
agar camp	<i>Agaricus campestris</i>
agos glau	<i>Agoseris glauca</i>

agro smit	<i>Agropyron smithii</i>
agro spic	<i>Agropyron spicatum</i>
agro trac	<i>Agropyron trachycaulon</i> var. <i>majus</i>
agro trah	<i>Agropyron trachycaulon</i>
agro ----	<i>Agropyron</i>
agsr bore	<i>Agrostis borealis</i>
agsr idah	<i>Agrostis idahoensis</i>
agsr ----	<i>Agrostis</i>
albi juli	<i>Albizia julibrissin</i>
alec juba	<i>Alectoria jubata</i>
alec sarm	<i>Alectoria sarmentosa</i>
alec ----	<i>Alectoria</i>
alli text	<i>Allium textile</i>
alnu cris	<i>Alnus crispa</i>
alnu crmo	<i>Alnus crispa</i> var. <i>mollis</i>
alnu glut	<i>Alnus glutinosa</i>
alnu inca	<i>Alnus incana</i>
alnu rubr	<i>Alnus rubra</i>
alnu rugo	<i>Alnus rugosa</i>
alnu sinu	<i>Alnus sinuata</i>
alnu sito	<i>Alnus sitkensis</i>
alnu ----	<i>Alnus</i>
amar palm	<i>Amaranthus palmeri</i>
amar retr	<i>Amaranthus retroflexus</i>
amar ----	<i>Amaranthus</i>
ambr arte	<i>Ambrosia artemisiifolia</i>
ambr psil	<i>Ambrosia psilostachya</i>
ambr trif	<i>Ambrosia trifida</i>
amel alni	<i>Amelanchier alnifolia</i>
amel arbo	<i>Amelanchier arborea</i>
amel bart	<i>Amelanchier bartramiana</i>
amel cana	<i>Amelanchier canadensis</i>
amel flor	<i>Amelanchier florida</i>
amel spic	<i>Amelanchier spicata</i>
amel utah	<i>Amelanchier utahensis</i>
amel ----	<i>Amelanchier</i>
amor cane	<i>Amorpha canescens</i>
ampe arbo	<i>Ampelopsis arborea</i>
anac cana	<i>Anacharis canadensis</i>
andm glau	<i>Andromeda glaucophylla</i>

andp dive	<i>Andropogon divergens</i>
andp elli	<i>Andropogon elliottii</i>
andp gera	<i>Andropogon gerardi</i>
andp subt	<i>Andropogon subtenuis</i>
andp tene	<i>Andropogon tener</i>
andp tern	<i>Andropogon ternarius</i>
andp virg	<i>Andropogon virginicus</i>
andp ----	<i>Andropogon</i>
ante plan	<i>Antennaria plantaginifolia</i>
arab drum	<i>Arabis drummondii</i>
aral nudi	<i>Aralia nudicaulis</i>
aral race	<i>Aralia racemosa</i>
arcg lati	<i>Arctagrostis latifolia</i>
arcs alpi	<i>Arctostaphylos alpina</i>
arcs glan	<i>Arctostaphylos glandulosa</i>
arcs patu	<i>Arctostaphylos patula</i>
arcs pung	<i>Arctostaphylos pungens</i>
arcs stan	<i>Arctostaphylos stanfordiana</i>
arcs uvur	<i>Arctostaphylos uva-ursi</i>
arcs visc	<i>Arctostaphylos viscosa</i>
arcs ----	<i>Arctostaphylos</i>
arct cale	<i>Arctotheca calendula</i>
aren groe	<i>Arenaria groenlandica</i>
aris long	<i>Aristida longiseta</i>
aris stri	<i>Aristida stricta</i>
aris ----	<i>Aristida</i>
arni fulg	<i>Arnica fulgens</i>
aron arbu	<i>Aronia arbutifolia</i>
aron mela	<i>Aronia melanocarpa</i>
arte arbu	<i>Artemisia arbuscula</i>
arte arno	<i>Artemisia arbuscula nova</i>
arte arct	<i>Artemisia arctica</i>
arte cali	<i>Artemisia californica</i>
arte camp	<i>Artemisia campestris</i>
arte cana	<i>Artemisia cana</i>
arte fili	<i>Artemisia filifolia</i>
arte frig	<i>Artemisia frigida</i>
arte gnap	<i>Artemisia gnaphalodes</i> (see also arte vulg)
arte ludo	<i>Artemisia ludoviciana</i>
arte nova	<i>Artemisia nova</i>
arte peda	<i>Artemisia pedatifida</i>

arte spin	<i>Artemisia spinescens</i>
arte trid	<i>Artemisia tridentata</i>
arte trip	<i>Artemisia tripartita</i>
arte vulg	<i>Artemisia vulgaris</i> (see also arte gnap)
arte ----	<i>Artemisia</i>
aste chil	<i>Aster chilensis</i>
aste comm	<i>Aster commutatus</i>
aste foli	<i>Aster foliaceus</i>
aste pilo	<i>Aster pilosus</i>
aste tena	<i>Aster tenacetifolius</i>
aste ----	<i>Aster</i>
astr bisu	<i>Astragalus bisulcatus</i>
astr emor	<i>Astragalus emoryanus</i>
astr mise	<i>Astragalus miser</i>
astr patt	<i>Astragalus pattersonii</i>
astr recu	<i>Astragalus recurvus</i>
astr tetr	<i>Astragalus tetrapterus</i>
astr ----	<i>Astragalus</i>
atri cane	<i>Atriplex canescens</i>
atri conf	<i>Atriplex confertifolia</i>
atri coro	<i>Atriplex coronata</i>
atri eleg	<i>Atriplex elegans</i>
atri hali	<i>Atriplex halimoides</i>
atri holo	<i>Atriplex holocarpa</i>
atri lent	<i>Atriplex lentiformis</i>
atri line	<i>Atriplex linearis</i>
atri nutt	<i>Atriplex nuttallii</i>
atri poly	<i>Atriplex polycarpa</i>
atri rose	<i>Atriplex rosea</i>
atri semi	<i>Atriplex semibaccata</i>
atri volu	<i>Atriplex volutans</i>
aula turg	<i>Aulacomnium turgidum</i>
aven fatu	<i>Avena fatua</i>
aven sati	<i>Avena sativa</i>

bahi oppo	<i>Bahia oppositifolia</i>
bals sagg	<i>Balsamorhiza sagittata</i>
benz aest	<i>Benzoin aestivale</i> (see also lind benz)
berb nerv	<i>Berberis nervosa</i>
berb thun	<i>Berberis thunbergii</i>
berb trif	<i>Berberis trifoliata</i>

berc scan	<i>Berchemia scandens</i>
betu alba	<i>Betula alba</i>
betu alle	<i>Betula alleghaniensis</i>
betu glan	<i>Betula glandulosa</i>
betu lent	<i>Betula lenta</i>
betu lute	<i>Betula lutea</i>
betu mino	<i>Betula minor</i>
betu nana	<i>Betula nana</i>
betu nigr	<i>Betula nigra</i>
betu papy	<i>Betula papyrifera</i>
betu pend	<i>Betula pendula</i>
betu popu	<i>Betula populifolia</i>
betu pube	<i>Betula pubescens</i>
betu pulm	<i>Betula pulma</i>
betu pumi	<i>Betula pumila</i>
betu verr	<i>Betula verrucosa</i>
betu ----	<i>Betula</i>
boer tenu	<i>Boerhaavia tenuifolia</i>
bole edul	<i>Boletus edulis</i>
bout grac	<i>Bouteloua gracilis</i>
bout ----	<i>Bouteloua</i>
brac decu	<i>Bracharia decumbens</i>
bras kabe	<i>Brassica kabér</i>
brom briz	<i>Bromus brizaeformis</i>
brom moll	<i>Bromus mollis</i>
brom pump	<i>Bromus pumpellianus</i>
brom rigi	<i>Bromus rigidus</i>
brom tect	<i>Bromus tectorum</i>
brun cirr	<i>Brunnichia cirrhosa</i>
buch dact	<i>Buchloe dactyloides</i>
bume texa	<i>Bumelia texana</i>
cala cana	<i>Calamagrostis canadensis</i>
cala casc	<i>Calamagrostis canadensis</i> var. <i>scabra</i>
calc amer	<i>Callicarpa americana</i>

cale stra	<i>Calliergon stramineum</i>
calg schi	<i>Calliergonella schieberi</i>
cali erio	<i>Calliandra eriophylla</i>
call palu	<i>Calla palustris</i>
calm cana	<i>Calmagrostis canadensis</i>
calt lept	<i>Caltha leptosepala</i>
calu vulg	<i>Calluna vulgaris</i>
came micr	<i>Camelina microcarpa</i>
camp rotu	<i>Campanula rotundifolia</i>
cams radi	<i>Campsis radicans</i>
cant ciba	<i>Cantharellus cibarius</i>
care aqua	<i>Carex aquatilis</i>
care bige	<i>Carex bigelowii</i>
care brev	<i>Carex brevipes</i>
care cane	<i>Carex canescens</i>
care eben	<i>Carex ebenea</i>
care geop	<i>Carex geophila</i>
care heli	<i>Carex heliophora</i>
care lacu	<i>Carex lacustris</i>
care mult	<i>Carex multicaulis</i>
care scir	<i>Carex scirpoidea</i>
care stri	<i>Carex stricta</i>
care ----	<i>Carex</i>
carn giga	<i>Carnegiea gigantea</i>
carp betu	<i>Carpinus betulus</i>
cart tinc	<i>Carthamus tinctorius</i>
cary aqua	<i>Carya aquatica</i>
cary cord	<i>Carya cordiformis</i>
cary glab	<i>Carya glabra</i>
cary illi	<i>Carya illinoensis</i>
cary leid	<i>Carya leidodermus</i>
cary ovat	<i>Carya ovata</i> (see also <i>hico ovat</i>)
cary texa	<i>Carya texana</i>
cary tome	<i>Carya tomentosa</i>
cary ----	<i>Carya</i>

casa cham	Cassia chamaecrista (see also casa fasc)
casa fasc	Cassia fasciculata (see also casa cham)
casa mari	Cassia marilandica
casa nict	Cassia nictitans
casa roem	Cassia roemeriana
casa ----	Cassia
casi hypn	Cassiope hypnoides
casi tetr	Cassiope tetragona
casn dent	Castanea dentata
casn vulg	Castanea vulgaris
cata spec	Catalpa speciosa
cean amer	Ceanothus americanus
cean cord	Ceanothus cordulatus
cean cune	Ceanothus cuneatus
cean diva	Ceanothus divaricatus (see also cean leuc)
cean fend	Ceanothus fenderli
cean foli	Ceanothus foliosus
cean greg	Ceanothus greggii
cean inte	Ceanothus integerrimus
cean leuc	Ceanothus leucodermis (see also cean diva)
cean parv	Ceanothus parvifolius
cean pros	Ceanothus prostratus
cean velu	Ceanothus velutinus
cean ----	Ceanothus
celt laev	Celtis laevigata
celt occi	Celtis occidentalis
celt pall	Celtis pallida
celt reti	Celtis reticulata
cera arve	Cerastium arvense
cerc betu	Cercocarpus betuloides
cerc brev	Cercocarpus breviflorus
cerc ledi	Cercocarpus ledifolius
cerc mont	Cercocarpus montanus
cerd ----	Cercidium
cerp deme	Ceratophyllum demersum
cers cana	Cercis canadensis
cetr cucu	Cetraria cucullata
cetr isla	Cetraria islandica
cetr niva	Cetraria nivalis
chae caly	Chaemaedaphne calyculata

cham foli	<i>Chamaebatia foliolosa</i>
chan caly	<i>Chamaedaphne calyculata</i>
chap thyo	<i>Chamaecyparis thyoides</i>
char vulg	<i>Chara vulgaris</i>
chen albu	<i>Chenopodium album</i>
chlo cucu	<i>Chloris cucullata</i>
chlo gaya	<i>Chloris gayana</i>
chrs gram	<i>Chrysopsis graminifolia</i>
chry lanc	<i>Chrysothamnus lanceolatus</i>
chry naus	<i>Chrysothamnus nauseosus</i>
chry pulc	<i>Chrysothamnus pulcherrimus</i>
chry sten	<i>Chrysothamnus stenophyllus</i>
chry tere	<i>Chrysothamnus teretifolius</i>
chry visc	<i>Chrysothamnus viscidiflorus</i>
chry vise	<i>Chrysothamnus viscidiflorus serrulatus</i>
chry ----	<i>Chrysothamnus</i>
cicu ----	<i>Cicuta</i>
cirs hook	<i>Cirsium hookerianum</i>
cirs undu	<i>Cirsium undulatum</i>
citr limo	<i>Citrus limon</i>
clad alpe	<i>Cladonia alpestris</i>
clad grac	<i>Cladonia gracilllis</i>
clad miti	<i>Cladonia mitis</i>
clad rang	<i>Cladonia rangifera</i>
clad rani	<i>Cladonia rangiferina</i>
clad ----	<i>Cladonia</i>
clar lute	<i>Cladrastis lutea</i>
clav purp	<i>Claviceps purpurea</i>
clem drum	<i>Clematis drummondii</i>
clet alni	<i>Clethra alnifolia</i>
cole ramo	<i>Coleogyne ramosissima</i>
coll line	<i>Collomia linearis</i>
colu texe	<i>Colubrina texensis</i>

coma pall	<i>Comandra pallida</i>
comp pere	<i>Comptonia peregrina</i>
cond obtu	<i>Condalia obtusifolia</i>
conv arve	<i>Convolvulus arvensis</i>
corn alte	<i>Cornus alternifolia</i>
corn cana	<i>Cornus canadensis</i>
corn drum	<i>Cornus drummondii</i>
corn flor	<i>Cornus florida</i>
corn nutt	<i>Cornus nuttallii</i>
corn pani	<i>Cornus paniculata</i> (see also corn race)
corn race	<i>Cornus racemosa</i> (see also corn pani)
corn rugo	<i>Cornus rugosa</i>
corn stol	<i>Cornus stolonifera</i>
corn ----	<i>Cornus</i>
coro vari	<i>Coronilla varia</i>
cory amer	<i>Corylus americana</i>
cory avel	<i>Corylus avellana</i>
cory cali	<i>Corylus californica</i>
cory corn	<i>Corylus cornuta</i> (see also cory rost)
cory rost	<i>Corylus rostrata</i> (see also cory corn)
cory ----	<i>Corylus</i>
coto pyra	<i>Cotoneaster pyracantha</i>
covi trid	<i>Covillea tridentata</i>
cowa stan	<i>Cowania stansburiana</i>
crat crus	<i>Crataegus crus-galli</i>
crat mono	<i>Crataegus monogyna</i>
crat poli	<i>Crataegus polita</i>
crat ----	<i>Crataegus</i>
crot cory	<i>Croton corymbulosus</i>
crot mona	<i>Croton monanthogynus</i>
crot neom	<i>Croton neomexicanus</i>
crot ----	<i>Croton</i>
cryp japo	<i>Cryptomeria japonica</i>
cucu foet	<i>Cucurbita foetidissima</i>
cucu pepo	<i>Cucurbita pepo</i>
cusc exal	<i>Cuscuta exaltata</i>
cymo wats	<i>Cymopterus watsonii</i>

cyna vinc Cynanchum vincetoxicum
cyno dact Cynodon dactylon
cypc ---- Cyperaceae
cype rotu Cyperus rotundus
cyri race Cyrilla racemiflora

dact glom Dactylis glomerata
dale albi Dalea albiflora
daph meze Daphne mezereum
delp ande Delphinium andersonii
delp barb Delphinium barbeyi
delp glac Delphinium glaucescens
delp glam Delphinium glaucum
delp nels Delphinium nelsonii
delp occi Delphinium occidentale
delp ---- Delphinium

desc soph Descarainia sophia

desh caes Deschampsia caespitosa
desh flex Deschampsia flexuosa

desi soph Descurainia sophia

desm cool Desmanthus cooleyi
desm fall Desmanthus fallax

deso glut Desmodium glutinosum

diap lapp Diapensia lapponica

dicr beig Dicranum beigeri

dier loni Diervilla lonicera

dioc mult Dioclea multiflora

dios texa Diospyros texana
dios virg Diospyros virginiana

drab aure Draba aurea

echi crus	<i>Echinochloa crusgalli</i>
ecin crus	<i>Echinochloa crusgalli</i>
elae angu	<i>Elaeagnus angustifolia</i>
eleo smal	<i>Eleocharis smallii</i>
elym glau	<i>Elymus glauca</i>
empe eame	<i>Empetrum eamesii</i> ssp. <i>hermaphroditum</i>
empe nigr	<i>Empetrum nigrum</i>
ephe anti	<i>Ephedra antisiphilitica</i>
ephe neva	<i>Ephedra nevadensis</i>
ephe torr	<i>Ephedra torreyana</i>
ephe trif	<i>Ephedra trifurca</i>
ephe viri	<i>Ephedra viridis</i>
ephe ----	<i>Ephedra</i>
epil angu	<i>Epilobium angustifolium</i>
epil lati	<i>Epilobium latifolium</i>
equi ----	<i>Equisetum</i>
erag ----	<i>Eragrostis</i>
ergo race	<i>Ergonum racemosum</i>
eria ----	<i>Eriastrum</i>
erid cali	<i>Eriodictyon californicum</i>
erie ----	<i>Erigeron</i>
erig hera	<i>Eriogonum heracleoides</i>
erig race	<i>Eriogonum racemosum</i>
erig wrig	<i>Eriogonum wrightii</i>
erig ----	<i>Eriogonum</i>
erip vagi	<i>Eriophorum vaginatum</i>
erip ----	<i>Eriophorum</i>
erod cicu	<i>Erodium cicutarium</i>
erod mosc	<i>Erodium moschatum</i>
erys aspe	<i>Erysimum asperum</i>
eryt amer	<i>Erythronium americanum</i>
eryt gran	<i>Erythronium grandiflorum</i>
euca obli	<i>Eucalyptus obliqua</i>

euon amer Euonymus americana
euph cict Euphorbia cictyesperma
euph coro Euphorbia corollata
euph fend Euphorbia fendleri
euph pros Euphorbia prostrata
euph ---- Euphorbia

euro lana Eurotia lanata

evol nutt Evolvulus nuttallianus

fagu gran Fagus grandifolia
fagu sylv Fagus sylvatica
fagu ---- Fagus

fero wisl Ferocactus wislizenii

fest alta Festuca altaica
fest ariz Festuca arizonia
fest idah Festuca idahoensis
fest octo Festuca octoflora
fest ---- Festuca

flou cern Flourensia cernua

fome ---- Fomes

fore neom Forestiera neomexicana

frag amer Fragaria americana

fras ---- Franseria

frax amer Fraxinus americana
frax exce Fraxinus excelsior
frax lati Fraxinus latifolia
frax quad Fraxinus quadrangulata
frax ---- Fraxinus

fuma offi Fumaria officinalis

gali bore Galium boreale
gaul proc Gaultheria procumbens

gayl bacc Gaylussacia baccata
gels semp Gelsemium sempervirens
gera rich Geranium richardsoni
gera ---- Geranium
geum peck Geum peckii
geum trif Geum triflorum
geum ---- Geum
gili mult Gilia multiflora
gled tria gleditsia triacanthos
gram ---- gramineae
gray spin Grayia spinosa
guai coul Guaiacum coulteri
guar cocc Guara coccinea
guti micr Gutierrezia microcephala
guti saro Gutierrezia sarocephala
guti spha Gutierrezia sphaerocephala
guti texa Gutierrezia texana
gyro escu Gyromitra esculenta

halo glom Halogeton glomeratus
hama vern Hamamelis vernalis
hama virg Hamamelis virginiana
hedy bore Hedysarum boreale
hele hoop Helenium hoopesii
heli quin Helianthella quinquenervis
heln angu Helianthus angustifolius
heln annu Helianthus annuus
heln hirs Helianthus hirsutus
helo heli Heliopsis helianthoides
heve ---- Hevea

hevl escu *Hevella esculenta*
hico ovat *Hicoria ovata*
hila bela *Hilaria belangeri*
hoff brac *Hoffmannseggia brachycarpa*
hord lepo *Hordeum leporinum*
hous caer *Houstonia caerulea var. faxonorum*
hydr arbo *Hydrangea arborescens*
hyme odor *Hymenoxys odorata*
hyme rich *Hymenoxys richardsonii var. floribunda*
hype macu *Hypericum maculatum*
hyprr perf *Hypericum perforatum*

ilex cori *Ilex coriacea*
ilex deci *Ilex decidua*
ilex glab *Ilex glabra*
ilex opac *Ilex opaca*
ilex vert *Ilex verticillata*
ilex vomi *Ilex vomitoria*

impa cape *Impatiens capensis*
ipom ---- *Ipomoea*
iva axil *Iva axillaris*

jacq tamn *Jacquemontia tamnifolia*
jugl cine *Juglans cinerea*
jugl nigr *Juglans nigra*

junc comm *Juncus communis*
junc trif *Juncus trifidus*

juni hori *Juniperus horizontalis*
juni knig *Juniperus knightii*
juni occi *Juniperus occidentalis*

juni pinc Juniperus pinchotii
juni scop Juniperus scopulorum
juni utah Juniperus utahensis
juni virg Juniperus virginiana

kalm angu Kalmia angustifolia
kalm lati Kalmia latifolia
kalm poli Kalmia polifolia

koch amer Kochia americana
koch chil Kochia childsii
koch scop Kochia scoparia
koch vest Kochia vestita

kram secu Krameria secundiflora

kunz trid Kunzia tridentata (see also purs trid)

lact cana Lactuca canadensis
lact ---- Lactuca

lamn minr Lamna minor

lari lari Larix laricina
lari lept Larix leptolepis
lari occi Larix occidentalis

lath ---- Lathyrsus

laty ochr Lathyrus ochroleucus
laty ---- Lathyrus

ledu groe Ledum groenlandicum
ledu palu Ledum palustre v. decumbens

lept mult Leptoptaenia multifida

lesp stip Lespediza stipulacea

leuc retu Leucaena retusa

leuo minu Leucophyllum minus

leut edit Leucothoe editorum

lewi pygm Lewisia pygmaea

liat punc	Liatrus punctata
libo decu	Libocedrus decurrens
ligs port	Ligusticum porteri
ligu obtu	Ligustrum obtusifolium
lind benz	Lindera benzoin (see also benz aest)
linm usit	Linum usitatissimum
liqu styr	Liquidambar styraciflua
liri tuli	Liriodendron tulipifera
lois proc	Loiseleuria procumbens
loli rigi	Lolium rigidum
loni albi	Lonicera albinura
loni cana	Lonicera canadensis
loni invo	Lonicera involucrata
loni japo	Lonicera japonica
loni morr	Lonicera morrowii
loni vill	Lonicera villosa
loni xylo	Lonicera xylosteum
lotu wrig	Lotus wrightii
lotu ----	Lotus
lupi arct	Lupinus arcticus
lupi caud	Lupinus caudatus
lupi seri	Lupinus sericeus
lupi ----	Lupinus
luzu spic	Luzula spicata
lyco anno	Lycopodium annotinum var. pungens
lyco sela	Lycopodium selago var. appressum
lyon luci	Lyonia lucida
mac1 pom1	Maclura pomifera
magn acum	Magnolia acuminata
magn macr	Magnolia macrophylla
magn virg	Magnolia virginiana

maho repe	<i>Mahonia repens</i>
maia cana	<i>Maianthemum canadense</i>
mani escu	<i>Manihot esculenta</i>
medi sati	<i>Medicago sativa</i>
meli offi	<i>Melilotus officinalis</i>
meli ----	<i>Melilotus</i>
ment pipe	<i>Mentha piperita</i>
menz pilo	<i>Menziesia pilosa</i>
mert cili	<i>Mertensia ciliata</i>
mimo frag	<i>Mimosa fragrans</i>
mira line	<i>Mirabilis linearis</i>
mona odor	<i>Monardella odoratissima</i>
morc angu	<i>Morchella angusticeps</i>
morc coni	<i>Morchella conica</i>
morc cras	<i>Morchella crassipes</i>
morc deli	<i>Morchella deliciosa</i>
morc escu	<i>Morchella esculenta</i>
moru alba	<i>Morus alba</i>
moru micr	<i>Morus microphylla</i>
moru rubr	<i>Morus rubra</i>
muhl mont	<i>Muhlenbergia montana</i>
muhl ----	<i>Muhlenbergia</i>
musa acum	<i>Musa acuminata</i>
myos alpe	<i>Myosotis alpestris</i>
myri aspl	<i>Myrica asplenifolia</i>
myri ceri	<i>Myrica cerifera</i>
myri gale	<i>Myrica gale</i>
myri pens	<i>Myrica pensylvanica</i>
myro exal	<i>Myriophyllum exalbescens</i>

nemo mucr *Nemopanthus mucronata*
neph arct *Nephroma arcticum*
noli macr *Nolina macrocarpa*
noli micr *Nolina microcarpa*
noli texa *Nolina texana*
nuph vari *Nuphar variegatum*
nuph ---- *Nuphar*
nymp tube *Nymphaea tuberosa*
nyss sylv *Nyssa sylvatica*

oeno ---- *Oenothera*
opun enge *Opuntia engelmannii*
opun fulg *Opuntia fulgida*
opun poly *Opuntia polycantha*
opun spin *Opuntia spinosior*
opun ---- *Opuntia*
orth lute *Orthocarpus luteus*
oryz aspe *Oryzopsis asperfolia*
oryz hyme *Oryzopsis hymenoides*
osmo clay *Osmorrhiza claytoni*
osmo occi *Osmorrhiza occidentalis*
ostr virg *Ostrya virginiana*
oxyd arbo *Oxydendrum arboreum*
oxyt ---- *Oxytropis*

pani ---- *Panicum*
park ---- *Parkinsonia*
parm ---- *Parmelia*
part quin *Parthenocissus quinquefolia*
part vita *Parthenocissus vitacea*
pasp ---- *Paspalum*

pelg ----	Peltigera
pens leon	Penstemon leonardi
pens whip	Penstemon whippleanus
pere nana	Perezia nana
peri gair	Perideridia gairdneri
pers borb	Persea borbonia
pers ----	Persea
phal arun	Phalaris arundinacea
phas ----	Phaseolus
phle prat	Phleum pratense
phyl caer	Phyllodoce caerulea
phys malv	Physocarpus malvaceus
phys pube	Physocarpus pubescens
phyt amer	Phytolacca americana
pice abie	Picea abies
pice enge	Picea engelmanni
pice glau	Picea glauca
pice mari	Picea mariana
pice obov	Picea obovata
pice rube	Picea rubens
pice sitc	Picea sitchensis
pice ----	Picea
pins albi	Pinus albicaulis
pins bank	Pinus banksiana
pins cemb	Pinus cembroides
pins clau	Pinus clausa
pins cont	Pinus contorta
pins ctla	Pinus contorta latifolia
pins echi	Pinus echinata
pins elli	Pinus elliottii
pins lamb	Pinus lambertiana
pins mont	Pinus monticola
pins nigr	Pinus nigra
pins nrca	Pinus nigra var. calabrica
pins nrma	Pinus nigra var. maritima
pins palu	Pinus palustris
pins pond	Pinus ponderosa
pins radi	Pinus radiata
pins resi	Pinus resinosa
pins rigi	Pinus rigida

pins sibe	<i>Pinus siberica</i>
pins silv	<i>Pinus silvestris</i>
pins stro	<i>Pinus strobus</i>
pins sylv	<i>Pinus sylvestris</i>
pins taed	<i>Pinus taeda</i>
pins virg	<i>Pinus virginiana</i>
pins ----	<i>Pinus</i>
plan majo	<i>Plantago major</i>
plan purs	<i>Plantago purshii</i>
plat occi	<i>Platanus occidentalis</i>
pleu schr	<i>Pleurozium schreberi</i>
poa cusci	<i>Poa cusickii</i>
poa fend	<i>Poa fendleriana</i>
poa fern	<i>Poa fernaldiana</i>
poa patt	<i>Poa pattersoni</i>
poa rupi	<i>Poa rupicola</i>
poa sand	<i>Poa sandbergii</i>
poa ----	<i>Poa</i>
poly alas	<i>Polygonum alaskanum</i>
poly avic	<i>Polygonum aviculare</i>
poly conv	<i>Polygonum convolvulus</i>
poly fago	<i>Polygonum fagopyrum</i>
poly pens	<i>Polygonum pensylvanicum</i>
poly vivi	<i>Polygonum viviparum</i>
poys muni	<i>Polystichum munitum</i>
poyt juni	<i>Polytrichum juniperinum</i> var. <i>alpestre</i>
poyt pili	<i>Polytrichum piliferum</i>
popu delt	<i>Populus deltoides</i>
popu gran	<i>Populus grandidentata</i>
popu trem	<i>Populus tremula</i>
popu treu	<i>Populus tremuloides</i>
popu tric	<i>Populus trichocarpa</i>
popu ----	<i>Populus</i>
pota ampl	<i>Potamogeton amplifolius</i>
pota epih	<i>Potamogeton epihydrus</i>
pota pect	<i>Potamogeton pectinatus</i>
pota rich	<i>Potamogeton richardsonii</i>
pota robi	<i>Potamogeton robinsii</i>
pota zost	<i>Potamogeton zosteriformis</i>
pota ----	<i>Potamogeton</i>
pote dive	<i>Potentilla diversifolia</i>
pote grac	<i>Potentilla gracilis</i>

pote trid	<i>Potentilla tridentata</i>
pote ----	<i>Potentilla</i>
pros chil	<i>Prosopis chilensis</i>
pros juli	<i>Prosopis juliflora</i>
pros velu	<i>Prosopis velutina</i>
pros ----	<i>Prosopis</i>
prun amer	<i>Prunus americana</i>
prun ande	<i>Prunus andersonii</i>
prun demi	<i>Prunus demissa</i> (see also prun virg)
prun emar	<i>Prunus emarginata</i>
prun fasc	<i>Prunus fasciculata</i>
prun ilic	<i>Prunus ilicifolia</i>
prun mari	<i>Prunus maritima</i>
prun mela	<i>Prunus melanocarpa</i> (see also prun vrme)
prun minu	<i>Prunus minutiflora</i>
prun pens	<i>Prunus pensylvanica</i>
prun sero	<i>Prunus serotina</i>
prun subc	<i>Prunus subcordata</i>
prun umbe	<i>Prunus umbellata</i>
prun virg	<i>Prunus virginiana</i> (see also prun demi)
prun vrme	<i>Prunus virginiana melanocarpa</i> (see also prun mela)
prun ----	<i>Prunus</i>
pscym mont	<i>Pseudocymopterus montanus</i>
pseu menz	<i>Pseudotsuga menziesii</i>
pseu taxi	<i>Pseudotsuga taxifolia</i>
pseu tagl	<i>Pseudotsuga taxifolia glauca</i>
psor tenu	<i>Psoralea tenuiflora</i>
pter aqui	<i>Pteridium aquilinum</i>
puer loba	<i>Pueraria lobata</i>
purs trid	<i>Purshia tridentata</i> (see also kunz trid)
purs ----	<i>Purshia</i>
pyrl pube	<i>Pyrularia pubera</i>
pyru angu	<i>Pyrus angustifolia</i>
pyru comm	<i>Pyrus communis</i>
pyru coro	<i>Pyrus coronaria</i>
pyru malu	<i>Pyrus malus</i>
pyru ----	<i>Pyrus</i>

quer acut	<i>Quercus acutissima</i>
quer agri	<i>Quercus agrifolia</i>
quer alba	<i>Quercus alba</i>
quer bico	<i>Quercus bicolor</i>
quer bore	<i>Quercus borealis</i>
quer bref	<i>Quercus brevifolia</i> (see also quer cine)
quer brev	<i>Quercus brevirostra</i>
quer cali	<i>Quercus californica</i>
quer chry	<i>Quercus chrysophylla</i>
quer cine	<i>Quercus cinnamomea</i> (see also quer bref)
quer cocc	<i>Quercus coccinea</i>
quer doug	<i>Quercus douglasii</i>
quer dumo	<i>Quercus dumosa</i>
quer dura	<i>Quercus durata</i>
quer elli	<i>Quercus ellipsoidalis</i>
quer emor	<i>Quercus emoryi</i>
quer falc	<i>Quercus falcata</i> (see also quer rubr)
quer gamb	<i>Quercus gambelii</i>
quer garr	<i>Quercus garryana</i>
quer harv	<i>Quercus harvardii</i>
quer ilic	<i>Quercus ilicifolia</i>
quer inca	<i>Quercus incana</i>
quer kell	<i>Quercus kelloggii</i>
quer lyra	<i>Quercus lyrata</i>
quer macr	<i>Quercus macrocarpa</i>
quer mari	<i>Quercus marilandica</i>
quer mini	<i>Quercus minima</i>
quer mino	<i>Quercus minima</i> (see also quer stel)
quer mont	<i>Quercus montana</i>
quer nigr	<i>Quercus nigra</i>
quer nutt	<i>Quercus nuttallii</i>
quer obtu	<i>Quercus obtusa</i>
quer palu	<i>Quercus palustris</i>
quer petr	<i>Quercus petraea</i>
quer phel	<i>Quercus phellos</i>
quer phil	<i>Quercus philtas</i>
quer prin	<i>Quercus prinoides</i>
quer prnu	<i>Quercus prinus</i>
quer robu	<i>Quercus robur</i>
quer rubr	<i>Quercus rubra</i> (see also quer falc)
quer shum	<i>Quercus shumardii</i>
quer stel	<i>Quercus stellata</i> (see also quer mino)
quer turb	<i>Quercus turbinella</i>
quer velu	<i>Quercus velutina</i>
quer virg	<i>Quercus virginiana</i>
quer wisl	<i>Quercus wislizenii</i>
quer ----	<i>Quercus</i>

ranu ----	Ranunculus
rham cali	Rhamnus californica
rham caro	Rhamnus caroliniana
rham fran	Rhamnus frangula
rham purs	Rhamnus purshiana
rhod lapp	Rhododendron lapponicum
rhod maxi	Rhododendron maximum
rhod rose	Rhododendron roseum
rhod ----	Rhododendron
rhoo cana	Rhododendron canadense
rhus arom	Rhus aromatica
rhus cana	Rhus canadensis
rhus copa	Rhus copallina
rhus dive	Rhus diversiloba
rhus glab	Rhus glabra
rhus hirt	Rhus hirta (see also rhus typh)
rhus micr	Rhus microphylla
rhus radi	Rhus radicans
rhus tril	Rhus trilobata (see also schm tril)
rhus typh	Rhus typhina (see also rhus hirt)
rhus vire	Rhus virens
rhus ----	Rhus
robi neom	Robinia neomexicana
robi pseu	Robinia pseudoacacia
rosa acic	Rosa acicularis
rosa egl	Rosa eglanteria
rosa fend	Rosa fendleri (see also rosa wood)
rosa humi	Rosa humilis
rosa maco	Rosa macounii
rosa maja	Rosa majalis
rosa mult	Rosa multiflora
rosa palu	Rosa palustris
rosa rugo	Rosa rugosa
rosa setg	Rosa setigera
rosa setr	Rosa setigeria
rosa spal	Rosa spaldingii
rosa spin	Rosa spinosissima
rosa wood	Rosa woodsii (see also rosa fend)
rosa woul	Rosa woodsii ultralmontana
rosa ----	Rosa
rubu alle	Rubus allegheniensis
rubu arct	Rubus arcticus
rubu bail	Rubus baileyanus
rubu cham	Rubus chamaemorus
rubu idae	Rubus idaeus

rubu occi Rubus occidentalis
rubu parv Rubus parviflorus
rubu saxa Rubus saxatilis
rubu spec Rubus spectabilis
rubu viti Rubus vitifolius
rubu ---- Rubus

rudb hirt Rudbeckia hirta

rume acet Rumex acetosella
rume cris Rumex crispus

saba ---- Sabal

sagi cune Sagittaria cuneata
sagi rigi Sagittaria rigida

sali alax Salix alaxensis
sali amyg Salix amygdaloïdes
sali arbu Salix arbusculoïdes
sali arct Salix arctica
sali bebb Salix bebbiana
sali capr Salix caprea
sali glau Salix glauca
sali humi Salix humilis
sali inte Salix interior
sali lasi Salix lasiolepis
sali lute Salix lutea
sali nigr Salix nigra
sali phyl Salix phylicifolia
sali plan Salix planifolia
sali scou Salix scouleriana
sali uvur Salix uva-ursi
sali ---- Salix

sals kali Salsola kali

salv mell Salvia mellifera
salv refl Salvia reflexa

samb call Sambucus callicarpa
samb cana Sambucus canadensis
samb glau Sambucus glauca
samb pube Sambucus pubens
samb ---- Sambucus

sang cana Sanguisorba canadensis

sapi sebi Sapium sebiferum

sarc verm	<i>Sarcobatus vermiculatus</i>
sass albi	<i>Sassafras albidum</i>
schm tril	<i>Schmaltzia trilobata</i> (see also <i>rhus tril</i>)
scir cesp	<i>Scirpus cespitosus</i>
scol fest	<i>Scolochloa festucacea</i>
sene jaco	<i>Senecio jacobaea</i>
sene long	<i>Senecio longilobus</i>
sene neom	<i>Senecio neomexicanus</i>
sene ridd	<i>Senecio riddellii</i>
sene tria	<i>Senecio triangularis</i>
sene ----	<i>Senecio</i>
sequ giga	<i>Sequoiadendron giganteum</i>
seta fabr	<i>Setaria faberi</i>
seta glau	<i>Setaria glauca</i>
seta lute	<i>Setaria lutescens</i>
seta viri	<i>Setaria viridis</i>
shep cana	<i>Shepherdia canadensis</i>
sibb proc	<i>Sibbaldia procumbens</i>
sile acau	<i>Silene acaulis</i> var. <i>exscapa</i>
sile doug	<i>Silene douglasii</i>
sile noct	<i>Silene noctiflora</i>
simm chin	<i>Simmondsia chinensis</i>
sita hyst	<i>Sitanion hystrix</i>
smia trif	<i>Smilacina trifolia</i>
smil bona	<i>Smilax bona-nox</i>
smil glau	<i>Smilax glauca</i>
smil rotu	<i>Smilax rotundifolia</i>
smil smal	<i>Smilax smallii</i>
smil tamm	<i>Smilax tamoides</i>
smil tamn	<i>Smilax tamnoides</i>
smil ----	<i>Smilax</i>
sola ----	<i>Solanum</i>
soli cutl	<i>Solidago cutleri</i>
soli macr	<i>Solidago macrophylla</i> var. <i>thyrsoides</i>
soli nemo	<i>Solidago nemoralis</i>

sorb amer	<i>Sorbus americana</i>
sorb aucu	<i>Sorbus aucuparia</i>
sorg hale	<i>Sorghum halepense</i>
spar eury	<i>Sparganium eurycarpum</i>
spar fluc	<i>Sparganium fluctuans</i>
spat ----	<i>Spartina</i>
spha cocc	<i>Sphaeralcea coccinea</i>
spha gros	<i>Sphaeralcea grossulariaefolia</i>
sphg fusc	<i>Sphagnum fuscum</i>
sphg girg	<i>Sphagnum girgensohnii</i>
spir alba	<i>Spiraea alba</i>
spir lati	<i>Spiraea latifolia</i>
spir ----	<i>Spiraea</i>
spor cryp	<i>Sporobolus cryptandrus</i>
spor curt	<i>Sporobolus curtissii</i>
spor ----	<i>Sporobolus</i>
stan pinn	<i>Stanleya pinnata</i>
ster pasc	<i>Stereocaulon paschale</i>
ster ----	<i>Stereocaulon</i>
stip colu	<i>Stipa columbiana</i>
stip coma	<i>Stipa comata</i>
stip pine	<i>Stipa pinetorum</i>
symp occi	<i>Symporicarpos occidentalis</i>
symp orbi	<i>Symporicarpos orbiculatus</i>
symp oreo	<i>Symporicarpos oreophilus</i>
symp rotu	<i>Symporicarpos rotundifolius</i>
symp vacc	<i>Symporicarpos vaccinioides</i>
sypl tinc	<i>Symplocos tinctoria</i>
syri vulg	<i>Syringa vulgaris</i>

tara offi	<i>Taraxacum officinale</i>
tara ----	<i>Taraxacum</i>
taxu bacc	<i>Taxus baccata</i>
taxu cana	<i>Taxus canadensis</i>

tetr cane	Tetradymia canescens
tetr glab	Tetradymia glabrata
thel ----	Thelesperma
thla alpe	Thlaspi alpestre
thuj occi	Thuja occidentalis
thuj plic	Thuja plicata
tide lanu	Tidestromia lanuginosa
tili amer	Tilia americana
tili cord	Tilia cordata
trad occi	Tradescantia occidentalis
trad suba	Tradescantia subaspera
trag dubi	Tragopogon dubius
trag ----	Tragopogon
trib terr	Tribulus terrestris
trif hybr	Trifolium hybridum
trif parr	Trifolium parryi
trif prat	Trifolium pratense
trif subt	Trifolium subterraneum
trif ----	Trifolium
trig mari	Triglochin maritima
tris spic	Trisetum spicatum var. pilosiglume
tsug cana	Tsuga canadensis
tsug hete	Tsuga heterophylla
typh augu	Typha augustifolia

ulmu alat	Ulmus alata
ulmu amer	Ulmus americana
ulmu glab	Ulmus glabra
umbi ----	Umbilicaria
umbr cali	Umbrellularia californica
ungn spec	Ungnadia speciosa
unio lati	Uniola latifolia

vacc angu	Vaccinium angustifolium
vacc arbo	Vaccinium arboreum
vacc caes	Vaccinium caespitosum
vacc cras	Vaccinium crassifolium
vacc elli	Vaccinium elliottii
vacc idae	Vaccinium idaea
vacc memb	Vaccinium membranaceum
vacc myrt	Vaccinium myrtilloides (see also vacc myti)
vacc myti	Vaccinium myrtillus (see also vacc myrt)
vacc oval	Vaccinium ovalifolium
vacc oxyc	Vaccinium oxycoccus
vacc parv	Vaccinium parvifolium
vacc scop	Vaccinium scoparium
vacc stam	Vaccinium stamineum
vacc ulig	Vaccinium uliginosum
vacc ulal	Vaccinium uliginosum var. alpinum
vacc vaci	Vaccinium vacillans
vacc viti	Vaccinium vitis-idaea
vacc vitm	Vaccinium vitis-idaea var. minus
vacc ----	Vaccinium
vale occi	Valeriana occidentalis
vale sitc	Valeriana sitchensis
vall amer	Vallisneria americana
vera cali	Veratrum californicum
vera viri	Veratrum viride
verb ence	Verbesina encelioides
vero worm	Veronica wormskjoldii
verp bohe	Verpa bohemica
vibu alni	Viburnum alnifolium
vibu cass	Viburnum cassinoides
vibu dent	Viburnum dentatum
vibu lent	Viburnum lentago
vibu moll	Viburnum molle
vibu opul	Viburnum opulus
vibu prun	Viburnum prunifolium
vibu rufi	Viburnum rufidulum
vici pulc	Vicia pulchella
viti aest	Vitis aestivalis
viti bico	Vitis bicolor
viti cord	Vitis cordifolia (see also viti vulp)
viti ripa	Vitis riparia
viti rotu	Vitis rotundifolia
viti vulp	Vitis vulpina (see also viti cord)
viti ----	Vitis

xant pens Xanthium pensylvanicum

xanx frut Xanthoxylum fruticosum

yucc elat Yucca elata
yucc glau Yucca glauca
yucc moha Yucca mohavensis
yucc reve Yucca reverchonii
yucc thom Yucca thompsoniana
yucc trec Yucca treculeana
yucc ---- Yucca

zea mays Zea mays

ziza equa Zizania aquatica

zyga eleg Zygadenus elegans

JULIAN DAY: MONTH AND DAY EQUIVALENTS*

Day	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Day
1	001	032	060	091	121	152	182	213	244	274	305	335	1
2	002	033	061	092	122	153	183	214	245	275	306	336	2
3	003	034	062	093	123	154	184	215	246	276	307	337	3
4	004	035	063	094	124	155	185	216	247	277	308	338	4
5	005	036	064	095	125	156	186	217	248	278	309	339	5
6	006	037	065	096	126	157	187	218	249	279	310	340	6
7	007	038	066	097	127	158	188	219	250	280	311	341	7
8	008	039	067	098	128	159	189	220	251	281	312	342	8
9	009	040	068	099	129	160	190	221	252	282	313	343	9
10	010	041	069	100	130	161	191	222	253	283	314	344	10
11	011	042	070	101	131	162	192	223	254	284	315	345	11
12	012	043	071	102	132	163	193	224	255	285	316	346	12
13	013	044	072	103	133	164	194	225	256	286	317	347	13
14	014	045	073	104	134	165	195	226	257	287	318	348	14
15	015	046	074	105	135	166	196	227	258	288	319	349	15
16	016	047	075	106	136	167	197	228	259	289	320	350	16
17	017	048	076	107	137	168	198	229	260	290	321	351	17
18	018	049	077	108	138	169	199	230	261	291	322	352	18
19	019	050	078	109	139	170	200	231	262	292	323	353	19
20	020	051	079	110	140	171	201	232	263	293	324	354	20
21	021	052	080	111	141	172	202	233	264	294	325	355	21
22	022	053	081	112	142	173	203	234	265	295	326	356	22
23	023	054	082	113	143	174	204	235	266	296	327	357	23
24	024	055	083	114	144	175	205	236	267	297	328	358	24
25	025	056	084	115	145	176	206	237	268	298	329	359	25
26	026	057	085	116	146	177	207	238	269	299	330	360	26
27	027	058	086	117	147	178	208	239	270	300	331	361	27
28	028	059	087	118	148	179	209	240	271	301	332	362	28
29	029	[060]	088	119	149	180	210	241	272	302	333	363	29
30	030		089	120	150	181	211	242	273	303	334	364	30
31	031		090		151		212	243		304		365	31

* For leap year, February 29 = JDAY 60. Add 1 to all subsequent JDAYS.

