INTRODUCTION

TO PART III

PHYSIOLOGY AND METABOLISM OF WILD RUMINANTS

Physiology is an often-overlooked aspect of the ecology of an individual or a species. Yet, no behavioral act, no maintenance activity, or no productive process can occur without basic physiological processes.

The first two PARTS of this book have included descriptions of physical and chemical characteristics (PART I) and behavior (PART II). Both contained factual information based on observations on many different species. PART III is directed more toward the functional characteristics of various systems, emphasizing the processes rather than the products of life functions.

Understanding of the functions and metabolic processes described in the next five CHAPTERS is very important for an understanding of productivity. Energy metabolism, possible only with a supply of digestible nutrients, is a fundamental characteristic of life itself, as gastrointestinal functions convert range forage energy into chemical forms that can be metabolized for maintenance and production.

Various systems of the body perform specific functions; these are discussed in CHAPTER 6. SYSTEMS PHYSIOLOGY. Metabolic processes, including ENERGY METABOLISM (CHAPTER 7), PROTEIN METABOLISM (CHAPTER 8), and MINERAL, WATER, AND VITAMIN METABOLISM (CHAPTER 9) support the functions of the systems, providing the nutrients and enzymes necessary for tissue maintenance and production. Deviations from normal metabolic functions are discussed in CHAPTER 10: FACTORS AFFECTING THE PHYSIOLOGY AND METABOLISM OF WILD RUMINANTS.

Some of the topics in these CHAPTERS are discussed in only a general way because there are no specific data available for any species of wild ruminants. In some cases, domestic ruminants have been studied intensively and the basic principles applied to wild ruminants.

There is an emphasis, whenever possible, on mathematical expressions of basic measurements in order to relate these physiological characteristics to other ecological functions. The use of mathematics to evaluate interrelationships increases as more functions of ecological importance are evaluated together. <u>Ecological accounting</u> is very important, and physiological processes, especially metabolism, are very well suited for mathematical expressions and ecological accounting.

The necessary reference information, including a glossary of symbols used, glossary of code names for the serials, and list of publishers is given at the end of each chapter. A Julian Day Calender is provided at the ends of those chapters which include time-related calculations.

The lists of REFERENCES, PART III, that follow provide essential bibliographic information for books and articles containing general information on several biological characteristics. These general references will be helpful in many of the UNITS that follow, but they are not listed again after each UNIT as the UNIT lists are limited to more specific articles of direct application to the material discussed in each UNIT.

REFERENCES, PART III

PHYSIOLOGY AND METABOLISM OF WILD RUMINANTS

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS--- YEAR

aubo	repu	nyny	1023		bioenergetics and growth	brody.s	1945
aubo	mhbc	nyny	895		a text of general physiolo	mitchell,ph	1956
aubo	saco	phpa	1180		textbook of mediacl physio	guyton, ac	1961
aubo	moco	slmo	547		textbook of physiology	<pre>tuttle,ww; schotte</pre>	1961
aubo	jwis	nyny	468		textbook of comparat endoc	gorbman,a bern,ha	1962
aubo	saco	phpa	535		body cell mass, suppor env	<pre>moore,fd; oleson,/</pre>	1963
edbo	mhbc	nyny	1427		hawk's physiological chemi	oser,b1,ed	1965
edbo	saco	phpa	1308		clinical diagn, lab method	davidsohn,i; henry	1969
edbo	orpr	nute	522		energy metabol, farm anima	blaxter,kl; kielan	1969
edbo	lefe	phpa	402		animal growth and nutritio	hafez,ese;dyer,ia	1969
edbo	acpr	nyny	439		clinical biochem, dom anim	kaenko jj; corneli	1970
edbo	coup	itny	1463		duke's physiol of dom anim	swenson,mj,ed	1970
edbo	acpr	nyny	352		clin bloch of dom anim, v2	kaenko,jj; corneli	1971
edbo	lefe	phpa	573		textbook of veterin physio	breazile,je,ed	1971
aubo	lmep	11ca	648		review of physiolog chemis	harper,ha	1977
aubo	rokp	loen	597	cerv	deer of g. britain, irelan	whitehead,gk	1964
_				_			
aubo	huho	nyny	426	od	deer, antelope of america	caton, jd	1877
edbo	stac	hapa	668	od	deer of north america	taylor,wp	1956
aubo	stac	hapa	128	od	if deer are to survive	dasmann,w	1971
aubo	vipr	nyny	194	od	deer of the world	whitehead,gk	1972
			107			. .	
aubo	omee	eail	107	odv1	the white-tailed deer	madson, j	1961
edbo	nnrg	conh	256	odvi	the white-tal deer, new ha	siegler, hr	1968
		L	F(7	. 11		1	1050
aubo	ucap	beca	201	oane	a nerd of mule deer	linsdale, jm; tomic	1923
h -		1	01 E	1	hand of models have belowed an	Jamling 66	1027
aubo	oxup	hone	306	ceel	nera or red deer, benavior	dariing, ii	193/
aubo	stat	napa	105	ceel	the all	murie,oj	1909
aubo	wiwe	beec	200	ceel		mauson, j	1071
aubo	ucap	Deca	209	ceel	LUTE ETK	mecurrougn, ar	19/1
aubo	utor	toor	280	0101	north amoridan moore	notorgon ml	1055
aubo	acop	2000	20V	arar	north american moose	pererson, rr	1977

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TYPE PUBL CITY PGES ANIM KEY WORDS-----AUTHORS/EDITORS--- YEAR aubo macm nyny 300 rata bar-gr car of north canada pike,w 1892 aubo ukap laka rata bar-ground carib, keewatin harper,f 1955 aubo qupr oton 339 rata migratory, barren-ground c kelsall, jp 1968 anam the pronghorn antelope 1948 aubo stac hapa 238 einarsen,as utop toon 957 bibi the north american buffalo roe, fg 1951 aubo aubo ther nyny 242 bibi the buffalo haines,f 1970 aubo aakn nyny 339 bibi time of the buffalo 1972 mchugh, t aubo swap atoh 374 bibi the buffalo book, saga ani dary d 1974 aubo uchp chil 383 ov-- mt sheep, behavior, evolut geist, v 1971 aubo coup itny 248 ov-- mt sheep, man, norther wil geist, v 1975 aubo usgp wadc 242 ovca the bighorn of death valley welles, re; welles 1961 aubo qupr oton 166 obmo muskoxen in canada tener, js 1965 aubo dalt laen 271 dada fal de: histor, distr, bio chapman,d; chapman 1975 doup nyny 318 many americ anim; popular guide stone,w; cram,we aubo 1902 aubo cscs nyny 347 many our big game huntington.d 1904 aubo cscs nyny 1267 many life hist northern animals seton, et 1909 many wildlife in alaska, ecolog leopold, as; darlin 1953 aubo ropr nyny 129 holt nyny 264 many records of n a big game an boone & crockett c 1958 edbo aubo ropr nyny 547 many mammals of north america hall, er; kelson, kr 1959 aubo ucap beca 586 many wildlife of mexico leopold, as 1959 vipr nyny 304 many wildlife in america aubo matthiessen,p 1959 many principals of mammalogy aubo repu nyny 335 davis, de; golley, f 1963 blsp loen 308 aubo many guide, study of productivi golley, fb; buechne 1968 aubo jhpr bamd 769 many mammals of the world walker,ep; paradis 1968 whfr sfca 458 aubo many wildlife ecology moen, an 1973 aubo utop toon 438 many the mammals of canada 1974 banfield, awf aubo repu nyny 1023 dome bioenergetics and growth brody,s 1945 edbo coup itny 1463 dome duke's physiol domest anim swenson, mj 1970

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SERIALS

CODEN	vo-nu	BEPA	ENPA	GENS	SPEC KEWO	AUTHORS	YEAR
MDCBA	5	1	64	odvi	w-tailed deer of minnesota	erickson,ab; gunv	1961
MDCRA	14	1	80	odvi	michigan white-tailed deer	jenkins,dh; bartl	1959
WCDBA	14	1	282	odvi	white-tailed deer, wiscons	dahlberg,bl; guet	1956
CODEN	vo-nu	BEPA	ENPA	GENS	SPEC KEWO	AUTHOR S	YEAR
AZWBA	3	1	10 9	odhe	in arizona chaparral	swank,wg	1958
CFGGA	8	1	163	odhe	life hist, managemt, calif	taber,rd; dasmann	1958
CODEN	vo-nu	BEPA	ENPA	GENS	SPEC KEWO	AUTHORS	YEAR
UCPZA	88	1	209	ceel	tule elk: hist, behav, eco	mccullough,dr	1969
WLMOA	16	1	49	ceel	status, ecol, roosevel elk	harper,ja; harn/	1967
WLMOA	24	L	00	ceet	the sun river elk herd	knight, rr	1970
60.DEN				071170			
CODEN	VO-NU	BEPA	ENPA	GENS	SPEC KEWO	AUTHORS	YEAK
NCANA NCANA	101	1 437	436 735	alal alal	ecol, proc inter sym, pt 1 ecol, proc inter sym, pt 2	bedard,j bedard,j	1974 1974
CODEN	vo-nu	BEPA	ENPA	GENS	SPEC KEWO	AUTHORS	YEAR
BPURD	2	1	215	rata	ecol, caribou, prudhoe bay	white, rg; thomso/	1975
CWRSB	38	1	71	rata	biology, kaminuriak popula	dauphine,tc,jr	1976
UABPA	8	1	82	rata	ecology, managment, sweden	skunke,f	1969
CODEN	vo-nu	BEPA	ENPA	GENS	SPEC KEWO	AUTHORS	YEAR
WMBAA	10 A	1	79	rata	prelim investigation, pt l	banfield,awf	1954
WMBAA WMBAA WMBAA	10A 10B 12	1 1 1	79 112 148	rata rata rata	prelim investigation, pt 1 prelim investigation, pt 2 caribou, continued studies	banfield,awf banfield,awf kelsall.ip	1954 1954 1957

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CODEN	VO-NU	BEPA	ENPA	GENS	SPEC	KEWO			AUTHORS	YEAR
AMNAA	432	257	354	anam	life	hist,	ecology,	texas	buechner,hk	1950
JOMAA	3	82	105	anam	the p	prong-1	norn		skinner,mp	1922

CODEN VO-NU BEPA ENPA GENS SPEC KEWO------ AUTHORS------ YEARAMNAA 24--3 505580 ov-- distribut, variat, no amer cowan, imct1940AZWBA 1---- 1153 ov-- desert bighornrusso, jp1956WLMOA 4---- 1174 ov-- united sta, past to future buechner, hk1960

CODEN VO-NU BEPA ENPA GENS SPEC KEWO------ AUTHORS------ YEARAMNAA 56--2 297 324 ovca ecology of mountain sheep mccann,1j1956WGFBA 1---- 1127 ovca wyoming bighorn studyhoness,rf; frost, 1942XNFSA 6---- 1242 ovca th bighorn of death valley welles,re; welles 1961

CODEN	VO-NU	BEPA	ENPA	GENS	SPEC KEWO	AUTHOR S	YEAR
CAFNA	811	1	22	oram	observat, brit col, canada	holroyd,jc	1967
CGFPA	8	1	23	oram	liter review on ecology of	hibbs,1d	1966

CODEN VO-NU BEPA ENPA GENS SPEC KEWO----- AUTHORS----- YEAR NATUA 221-- 59 60 dada geographi var, fallow deer chapman,di; chapm 1969

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Proceedings of the White-tailed Deer in the Southern Forest Habitat

Symposium Transactions of the Annual Meeting of the Northeast Deer Study Group (Annual meeting beginning 1964) Proceedings of the North American Moose Conference (Vol. 5, 1968) Proceedings of the International Reindeer/Caribou Symposium (First meeting in 1977, second in 1980) Proceedings of the Biennial Antelope States Workshop Transactions of the Interstate Antelope Conference Transactions of the North American Wild Sheep Conference- (Second meeting in 1976) Transactions of the Desert Bighorn Council (Annual meeting beginning 1957) Proceedings of the International Mountain Goat Symposium Proceedings of the Annual Conference of Western Association of State Game &

Fish Commissioners (Annual conference beginning 1921)

LIST OF PUBLISHERS - PART III

aakn	Alfred A. Knopf	New York	nyny
acpr	Academic Press	New York	nyny
blsp	Blackwell Scientific Publications	London	loen
coup	Cornell University Press	Ithaca, NY	itny
cscs	Charles Scribner's Sons	New York	nyny
dalt	Dalton	Lavenheim, England	laen
dipr	Dial Press, The	New York	nyny
dohr	Dowden, Hutchinson & Ross	Stroudsburg, PA	stpa
doup	Doubleday, Pace, & Co.	New York	nyny
hocl	Hollis & Carter Ltd.	London	loen
huho	Hurd Houghton	New York	nyny
holt	Holt	New York	nyny

Publishers continued on the next page

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jhpr	John Hopkins Press	Baltimore, MD	bamd
jwis	John Wiley and Sons, Inc.	New York	nyny
lefe	Lea and Febiger	Philadelphia, PA	phpa
1mep	Lange Medical Publishers	Los Altos, CA	11ca
macm	MacMillan Co.	New York	nyny
mhbc	McGraw-Hill Book Co., Inc.	New York	nyny
moco	C. V. Mosley Company	St. Louis, MO	salo
nhfg	New Hampshire Fish & Game Dept.	Concord, NH	conh
omcc	Olin Mathieson Chem. Corp.	E. Alton, IL	eail
orpr	Oriel Press	Newcastle Upon Tyne, Eng.	nute
oxup	Oxford University Press	London	loen
qupr	Queen's Printer	Ottowa, Canada	oton
ropr	Ronald Press	New York	nyny
repu	Reinhold Publishing	New York	nyny
rokp	Routledge & K. Paul	London	10en
saco	Saunders Publishing Company	Philadelphia, PA	phpa
stac	The Stackpole Company	Harrisburg, PA	hapa
swap	Swallow Press	Athens, OH	atoh
t hc r	Thomas Crowell Co.	New York	nyny
ucap	University of California Press	Berkely, CA	beca
uchp	University of Chicago Press	Chicago, IL	chil
ukap	University of Kansas Press	Lawrence, KA	1aka
utop	University of Toronto Press	Toronto, Ontario	toon
usgp	U. S. Government Printing Office	Washington D. C.	wadc
vipr	Viking Press	New York	nyny
wbsc	W. B. Saunders Co.	Philadelphia	phpa
whfr	W. H. Freeman Co.	San Francisco, CA	sfca
wiwe	Winchester-Western Press	East Alton, Il	eail

x

GLOSSARY OF CODE NAMES, PART III

Code names (CODEN) of Serials are defined in a GLOSSARY OF CODENS at the end of each CHAPTER. The GLOSSARY below includes the CODENS listed as Serials in this PART I. It is a miniature version of the lists given at the ends of CHAPTERS.

AMNAA American Midland Naturalist AZWBA Arizona Game and Fish Department Wildlife Bulletin BPURD Biol. Pap. Univ. Alaska Spec. Rep. CAFNA Canadian Field Naturalist CFGGA California Department of Fish and Game, Game Bulletin CGFPA Colorado Division of Game, Fish, and Parks Special Report CWRSB Canadian Wildlife Service Report and Management Bull. Series JOMAA Journal of Mammalogy MDCBA Minnesota Deptartment of Conservation Technical Bulletin MDCRA Michigan Department of Conservation Game Division Report NATUA Nature NCANA Naturaliste Canadien, Le UABPA Proceedings of the Utah Academy of Sciences, Arts and Letters UCPZA University of California Publications in Zoology WCDBA Wisconsin Department of Natural Resources Technical Bulletin WGFBA Wyoming Game and Fish Commission Bulletin WLMOA Wildlife Monographs WMBAA Wildlife Management Bulletin

XNFSA U S National Park Service Fauna of the National Parks of the United States, Fauna Series

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THE BIOLOGY AND MANAGEMENT OF WILD RUMINANTS

CHAPTER SIX

SYSTEMS PHYSIOLOGY

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Published by

CornerBrook Press Box 106 Lansing, N.Y. 14882

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Library of Congress Catalog Card Number 80-70984

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Animals are complex biochemical entities, with specialized organs that perform particular functions. Organs have functional relationships with other body parts, and those with related functions are grouped together as systems.

The physiological functions of the systems of wild ruminants have not been studied very thoroughly. This is understandable because of the costs and difficulties involved. Their importance, however, is great, since physiological functions encompass all of the biochemistry of life. They have been given considerable attention by domestic animal scientists, and <u>Duke's</u> <u>Physiology of Domestic Animals</u> (Swenson 1970) is a classic text on this general subject.

A number of vital signs, including heart rates, respiration rates, and body temperatures are considered in this chapter. Signs indicate the status of the functions of some rather important body systems. Many of the functions are of direct importance to two or more systems. Cardiovascular and respiratory functions are interrelated, and these interact with the products of gastrointestinal functions. They also indicate the presence of internal responses that are not always accompanied by overt behavioral responses. Heart rates, for example, may accelerate as a fright response, even though the animal may not run away.

Vital signs also show diurnal and seasonal variations. Heart rates of deer, for example, are reduced when an animal is at rest each day, a short-term energy conservation adaptation, and they are reduced in winter when deer are less active, a long-term and very important energy conservation adaptation.

The organization of CHAPTER 6 is based on commonly-recognized systems of the body. The TOPICS describing the functions of each system are presented in the same order as the UNITS in CHAPTER 1 which contained descriptions of system characteristics.

LITERATURE CITED

Swenson, M. J., Ed. 1970. Dukes's physiology of domestic animals. Comstock Publishing Co., Cornell University, Ithaca, NY. 1463 pp.

REFERENCES, CHAPTER 6

SYSTEMS PHYSIOLOGY

BOOKS

TYPE	PUBL	CITY	PGES A	ANIM	KEY	WORDS					AUTHORS/EDITORS	YEAR
aubo	mhbc	nyny	895		a te	extboo	ok of	genl	phys	iol	mitchell,ph	1956
aubo	jwwa	pome	194		intr	oduc	to m	ammal	biol.	ogy	avis,fr	1 9 57
aubo	uchp	chil	642		basi	lc phy	rsiol	ogy			d'amour,fe	1961
aubo	moco	salo	547		text	book	of p	hysiol	Logy		<pre>tuttle,ww; schott</pre>	1961
aubo	haro	nyny	619		genl	l phys	iol,	molec	app:	rch	dowben,rm	1969
aubo	wiwi	bama	1964		text	bk ge	enl p	hysio,	, 4th	ed	davson,h	1970
aubo	hrwi	nyny	231		stru	ictu a	nd f	unctn,	2 nd	ed	<pre>griffin,;dr; novi</pre>	1970
edbo	coup	itny	1463		duke	e's ph	ysio	l of d	lom an	nim	swenson,mj,ed	1970
edbo	lefe	phpa	573		text	:book	of v	eterin	ı phys	sio	breazile, je, ed	1971
aubo	saco	phpa	.422		vert	ebrat	e ph	ysiolo	ogy		mccauley,wj	1971
aubo	vare	nyny	691		prir	nciple	s of	physi	lolog	у	hartenstein,r	1972
aubo	edar	loen	342		prin	nc ani	m ph	ysiol,	2nd	\mathbf{ed}	wood,dw	1974
aubo	hein	loen	660		bio]	logy d	f mar	mmals,	4th	ed	<pre>clegg,pc; clegg,a</pre>	1975
aubo	grst	nyny			intr	o to	phys	iolog,	5 v	ols	davson,h; segal,m	1975
aubo	prha	ecnj	848		gen	& con	ip phy	ysiol,	, 2nd	ed	hoar,ws	1975
aubo	haro	nyny	656		comp	phy	of a	nim, e	env aj	ppr	hill,rw	1976
aubo	hrwi	nyny	53 3		intr	o to	comp	aratv	phys	iol	goldstein,1	1977
aubo	macm	nyny	699		phys	, pri	nc,	adapt,	3rd	eđ	gordon,ms	1977
aubo	whfr	sfca	558		anim	nal ph	ysio	logy			<pre>eckert,r; randall</pre>	1978
aubo	cupr	nyny	560		phys	, ada	pt,	envir,	2nd	\mathbf{ed}	<pre>schmidt-nielsen,k</pre>	1979
aubo	macm	nyny	891		prin	ic ani	m ph	ysiol,	2 nd	eď	wilson,ja	1979

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TOPIC 1. GASTROINTESTINAL SYSTEM FUNCTIONS

The gastrointestinal system functions both mechanically and chemically as foods are ingested and broken down. The process of digestion involves the conversion of ingested food into forms that can be absorbed by the body and assimilated into body tissue. The preparation of ingested food for digestion begins with mechanical processes, and ends with biochemical processes.

Foods eaten "are merely the carriers of the nutrients and the potential energy . . . in a satisfactory diet." (Crampton and Harris 1969:5). Nutritive analyses must include not only investigations of food habits but also the digestion and assimilation of nutrients at different times of the year and under different range conditions. Microorganisms are absoslutely essential in ruminant digestion as there are no glands in the lining of the rumen and reticulum to secrete digestive enzymes.

The four functional parts of the mature ruminant's stomach are, in order of food passage: rumen, reticulum, omasum, and abomasum. At birth, the abomasum, or "true stomach," is the most well-developed, and the neonates are essentially monogastric animals. Ingested milk goes directly into the omasum via the oral groove, a passage that opens to allow milk to go to the omasum but closes when forage is swallowed, diverting that to the rumen. As the amount of forage ingested increases, the rumen develops thick muscular walls that contract rhythmically, promoting the breakdown and sorting of the ingesta for additional mastication of the coarser particles and passage of the finer particles to the reticulum, omasum, and abomasum.

Saliva functions as a lubricant and has a part in maintaining the fluid volume in the rumen. It assists in digestion to a very limited extent as the microorganisms in the rumen are almost totally responsible for the chemical breakdown of food in the rumen and reticulum. Nitrogen in the saliva is a substrate for protein synthesis by the rumen microorganisms, and their protein is subsequently useful to the ruminant host (Annison and Lewis 1959).

The rumen is lined with very rudimentary papillae at birth, and their development is dependent on the presence of volatile fatty acids (VFA's), the end products of rumen fermentation (Flatt et al. 1958), which are an energy source for the host animal. The papillae lining the interior surface of the rumen elongate as the amount of rumen fermentation increases, greatly increasing the surface area inside the rumen and permiting more rapid absorption of the VFA'S.

The reticulum, the second stomach compartment, is not as muscular as the rumen, and the internal surface has only a few papillae and no glands. It is separated from the rumen by the rumenoreticular fold, which acts like a dam to retain the larger particles of ingesta in the rumen. The smaller particles of ingesta are washed into the reticulum, and then they move on to the third stomach compartment, the omasum. The absorption of water from the ingesta seems to be the principal function of the omasum. The internal surface has many granular papillae that greatly increase the surface area for absorption. Since the omasum is relatively small, the processed ingesta must move on to the fourth compartment, the abomasum, rather quickly.

The abomasum has a muscular layer composed of thin fibers in small bundles, an outer serous layer, and an interepithelial lining that includes secretory glands (Short 1964). It is the only part of the stomach which secretes digestive juices (Phillipson 1970:454). He points out that the movement of ingesta into the abomasum from the omasum and from the omasum to the duodenum is influenced by the quantities of material in the recipient organ. It appears that both the amounts and chemical characteristics of ingested materials have become fairly uniform by the time the processed ingesta reaches the abomasum.

Ruminants have a small and nonsacrulated large intestine (Dukes 1955: 416). A diverticulum at the proximal end, called the cecum, does have some fermentation of food materials going on (Dukes 1955:428), but this is of little overall importance.

A series of mechanical actions keeps the ingesta moving through the gastrointestinal tract. Pressure or strain gauges are used to measure movement, especially in the rumen and reticulum. Both in vivo and in vitro techniques are used to study fermentation and digestion. Rumen fistulas, openings to the rumen through lateral incisions in the body walls, have been used extensively in studies on cattle and sheep, making it possible to suspend nylon mesh sacks of forage in the rumen for in vivo fermentation analyses, and to collect rumen fluid with a minimum of disturbance to the animal for in vitro measurements of fermentation. Several researchers have used fistulas on wild ruminants held in captivity as well.

The use of <u>in vitro</u> fermentation techniques, with flasks of forage samples and rumen fluid maintained at rumen temperatures in a water bath, is an inexpensive way to study fermentation rates and processes. The products of fermentation can be conveniently analysed, but with caution since artificial rumens are not identical to natural (<u>in vivo</u>) ones. Additional information and photos of a fistulated deer and <u>an in vitro</u> fermentation bath are in Moen (1973:149-159).

This topic contains three units describing the mechanical and chemical processes of digestion. Since gastrointestinal functions are an important and well-studied aspect of domestic animal science, several books which deal primarily with this topic are listed as references after the LITERATURE CITED. Serials describing gastrointestinal functions are listed after each of the units.

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edbo	butt	10en	297	rumi	digestiv physiology of rum lewis,d,ed	1961
edbo	butt	wadc	480	rumi	physiology of digesti, rum dougherty, rw, ed	1965
edbo	libr	boma	261	many	recent advanc in anim nutr abrams, jt, ed	1966
aubo	acpr	nyny	533	rumi	the rumen and its microbes hungate, re	1966
edbo	acpr	nyny	427	many	comparativ nutrition, wild crawford, ma, ed	1968
aubo	amph	wash		rumi	handbook of physiol: alime code, cf, ed; heide	1968
aubo	шосо	salo	262	many	physiol of gastroint tract texter, ce, jr; ch/	1968
aubo	dcdh	coor	316	rumi	digest physiology, nutriti church,dc	1969
aubo	whfr	sfca	753	many	applied animal nutrition crampton, ew; harr	1969
edbo	orpr	nute	636	rumi	physiolog digest, metaboli phillipson, at, ed	1970
aubo	orst	coor	801	rumi	dig physiol, nut of ruminan church, dc	1971
edbo	dcch	coor	350	rumi	dig phys, nut of ruminants church, dc, ed	1972
aubo	whfr	sfca	458	rumi	wildlife ecology: an analy moen, an	1973
edbo	iepu	nyny	576	many	physiolog adapta to enviro vernberg fj,ed	1974
edbo	unep	arau	602	rumi	digesti, metaboli, ruminan mcdonald, iw; warn	1975
edbo	coup	itny	1020	rumi	physiolog domestic animals dukes,hh	1955

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UNIT 1.1: MECHANICAL FUNCTIONS

Mechanical functions keep the food materials moving through the alimentary canal. These begin with the prehension and ingestion of food, followed by mastication, swallowing, rumination (including regurgitation, mastication again, swallowing, and rumen contractions), passage to the omasum and abomasum, intestinal contractions, and the opening and closing of valves between the parts of the gastrointestinal tract. As the food passes through the alimentary canal, saliva and digestive juices are mixed with the food, and the enzymes in these secretions compliment the fermentations by rumen microorgaisms in the overall process of digestion in the ruminant. Food in the rumen and reticulum is actively moved and churned as it is metabolized by rumen microorganisms. The movement results in the separation of the ingesta into layers, with the finer particles moving from the rumen and reticulum onto the omasum and abomasum. Finally, defecation of undigested food materials occurs.

Prehension. Prehension is accomplished in different ways among wild ruminants. Browse is removed by grinding it off with the molars, and low herbaceous vegetation is ingested by seizing the plant material between the lower incisors and upper gum. Plant parts, such as flowers, can be carefully and selectively removed. Moose sometimes feed underwater, submerging their heads, blocking off the nasal passages, and grasping the vegetation in their mouths. The different abilities of wild ruminants in selective foraging contribute to differences in their food habits; Cervids are generally more selective feeders than Bovids.

<u>Mastication</u>. Ruminants chew the forage some when it is ingested and again after it has been regurgitated. Most of the mechanical reductions in particle size occurs when the animal chews the regurgitated bolus, usually while bedded between feedings. The vertical and lateral movements of the jaws result in a rotary motion centered on one side at a time. The teeth wear roughly, increasing their grinding efficiency as particle sizes are reduced and surface areas increased, resulting in greater efficiencies in fermentation.

Swallowing. Swallowing is a complex reflex act in the ruminant. Milk ingested by nursing young goes into the omasum via the closed esophageal groove. After weaning, the esophageal groove no longer functions and water goes directly to the rumen with the rest of the ingesta. The water content of the forage is important for ruminants as foods that are not sufficiently moist do not start the muscular contractions involved in swallowing. Salivation also occurs, and the saliva is mixed with the food during mastication until the bolus is swallowed. When swallowing occurs, respiration is inhibited and the glottis and posterior end of the nares are closed, preventing the possibility of food entering the respiratory tract The food swallowed enters the rumen where rumination adds via the trachea. to the mechanical treatment of the food, followed by chemical fermentation in the rumen and reticulum.

includes Rumination. Rumination regurgitation, mastication and salivation again, followed by swallowing. The frequencies and durations of these events are dependent on the characteristics of the diet. Rumination begins a few days after birth, and adult-type rumination rhythms occur within a few weeks. Regurgitation involves contraction of the skeletal Dzuik et al. (1963:782) observed that white-tailed deer sometimes muscles. regurgitated ingesta, swallowed without mastication and regurgitated again in less than ten seconds. The rapid reswallowing may have been associated with characteristics of the bolus. The contraction patterns of the rumen and reticulum of a showed that contractions of the musculature of the rumen and reticulum were very closely associated (Dziuk et al. 1963). The first or primary rumen contraction, involving all parts of the rumen, followed two successive contractions of the reticulum, or "reticular doublet." A secondary rumen contraction, involving the dorsal sac, posterior dorsal blind sac, and the ventral sac of the rumen, sometimes occurred. The duration of the ruminoreticulum cycle in the white-tailed deer was 20-30 seconds. The frequency of reticular and primary rumen contractions varied with the activity of the deer, with 1.8 contractions per minute when standing and 2.2 contractions per minute when bedded. Secondary rumen contractions varied from 0.6 to 0.8 contractions per minute while standing and 0.2 to 0.5 contractions per minute when bedded. Dzuik et al. concluded that similarity of the pressure events in deer, cattle, and sheep were marked.

Stomach and intestinal movements. In the omasum, contractions of the muscular walls compress and pulverize the food and much of the water is absorbed. The food materials are then passed on to the abomasum where gastric juice is secreted and the fluid content is restored to the approximate level in the omasum. The hydrochloric acid content of the gastric juice causes the pH to fall to 1.5 - 3.0. The protozoa disintegrate there and some of the bacteria are killed. The food materials pass through the abomasum quite rapidly and move on into the small and then the large intestine (Annison and Lewis 1959:18).

Defecation. When the food reaches the rectum it consists largely of undigestible materials and water. The fecal mass also includes tissue that has been removed from the lining of the alimentary canal, as well as the remains of rumen microorganisms that have escaped digestion in the small intestine. The water content is variable, depending on species, diets, and time of year. Defecation occurs several times a day, often but not necessarily shortly after the animal leaves its bed. The number of defecations per day have been used in estimating populations from pellet group counts.

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 36--3 474 476 odhe values, alimen canal ph browman, 1g; sears 1955 JOMAA 46--2 196 199 odhe reminoreticular characteri short, hl; medin, / 1965 JWMAA 28--3 435 444 odhe defecation rates of mule d smith, ad 1964 JWMAA 34--1 29 36 odhe ceel, freq dist pellet grp mcconnell, br; smit 1970 UASPA 32--- 59 odhe indices of carc fat, color anderson, ae; med/ 1972 64

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR ATRLA 15--- 253 268 ceel relatio, age, size, poland dzieciolowski,r 1970 JWMAA 24--4 429 429 ceel dyes to mark ruminan feces kindel,f 1960 JWMAA 29--2 406 407 ceel determ defeca rate for elk neff,dj; wallmo,/ 1965 JWMAA 35--4 673 680 ceel rumen characteristi, red d prins,ra; geelen, 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 40--2 374 375 alal dail wint pell gr, bed, al franzmann,aw; ar/ 1976 NCANA 95--5 1153 1157 alal [numb pellet-gro each day] desmeules,p 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AVSPA 57--- 1 18 rata topograph, internal organs engebretsen, rh 1975 NJZOA 24--4 407 417 rata morph, fat stor, org wt, wint krog, j; wika, m; / 1976 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 13--- 499 509 bibo capac, weigh, walls, diges gill, j 1968 ATRLA 14--- 349 402 bibo morphology digestive tract pytel, sm 1969 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR FEPRA 27--6 1361 1366 rumi regulation of feed intake baile, ca 1968

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UNIT 1.2: FERMENTATION AND DIGESTION

Food materials are complex structures that must be broken down by rumen fermentation and digestion before the nutrients can be assimilated into new body tissue. Foods can be divided into two physical groups, water and dry matter, and the latter can be divided into chemical groups with similar characteristics as outlined below.

I. Water

II. Dry Matter

A. Organic substances

1. Nigrogenous compounds (crude protein)

a. True protein

b. Non-protein nitrogenous materials

- 2. Non-nitrogenus substances
 - a. Carbohydrates
 - 1. Soluble carbohydrates (nitrogen free extract)
 - 2. Insoluble carbohydrates (crude fiber)
- B. Inorganic substances (ash)

1. Salts

2. Mineral matter

Rumen microorganisms break the food materials down into substrates that can be used for their own metabolism, and the metabolic by-products of the microorganisms and the microbes themselves are then chemically processed by the ruminant host for use in their own energy, protein, mineral, vitamin, and water metabolism. The ruminant host and the rumen microorganism represent a highly developed symbiotic relationship. Nearly 900 strains of bacteria and more than 100 species of protozoa have been identified in the anaerobic rumen environment of different ruminants (Moen 1973:151). No individual has this many kinds, of course; variations in the number present of each strain are primarily due to differences in diets.

The actual number of microorganisms in the rumen is very large. The mean number of protozoa may be 10^6 or one million in each milliliter of rumen contents (Pantelouris 1967:193), and of bacteria about 10^9 to 10^{10} in each milliliter (Annison and Lewis 1959:33). Expressed in long-hand, the number is 10,000,000 (ten billion) bacteria in each milliliter of rumen contents.

The rumen is an open system that depends on the flow of food materials into it and the flow of microorganism metabolites and food residues out. When the supply of food in the rumen is maintained by frequent intake, conditions in the rumen remain quite constant as the microorganisms have a stable substrate for the maintenance of their own metabolic activities. If food remains undigested in the rumen, appetite and rumen movement stop. If the rumen is then filled with actively fermenting rumen fluid from other animals, both appetite and rumen movement begin again (Nagy et al. 1967:447).

The failure of deer to survive when fed artificially in the winter has been attributed to a lack of the types of rumen microorganisms necessary for breakdown of such things as alfalfa hay. Research by Nagy (1967:446) has shown that no major adjustments in the microbial spectrum were necessary when deer were fed alfalfa hay, and other factors appear to be more important when deer are found dead with their rumens full of nutritious hay and other artificial feeds. Nagy suggests that as the amount of forage ingested decreases under starvation conditions there is a concomitant loss in the number of properly functioning rumen microorganisms. When the rumen contains a relatively small number of active microorganisms, the food residues would, in normal passage through the intestinal tract, tend to remove these microorganisms at a faster rate than their population growth can sustain. A supply of new forage then becomes residual in the rumen as death occurs.

Digestion is also affected by oils in certain plant materials. Oils of big sage-brush, for example, had in inhibitory action on bacteria in the rumen of mule deer, with a subsequent decrease in the rate of cellulose digestion (Nagy et al. 1964:785). Appetite and rumen movement stopped completely when three 7-pound portions of sage brush were introduced into a steer through a rumen fistula. When range conditions deteriorate to the point where wild ruminants are forced to eat certain plants that might otherwise be avoided, reductions in the digestive efficiencies are inevitable.

It is clear that rumen fermentation continues only if the substrate is adequate for support of microorganism metabolism. Not only must there be an adequate quantity, but the balance between carbohydrates and nitrogen is also important. The use of starch is an important factor in maintaining a flourishing rumen flora, and the digestion of starch is influenced by the quantity and nature of the nitrogenous components of the diet. A minimal level of protein is essential for supplying the nitrogen requirements of the rumen nicroorganisms (Annison and Lewis 1959:79-83). Two main conclusions are usually drawn from experiments on carbohydrate and nitrogen effects. One, there is a more rapid attack by the rumen microorganisms upon the fibrous components of the ration as the protein intake is increased, and two, there is better use of protein in the presence of added carbohydrate (Annison and Lewis 1959:109).

The complex ecology of the rumen can be at least partially controlled when animals in captivity are fed known quantities of diets with selected nutritive characteristics. Wild ruminants are subject to natural changes in range conditions, resulting in a much more complex rumen ecology than that of penned domestic ruminants. This increase in complexity adds many problems to nutritive analyses of wild ruminants, but it also presents many new challenges to creative thinking. The opportunities for nutritive analyses beyond the food habits level are many in wild ruminant ecology.

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR XAMPA 1147-159 163 od biol rel, rumen flor, faun nagy, jg 1970

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ceel continued on the next page

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AZFEA 127-- 1 76 rata rumen ciliate fauna finla westerling, b 1970 rata seas variation, gluc metab luick, jr; person/ 1973 BJNUA 29--2 245 259 CJZOA 36--5 819 835 rata rumen ciliates, canad arct lubinsky,g 1958 CJZOA 36--6 937 959 rata rumen ciliates, canad arct lubinsky,g 1958 CJZOA 54--1 55 rata gluc metab, lactating rein white, rg; luick, j 1976 64 HLTPA 21--5 657 rata radiocesium, lichen, alask holleman, df; lui/ 1971 666 LAANA 28--- 175 180 rata comp, reindeer, sheep dige eriksson,s; schme 1962 1968 SZSLA 21--- 109 115 rata wint nutr, wld reind, norw gaare,e SZSLA 21--- 117 rata nutr semi-domestic reindee steen,e 1968 128

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARJWMAA 33--2 437 439 anam ruminoreticular vfa conten nagy,jg; williams 1969TAMSA 72--3 248 252 anam intestin amoeba from prong noble,ga1953XARRA 148-- 14 anam starvation with stomach fu pearson,ha1969CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARYEARAPMBA 15--6 1450 1451 bibi rumen microorganisms, utah pearson,ha1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR UCPZA 53--6 237 261 ovca ciliates, sierra nev bigho bush,m; kofoid,c 1948 JWMAA 36--3 924 932 ovca env sour of var, physi val franzmann,aw 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWIDA 7---3 139 141 ovda physiologic values of ston franzmann, aw 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR CJZOA 41--1 29 32 obmo rumen ciliate, north canad lubinsky,g 1963

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ZEJAA 8---3 97 111 caca [topog organ, neck, chest] hofmann,r 1962

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR EJBCA 63--2 441 448 many carbohyd, pancre ribonucle beintema, jj; gaa/ 1976 FEPRA 27--6 1361 1366 rumi regulation of feed intake baile, ca 1968 JANSA 35--6 1271 1274 dosh effect of watr, nutrnt dig asplund, jm; pfand 1972 JONUA 86--3 281 288 rumi blood urea, protein intake preston, rl; schn/ 1965 JPROA 21--1 26 32 wiru rumen ciliate fauna, 3 wr dehority, ba 1974 JWMAA 32--1 198 meth microb diges, pos mor bruggemann, j; gi/ 1968 207 JWMAA 38--4 948 949 wiru differentiatio, pellet, ph howard, vw, jr; del 1974 NARFA 29--2 511 511 rumi fact, rumina dur developme tulbaev, po 1959 NCANA 101-1 227 262 many energy req rumen fermenta gasaway, wc; coad 1974

UNIT 1.3: PRODUCTS OF FERMENTATION

Rumen microorganisms produce heat energy, gases, volatile fatty acids, nitrogen compounds, and vitamins as end-products of their metabolism. The heat energy given off is called the heat of fermentation, and contributes to the energy balance of the ruminant host. It is the result of exothermic metabolic reactions of the rumen microflora. Its role in the energy balance of free-ranging ruminants is discussed in PART V. The other products of rumen microorganism metabolism are discussed below.

GASES

Gases given off, primarily methane (CH_4) and carbon dioxide (CO_2) are either released directly into the atmosphere by eructation or absorbed by the blood and exhaled later as part of respiratory exchange. The composition of the rumen gases eructed do not vary much with rations. The total amount of gas formed is not affected much by the ration, but the quantity rises sharply after eating and then falls gradually (Dukes 1955: 382). The significance of eructation is in the loss of energy that it represents; about 8% of the digestible energy of the diet, varying with the digestibility of the food, is released in methane (Blaxter 1967:200; and Moen 1973:154).

VOLATILE FATTY ACIDS

Three volatile fatty acids (VFA's)--acetic, butyric, and proprionic-are invariably present in the active rumen as a result of microbial fermentation of carbohydrates (Annison and Lewis 1959:60). The total concentration and the amounts of individual acids are dependent on the composition of the ration and the feeding regime. Thus seasonal differences are expected in wild ruminants, and have been observed in mule deer (Short et al. 1966:466) and white-tailed deer (Short 1963).

Higher concentrations of VFA's are observed when easily fermented foods are ingested (Short 1963:186). There is an increased rate of rumen fermentation during the summer when the range forage is succulent, and a decrease in the fall and winter.

The concentration of volatile fatty acids in the rumen is not a simple and straight forward indication of the actual rate of VFA production, however. The concentration of VFA's at a given time depends not only on their rate of production in the rumen, but also on the rate of absorption from the rumen, rate of passage from the rumen to the omasum, dilution with saliva, use of VFA's by the rumen microorganisms, and conversion to other rumen metabolites (Annison and Lewis 1959:63).

The young ruminant has glucose and VFA concentrations like those of a monogastric animal in which most of the energy is absorbed in the form of glucose from the small intestine (Annison and Lewis 1959:73). As rumen fermentation increases, glucose concentrations fall to about one-half that of non-ruminants, and VFA concentrations increase. This change in carbohydrate metabolism is a physiological indication of the weaning process (Annison and Lewis 1959:20). Volatile fatty acids are readily absorbed from the rumen and are a major source of energy for the ruminant animal. Moen (1973:154) cites references indicating that at least 600 to 1200 Kcal of energy are absorbed as VFA's from the sheep rumen every 24 hours, and 6000 to 12000 Kcal from the cattle rumen. The latter has a much greater physical capacity than the former, of course. VFA production by deer is discussed in Short (1963) who stresses the need for additional understanding of such nutritional energy relationships in deer management.

NITROGEN COMPOUNDS

Microbial protein synthesized from amino acids and non-protein nitrogen sources is digested in the omasum and abomasum as microbial populations are rapidly turned over (Pantelouris 1967:192). A considerable portion of the protein requirement of ruminants is supplied in this way (Annison and Lewis 1959:19) as non-protein nitrogen is converted to microbial protein and digested and absorbed by the host. Urea is an example of a non-protein nitrogen compound that may be converted by rumen microbes to microbial tissue. Deer ingest urea and other non-protein nitrogenous compounds by consuming water that has been accumulating in pools where feces and urine have been deposited, by lapping urine from other deer, and by the cleaning of the fawns by the doe. Fawns at the Wildlife Ecology Laboratory have shown varying degrees of interest in lapping the urine from their penmates, sometimes refusing milk in preference for urine.

The protein contents of the rumen are often higher than the protein contents of the forages being ingested. The terminal two to three inches of growth on browse sampled by Bissel (1959:57) had 6.9% crude protein, but the rumen contents of nine deer on that range averaged 17.6% crude protein. Fourteen deer studied when range plants were dormant had range and rumen protein contents of 7.1% and 17.2% respectively in January and 6.1% and 15.1% in February. Three deer fed alfalfa pellets had rumen protein contents of 21.0%, 16.2% and 14.7%. Such increases in the nitrogen contents of rumens over those of ingested food materials can be accounted for by nitrogen recycling. The percent of urea recycled by white-tailed deer ranged from 92.3% to 40.6% as dietary protein content increased from 5% to 26% (Robbins et al. 1974).

VITAMINS

The vitamin requirements of adult ruminants are satisfied in part by the rumen microorganisms. Water soluble vitamins of the B complex and fatsoluble vitamin K are synthesized in the rumen if appropriate foods are eaten (Annison and Lewis 1959:20, 155). Vitamins A, D, and E must be supplied in the diet. The provision of at least part of the adult's vitamin requirements by rumen microflora illustrates how important the symbiotic relationship between microflora and ruminant host is. All of the metabolic products of the microflora, except the gases, are used by the ruminant host as sources of different nutrients. When each individual microbe dies, the cell is then digested and nutrients, especially protein, extracted. The rumen is a closed, anaerobic system that represents the link between the animal and its range.

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PRODUCTS OF FERMENTATION

BOOKS

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TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR edbo nhfg conh 256 odvi w-tail deer of new hampshi siegler, hr, ed 1968

SERIALS

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JANSA 38--1 186 191 odvi nitrogen metabolism of whi robbins, ct: prio/ 1974 JWMAA 27--2 184 195 odvi rumen ferment, energy rela short, hl 1963odvi var ruminoreticul contents short, h1; remmen/ 1969 JWMAA 33--1 187 191 JWMAA 33--1 204 odvi cage, metab, radioiso stud cowan, rl; hartso/ 1969 208 JWMAA 33--2 380 odvi rumino-retic char, 2 foods short, h1; segelq/ 1969 383 JWMAA 36--3 885 891 odvi digest, metaboli, aspen br ullrey, de; youat/ 1972 VJSCA 21--3 117 117 odvi prod enrgy, comp rum diges woodyard, gw; whel 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JANSA 39--1 236 236 odhe effect starva on rumen bac decalesta, ds; wa/ 1974 JWMAA 14--3 285 289 odhe sagebrush as a winter feed smith, ad 1950 JWMAA 28--4 785 **79**0 odhe essen oils, sageb, rum mic nagy, jg; steinho/ 1964 JWMAA 30--3 466 odhe seas varia, vol fatty acid short, hl; medin, / 1966 470 JWMAA 37--3 312 odhe effect nutr chan on captiv robinette,w1; ba/ 1973 326 JWMAA 39--3 601 604 odhe reticulo rum char mal nour dean, re; strick1/ 1975 JWMAA 39--4 663 669 odhe starving and re-feeding mu decalesta, ds; na/ 1975 NAWTA 36--- 153 162 odhe effects pestic, rumen bact barber, ta; nagy, j 1971 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR NCANA 101-1 227 262 alal energy req, rumen fermenta gassaway,wc; coad 1964

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BPURD 1---- 290 296 rata odvi, compar study rum met bruggemann,j; dr/ 1975 CJZOA 54--5 737 751 rata dig energ in, glucos synth mcewan,eh; white/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 33--2 437 439 anam ruminoreticula VFA content nagy,jg; williams 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR b1b1

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JASIA 60--3 393 398 doca exper, nutri, dairy heifer broster,wh⁻ tuck/ 1963 JONUA 101-- 1331 1342 doca diet nonprot N, urea kinet mugerwa, js; conra 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR AJBSA 20--5 967 973 dosh transf nitr, blood to rume weston,rh; hogan, 1967 BJNUA 21 -2 353 371 dosh metabolis of urea in sheep cocimano,mr; leng 1967 BJNUA 27--1 177 194 dosh ammonia and urea metabolis nolan,jv; leng,ra 1972 JANSA 35--6 1271 1274 dosh eff wat restric nutrnt dig asplund,jm; pfand 1972 PAANA VI--- 378 383 dosh urea metabolism in sheep cocimano,mr; leng 1966

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARAJPHA 197-1 115120rumi utilization of blood urea houpt,tr1959JDSCA 51--2265275rumi nitrogen utiliz, metabolis waldo,dr1968JONUA 22--2167182rumi value, urea, synth, protei harris,le; mitche 1941

CODEN	VO-NU	BEPA	ENPA ANIM	KEY WORDS	AUTHORS	YEAR
AJMEA	29	832	848	design, anal, isotop exper	zilversmit,db	1 9 60
CLCHA	179	92 1	925	singl reag meth, urea nitr	foster,1b; hochho	1971
ECOLA	525	93 5	939	model, absorp, ret ing ele	goldstein,ra; elw	1971
JANSA JANSA	252 261	593 119	593 128	dig prot est, nrc feed com develop, system, feed anal	knight,ad; harris van soest,pj	1966 1967
JBCHA JBCHA	583 583	873 905	904 922	meth, det biol value, prot biol val, prot dif int lev	mitchell,hh mitchell,hh	1924 1924

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR SCIEA 130-- 1192 1194 mamm microbial fermentation in hungate, re; phil/ 1959
CHAPTER 6, WORKSHEET 1.3a

Estimates of methane energy

The food energy lost in the methane (CH_4) represents about 8% of the total food energy (Blaxter 1967:198-200). The amount of energy lost in methane can be estimated for animals on a roughage diet with the equation:

MEPP = 4.28 + 0.059 ADEP

where MEPP = methane production as a percent of the gross energy of the forage ingested and ADEP is the apparent digestible energy of the forage expressed as a percent. Thus if ADEP = 50%, MEPP = 4.28 + (0.059)(50) = 72%. If ADEP = 80%, MEPP = 4.28 + (0.059)(80) = 9.0%. Note that these bracket the 8% figure given above.



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TOPIC 2. CARDIOVASCULAR SYSTEM FUNCTIONS

The cardiovascular system is responsible for the distribution of oxygen and nutrients to the cells throughout the body. The role of the heart in pumping blood has been known for a long time, and the exchange of O_2 and CO_2 at the respiratory surfaces of the lungs is an important function associated with the flow of blood. The number of beats per minute and the volume of blood pumped are both dependent on the level of activity and other factors affecting the oxygen and carbon dioxide concentrations in the blood.

The cardiac cycle may be described by beginning with ventricular systole, or contractions of the two ventricles. As these contractions occur, blood is forced out of the ventricles and into the pulmonary and systemic circiuts. The two atria, or receiving chambers of the heart, are in diastole, or a relaxed condition, and fill with blood that has been propelled through the arterial system to the venous system that leads to the atria. As the atria are filled with blood and ventricular diastole occurs, the ventricles are filled with blood from the atria in preparation for the next ventricular systole.

The amount of blood pumped with each cardiac cycle is dependent on the strength of ventricular systole. As the heart beats faster, stroke volumes may increase, but not necessarily in direct proportion to changes in heart rate. The total amount of blood present also affects the strength of the circulation pattern as decreased volumes will likely result in less flow from the atria to the ventricules and throughout the systemic circulation.

The circulating blood is an active nutrient and oxygen transport system. The chemistry of the blood is important because it determines how well the blood functions as a medium for the transport of oxygen and nutrients to the sites of metabolism. The red cell fraction of the circulating blood, for example, is important because oxygen is carried primarily by the chemical combination of oxygen with hemoglobin. Thus an anemic animal is as metabolically efficient as one with higher red cell and hemoglobin concentrations, and will consequently be less able to withstand the effects of cold weather, parasites, and other factors of metabolic and ecological importance.

Evaluations of heart rates when the animals are involved in different activities at different times of the year, the stroke volumes of each heart beat, the total amounts of blood present in the animal, red cell functions, chemical characteristics of the blood are all of potential importance when estimating the metabolic potentials of free-ranging animals.

LITERATURE CITED

McCauley, W. J. 1971. Vertebrate physiology. W. B. Saunders, Co., Philadelphia, PA. 422 pp.

REFERENCES, TOPIC 2

CARDIOVASCULAR SYSTEM FUNCTIONS

BOOKS

TYPE	PUBL	CITY	PGES	ANIM	КЕҮ	WORDS	AUTHORS/EDITORS	YEAR
aubo aubo aubo aubo aubo aubo	wbsc hrwi wbsc prha pepr wbsc	phpa nyny phpa ecnj oxen phpa	688 120 713 815 497 422		comp anin gen gene intr ver	parative animal physiol mal structure & functio and comparat anim phys eral & comparat physiol co anim physiol, geneti tebrate physiology	prosser,cl; brown griffin,dr florey,e hoar,ws pantelouris,lm mccauley,wj arabar rky inffac	1961 1962 1966 1966 1967 1971
aubo	cupr	caen	560		anir	n phys: adapta, environ	schmidt-nielson,k	1979

UNIT 2.1: HEART RATES

Electrocardiogram equipment, or even stethoscopes, are not standard equipment in the typical wildlife laboratory. Yet the heart rate is one of the first things to be checked when evaluating the health status of a person because it is a very important vital sign. Earlier societies considered the heart to be the center of life. Neglect of this very important organ and its vital signs in research on wild ruminants must be attributed to difficulties associated with cardiovascular research and not to a lack of importance of heart functions. Fortunately, radio telemetry now makes such research possible.

HEART RATES IN RELATION TO AGE

A heart rate pattern consistently observed in all species is that of a rapid decline in heart rates in the first few days or weeks or life. Heart rates of neonates are high, and they drop rapidly, logarithmically, until the rates characteristic of adults are reached. This shift from the high rates characteristic of neonates to the seasonally-variable characteristics of adult deer is observed by the end of the suckling period.

Curve-fitting of neonate and adult heart rates should be done without discontinuities where the two patterns merge, just as weight equations for different phases of growth were merged, as illustrated below.



Individual variations, experimental artifacts, and random errors contribute to mathematical discontinuities. Living animals exhibit a smooth transition in these functions from day to day, however, and mathematical representations must reflect that. The equations may have to be adjusted slightly to link directly with the equation for the other group. Adjustments are easily made in the equation for the young by elevating the entire equation, or by curve-fitting just two points--the intercept, a, and the point where the equation for the young merges with the adult equation. The adjustments illustrated below show how the equations can be merged without discontinuities.



Heart rates of the adults of large species are slower than adult heart rates of smaller species. This is characteristic of a wide range of species; very high heart rates are observed in some small animals, such as shrews, and very slow rates in large animals, such as elephants. It is apparently true for wild ruminants too. Elk have slower heart rates than deer, and moose are expected to have slower rates than elk. Data are too limited to derive mathematical expressions of heart rates in relation to the weights of different species, however.

HEART RATES IN RELATION TO ACTIVITY LEVELS

There is a general pattern of increased heart rates with increased physical activity. Heart rates in five major activities increase in the following order: bedded, standing, foraging, walking, and running. It would be very surprising if a species or even an individual showed a different pattern. Accelerations due to fright responses may occur in any activity, however, and upset the patterns based on physical activity alone.

The use of radio telemetry techniques for transmitting heart rates of unrestrained animals in their chosen activities provides the best information when activity patterns, other animals, and other stimuli are monitored concurrently. This has been done during experiments on white-tailed deer at the Wildlife Ecology Laboratory. Insights gained in such carefullycontrolled conditions are useful when interpreting variations observed in free-ranging animals.

HEART RATES IN RELATION TO TIME OF YEAR

Measurements of heart rates of wild ruminants must be made carefully over time. Heart rates of white-tailed deer varied during the year with the highest rates observed in the summer when metabolism was highest, and lowest in the winter when metabolism was lowest (Moen 1978). Sine wave variations in adult heart rates are illustrated on the previous page, showing highest heart rates in the summer and lowest in the winter.

The higher heart rate: higher metabolism pattern was evident in all activities, as heart rates were seasonally higher when the deer were walking than when they were standing, and higher when standing than bedded. The higher heart rates, and metabolism too, reflect a synchrony between range resources and animal requirements. When range resources are down, animal requirements are down, and when resources are up (during the growing season), productive functions rise to their highest levels. This is ecologically very reasonable, and represents a distinct energy conservation adaptation.

HEART RATES IN RELATION TO TRANSIENT STIMULI

While there are general heart-rate patterns in relation to age, weight, and time of year, there are also many variations due to transient stimuli. Many of these transients are very natural; the approach of a dam to its fawn may cause tachycardia, or a marked acceleration in the heart rate of the fawn. Rain and thunder also cause tachycardia; as do other animals and other natural noises. (Moen and Chevalier 1977). Sometimes heart rates are slowed; bradycardia is a characteristic fright response in neonates. Heart rate responses of whitetail fawns to recorded wolf howls included both tachycardia and bradycardia, as well as a patterned response to howl frequency and loudness (Moen et al. 1978).

The dynamics of the environment and their effects on heart rates are very great. Overall heart rate patterns in relation to both activity and time of year were observed at the Wildlife Ecology Laboratory because heart rates were recorded only when the experimental animals were being observed, relating behavior to heart rates. Further, deviations in the heart rates due to transient stimuli were also evaluated and the undisturbed heart rates in different activities determined. These kinds of data collections and analyses rather than strictly timed measurements at arbitrary intervals, were very important factors in recognizing the sine wave patterns described in Moen (1978). Another important factor in the successful recognition of patterns over the annual cycle was the careful observations and measurements on several animals for 24-hour or larger periods over several successive annual cycles. Attention to these details with animals that were treated as naturally as possible was very important in the synthesis of relationships into whole pictures, unencumbered by deviations due to experimental mathematical or statistical procedures.

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HEART RATES

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS AUTHORS	YEAR
AHEMA	3	250	261	odvi	maj arteries, shoulder,arm bisaillon,a	1974
AJVRA AJVRA	301 33-12	143 2545	148 2549	odvi odvi	<pre>serum prot, normal, arthri sikes,d: hayes,f/ blood seru, arthrit reumat sikes,d; kistner/</pre>	1969 1972
AKASA	30	50	51	odvi	electroph pattern, 2 subsp jackman,gs; garne	1976
CBPAB	62a-4	885	888	odvi	chngs hrtrate, grwth, act jacobsen,nk	1979
CJZOA CJZOA	535 536	1207 679	1210 685	odvi odvi	heart rate, fawn, wolf how moen, an; dellafe/ amb temp eff, physio trait holter,jb; urban/	1978 1975
JOMAA	602	343	349	odvi	alarm bradycardia in fawns jacobsen, n	1979
JWMAA	373	413	417	odvi	seasonal heart rates, meta moen, an	1978
SOVEA	261	51	58	odvi	electrocardiogram, wh-t de szabuniewicz,m; /	1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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odhe

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBCPA 61A-- 43 48 ceel oxygen utiliza elk calves cohen,y:robbins,/ 1977 JOMAA 49--4 790 791 ceel electrocard, rocky mountai herin,ra 1968 ZEJAA 4---4 171 177 ceel [regulation of blood pres] jaczewski,z; ja/ 1958

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JEBPA 12--4 347 349 alal rata, card comp emot stres roshchevskii,mp/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR APSSA 396-- 96 96 rata blood circulation, finnish hirvonen,1; jar/ 1973 AVSPA 57--- 1 18 rata topograph, internal organs engebretsen,rh 1975 LBANA 13--3 183 186 rata electcardiogram, reindeer timisjarvi,j; / 1979 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

anam

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JZAMD 1---2 20 22 bibi odvi, electrocrdgrph obsrv jankus, ef; good, a 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR VETRA 97-12 230 231 ovca electrocrdiogram, big-horn rezakhani,a; edj/ 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 39--4 554 559 obmo serologica evid, relations moody,pa 1958

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 12-32 1275 1276 bibo electrocardi, experi death nagorski,f; grodz 1967

CHAPTER 6, WORKSHEET 2.1a

Heart rates of white-tailed deer fawns (odvi) in relation to age and activity

Heart rates of whitetail fawns, measured at the Wildlife Ecology Laboratory with a radio telemetry system, are related to activities and the age of the fawn. There is a rapid decline in the heart rates in different activities shortly after birth, and the rates continue to decline until 105 days of age. After this age, the heart rates vary seasonally, without regard to the age of the deer, and the equations for adult deer apply. Equations for predicting heart rates for fawns from 1-105 AGDA in five activities are listed below.

Activity	Equation
Bedded	BHRM = 250 - 35.5 ln AGDA
Standing	SHRM = 306 - 43.8 ln AGDA
Walking	WHRM = 390 - 59.1 1n AGDA
Foraging	FHRM = 321 - 45.9 1n AGDA
Running	RHRM = 383 - 40.0 ln AGDA

Plot and label the heart rates (HERA) for each of the activities on the next page. Note especially the heart rates at AGDA = 105 (JDAY = 256 if birth date = 151) comparing them to rates calculated in WORKSHEET 2.1b for adult deer on JDAY 256.

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AGDA	$= 0 \frac{1}{0}$	$\frac{1}{030}$		<u>†</u> † 09	<u>†</u> 1	<u>†</u> † 120	$\frac{1}{150}$			$\frac{1}{210}$	$\frac{1}{2^{7}}$	<u>†</u> 40	1 270	<u>†</u> <u>†</u> 30	- 50
JDAY	=				256	28	6 3	316	346)	11	4	1	71	-

CHAPTER 6, WORKSHEET 2.1b

Heart rates of adult white-tailed deer (odvi) in relation to activity and time of year

Heart rates of adult white-tailed deer have been studied for several years at the Wildlife Ecology Laboratory, Cornell University. Equations for bedding, (BHRM), standing, (SHRM), walking, (WHRM), foraging (FHRM), and running (RHRM) heart rates per minute given in Moen (1978:722) are:

BHRM =	14.6878 si	n [(JDAY)(0.980	53) + 226] -	0.1640 + 72
SHRM =	17.2907 si	n [(JDAY)(0.980	53) + 226] +	0.4346] + 86
WHRM =	[15.5409 sim	n [(JDAY)(0.980	53) + 226] -	0.3256 + 102
FHRM =	[18.7914 si	n [(JDAY)(0.986	53) + 226] +	0.1934] + 90
RHRM =	32.0023 si	n [(JDAY)(0.986	53) + 226] +	14.7795 + 155

Plot the heart rates for AGDA 105 to 300 on the grid in WORKSHEET 2.1a, demonstrating how they merge with the results of the calculations in WORKSHEET 2.1a. Continue plotting heart rates below for each of these activities through AGDA = 600 and JDAY = 21. Plotting may be continued on the back of this page in the grid provided.



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Moen, A. N. 1978. Seasonal changes in heart rates, activity, metabolism, and forage intake of white-tailed deer. J. Wildl. Manage. 42(4):715-738.

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JDAY =

UNIT 2.2: STROKE VOLUME

The stroke voume is a characteristic of ventricular systole, but it is influenced by more factors than the heart itself. The amount of blood circulating depends on three major factors: the cardiac output, the general activity of the animal, and the peripheral resistance to blood flow (Hoar 1966:168-169). The cardiac output, or blood pumped per unit time, is a function of the ventricular volume, the force of the contraction, and the total blood volume in the body. The amount of blood in the heart, up to the limit of ventricular volume, is affected by the amount of venous blood returned. Muscular exercise results in an increase in the venous blood returned to the heart because of the pumping effects on the veins.

Ventricular contraction does not result in complete emptying of the ventricles; varying amounts of blood remain after each contraction. The force of ventricular contraction is limited by the muscle tissue itself, but modified up to the maximum possible by the amount of blood in the heart--it contracts more forcibly if it is full--and the effect of the hormone adrenaline, which causes an increase in the strength of cardiac muscle contraction (Schmidt-Nielson 1979;103). Total blood volume obviously affects the amount of blood in the heart and the subsequent force of the contraction.

The control of stroke volume or cardiac output is self-regulated due to the feedback effects of the return of venous blood to the heart, by hormonal regulation as adrenalin affects the strength of the contraction, and by the blood volume and venous system, all acting within the physical framework of heart size. Undisturbed animals in different activities likely have stroke volumes that are proportional to heart rates. Disturbed animals, responding to noises, by elevating heart rate, for example, likely have stroke volumes that are not in proportion to rate. Their heart rate increases are not accompanied by concommitant increases in stroke volume and metabolism is not elevated in direct proportion to heart rate increases. This is an important consideration when using heart rate as an independent variable when calculating energy metabolism.

Heart volume and heart weights in relation to body weights that were discussed in CHAPTER 1 are good starting points for further analyses of the effects of these functional effects on the realized stroke volumes in different cirumstances.

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Schmidt-Nielson, K. 1979. Animal physiology: adaptation and environment. Cambridge University, Press, Cambridge. 560 pp.

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STROKE VOLUME

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ZEJAA 4---4 171 177 ceel [regula of blood pressure] jaczewski,z; janu 1958

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JEBPA 12--4 347 349 alal rata, card comp emot stres roshchevskii,mp/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR APSSA 396-- 396 96 rata blood circulation, finnish hirvonen,1; jar/ 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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UNIT 2.3: BLOOD VOLUMES

Blood volumes are dependent on the mass of the animal and on the time of year. Larger animals are expected to contain more blood than smaller animals, and higher blood volumes are observed during seasons of the year when metabolic functions are highest.

Total blood volume includes of both the plasma, or fluid portion, and blood cells which are suspended in the plasma. Red cells make up a greater portion of the packed cell volume than white cells.

Blood plasma volumes are regulated by the exchange of water through the capillary wall. Two factors are given by Pantelouris (1967:252) as the cause of such exchange. One, water tends to move by osmosis from the tissue spaces (interstitial fluid) into the capillaries because of the higher protein content within the capillaries, and two, the greater hydrostatic pressure inside the blood vessels tends to force water out, counteracting the movement of water in by osmosis. Thus total blood volume is maintained properly in the healthy animal by fluid exchange through the walls of the capillary network.

LITERATURE CITED

Pantelouris, L. M. 1967. Introduction to animal physiology and physiological genetics. Pergamon Press, Oxford. 497 pp.

REFERENCES, UNIT 2.3

BLOOD VOLUMES

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS AUTHORS	YEAR
CBCPA	192	471	473	odvi	red cell life span, w-t de noyes,wd; kitchen	1966
CPSCA	74	217	218	odvi	organ:body weight relation robinson,pf	1966
JOMAA JOMAA	311 494	5 749	17 754	odvi odvi	weight relations, georg re hamerstrom,fm,jr/ hematologica volumes, mich johnson,he; youa/	1950 1968
VJSCA	262	61	61	odvi	immobi drug, pack cell vol wesson, ja III; s	1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JANSA 38--6 1331 1332 odhe blood compon, seas, wt, fe o'brien, jm; les/ 1974 JOMAA 36--3 474 476 odhe erythrocyte val, mule deer browman, 1g; sear 1955 JOMAA 52--3 628 630 odhe tiss, organs, tota body ma hakonson, te; whi 1971 JOMAA 53--2 384 387 odhe total serum protein in pop anderson, ae; med/ 1972 JWMAA 34--2 389 406 odhe erythrocyte, leukocy, colo anderson, ae; med/ 1970 PCZOA 2--10 46 46 odhe chang, plas lipid thr year stewart, sf; nor/ 1974 WLMOA 39--- 1 122 odhe carcas, bone, organ, gland anderson, ae; med/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR ATRLA 15--- 253 268 ceel relat age and size, poland dziecioloski,r 1970 ZEJAA 4---4 171 177 ceel [regulation of blood pres] jaczewski,z; ja/ 1958

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 27--1 90 91 alal weights of minnesota moose brechinridge,wj 1946 JOMAA 50--4 826 alal blood chemis, shiras moose houston,db 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR APSSA 396-- 96 96 rata blood circulation, finnish hirvonen,1; jarv/ 1973 CJZOA 47--4 557 562 rata hematologi studies, bar-gr mcewan,eh 1968 CJZOA 50--1 107 116 rata seas chang, blood vol, wat cameron,rd; luick 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR RSPYA 21-3 365 370 anam doga, cardiopulmonary para mckean,t; walker, 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

bibi

 CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR

 ovca

 CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR

 ovda

 CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR

 ovda

 CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR

 ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

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UNIT 2.4: BLOOD CHARACTERISTICS

Blood characteristics may be divided into two major categories: hematology and blood chemistry. Hematology involves analyses of the cellular components of the blood. Blood chemistry involves analyses of the chemical components of the blood. Hematology is a physical approach, blood-chemical determinations a chemical one.

HEMATOLOGY

Counting the cellular elements in the blood of animals is valuable in assessing conditions of health and disease (Siegmund and Eaton 1967:1461). They point out, however, that variations between animals of the same species (they are discussing domestic animals) are considerable and depend on age, sex, nutrition, diurnal and sexual cycles, and stresses such as strenous exercise and excessive heat and cold. Since these variations are very much part of the lives of wild ruminants in their natural habitats, large variations should be expected in this group. Further, collections cannot be made conveniently, regularly, and without immediate stress on the free-ranging ruminant, so a very incomplete hematology picture can be obtained at best. Captive animals provide some base-line values and patterns, of course, but their relationship to free-ranging ruminants will also be variable.

A "normal hemogram" table is given in Siegmund and Eaton (1967:1462) for the ox, sheep, and goat. Categories include hematocrit, erythrocytes, hemoglobin, leukocytes, neutrophils, and lymphocytes.

Hematocrit is the packed cell volume, expressed as a percent of the total blood volume. It is determined by centrifuging the blood sample. The red blood cell or erythrocyte count is made as a check for anemia. Malnutrition, iron deficiency, or chronic disease may result in a reduction in the number of circulating red blood cells. The hemoglobin content of each red blood cell is also variable; a reduction indicates an anemic condition.

The leukocyte count usually rises in response to an acute bacterial infrection, and declines in response to acute viral infections. Neutrophils increase markedly in response to pus-forming organisms, and an increase in lymphocytes usually indicates a more chronic process, or the end stage of an acute infection. Additional details and instructions for analyzing these blood characteristics are given in Siegmund and Eaton (1967).

BLOOD CHEMISTRY

Blood chemical determinations may be made for such things as blood sugar, blood urea nitrogen, blood creatinine, and minerals such as calcium and phosphorous. Uniform handling and collection procedures were suggested by Seal et al. (1972a) as white-tailed deer showed effects of handling (with no drugs) for up to 24 hours later. Comparisons were made for restraint and drug groups, and another paper by the same authors describes nutritional effects on thyroid activity and blood of white-tailed deer (1972b). One important result is that serum thyroxine decreases in the winter, indicating a state of hypothyrodism and decrease metabolic rate as an energy conservation adaptation. This is discussed further in CHAPTER 7.

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- Siegmund, O. H. and L. G. Eaton, Ed. 1967. The Merck Veterinary Manual. Merck & Co., Inc., Rahway, N. J. 1686 pp.

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BLOOD CHARACTERISTICS

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR

aubo	lefe	phpa 807	veteri hematology, 3rd ed	schalm,ow; jaina/	1975
aubo	lefe	phpa 1287	clinical hematology	wintrobe,mm	1967
edbo	coup	itny 1463	duke's physiol of dom anim	swenson,mj	1970

REFERENCES, UNIT 2.4

BLOOD CHARACTERISTICS.

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ZOOLA 27--4 17 23 arti qauntita serolog relations baier,jg jr; wolf 1942 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBPAB 23--1 149 157 cerv serum proteins, transferri nadler,cf; hughe/ 1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR CNJMA 31-12 317 319 od-- erythr infec, splenec, tex kuttler,kl; robi/ 1967 JWMAA 39--2 346 354 od-- blood protn chng, gestatio hartsook,ew; whe/ 1975 NATUA 187-- 333 334 od-- sickling phenomenon in dee undritz,e; betke/ 1960 PSEBA 34--5 738 739 od-- sickle cell anemia in deer o'roke,ec 1936 PSEBA 117-1 276 280 od-- sickling, heteroge, hemogl weisberger,as 1964

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ABBIA 127-- 711 717 odvi hemoglobin heterogeneity huisman,thj; doz/ 1968 ABBIA 151-2 540 548 odvi struct, hemogl alph chains harris,mj; wilso/ 1972 AJPHA 199-- 190 192 odvi tactoid forma, hemoglobin moon,jh 1960

odvi continued on the next page

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJVRA 30--1 143 148 odvi serum prot, normal, arthri sikes,d; hayes,f/ 1969 AJVRA 33-12 2545 2549 odvi blood seru, arthrit reumat sikes,d; kistner/ 1972 AKASA 30--- 50 51 odvi electroph pattern, 2 subsp jackman,gs; garne 1976 ANYAA 241-- 653 odvi dome, embryo, fetal hemoglo kitchen, h; brett, 1974 671 BLOOA 29--6 867 877 odvi hemoglobin polymorphism in kitchen, h; putna/ 1967 BUCDA 29--2 105 odvi geogr dist, hemoglo compon harris mjw 105 1971 CBPAB 30--4 695 713 odvi hemat, bloo chem prot poly seal, us; erickson 1969 CBPAB 58a-4 387 391 odvi short-term chan, corti, in bubenik,ga; bube/ 1977 CBPAB 62a-4 869 odvi circannl rhythm, estradiol bubenik,ga; buke/ 1979 872 CJCMA 34--1 66 71 odvi ser blochem, hemat par cap tumbleson, me; cu/ 1970 CJZOA 44--4 631 647 odvi odhe var, blood ser, elect van tets,p; cowan 1966 CJZOA 53--6 679 685 odvi amb temp eff, physic trait holter, jb; urban/ 1975 CJZOA 57--4 777 780 odvi death, sampling, blood mea wesson, ja; scanl/ 1979 CNJMA 34--1 66 odvi serum biochem, hemat fawns tumbleson, me; cu/ 1970 71 CNVJA 14-12 299 300 odvi chloramphenicol in blood sisodia, cs; gupt/ 1973 JBCHA 247-- 7320 7324 odvi heterog, hemogl alpha chai taylor, wj; easle/ 1972 JICRB 9---3 179 193 odvi seas, body cond. plas vol jacobsen, nk 1978 JOMAA 31--1 5 17 odvi weight relations, georg re hamerstrom, fm, jr/ 1950 JOMAA 39--2 269 274 odvi blood composition of w-t d teeri,ae; vircho/ 1958 JOMAA 39--2 309 311 odvi aspects of blood chemistry wilbur, cg: robin/ 1958 JOMAA 41--3 410 411 odvi vitamins a and e in blood haugen, ao; hove, e 1960 JOMAA 49--4 749 754 odvi hematologica volumes, mich johnson, he; youa/ 1968 JOMAA 54--1 270 274 odvi geograph dist, hemoglo var harris, mj; huism/ 1973 JOPAA 59--6 1091 1098 odvi hematol chan, fawns, ticks barker, rw; hoch,/ 1973 JWIDA 9---4 342 348 odvi combin etorphine, xylazine presidente, pja;/ 1973 JWIDA 10--- 18 24 odvi blood char, free-rang, tex white,m; cook,rs 1974 JWMAA 3---1 14 16 odvi studies on the blood of wt whitlock, sc 1939 JWMAA 29--1 79 84 odvi comp milk, blood, doe, fawn youatt, wg; verme/ 1965 JWMAA 29--4 717 723 odvi natural var blood proteins miller, wj; hauge/ 1965 JWMAA 31--4 679 685 odvi protein requirement, fawns ullrey, de; youat/ 1967 JWMAA 34--4 1034 1040 odvi effec immobil on blood ana seal, us; ozoga, j/ 1972 JWMAA 36--4 1041 1052 odvi nutrition, thyroid, blood seal,us; verme,1/ 1971 JWMAA 38--4 845 847 odvi restrain appar, blood samp mautz, ww davis/ 1974

odvi continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHOR S----- YEAR

JWMAA JWMAA JWMAA JWMAA JWMAA	392 392 394 403 432	342 346 692 442 454	345 354 698 446 460	odvi odvi odvi odvi odvi	ser cholesterol level chng blood prot, gestatn, suckl ener, prot, blood urea nit plasma progest, puber, faw season, nutri, serum nitro	<pre>coblenz,be hartsook,ew; whe/ kirkpatrick,rl;b/ abler,wa; buckla/ bahnak,br; holla/</pre>	1975 1975 1975 1976 1976 1979
MGQPA	32	113	138	odvi	physiol baselines, hematol	karns,pd	1972
NAWTA	3	890	892	odvi	glac nat par, enlar spleen	aiton,jf	19 <u>3</u> 8
PAARA	209	1	11	odvi	feed restr, seas, antl dev	long,ta; cowan,r/	1959
SCIEA	144	1237	1239	odvi	hemoglob polymor, sickling	kitchen,h; putn/	1964
SOVEA	281	25	26	odvi	blood, glucose, urolog par	hoff,gl; trainer,	1975
WDABB	31	32	34	odvi	serolog surv, 2 herds n y	friend,j; halter	1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 34--5 477 484 odhe age, nutrition, blood chem kitts,wd; bandy/ 1956 CJZOA 35--2 283 289 odhe age, nutrition, blood chem bandy, pj; kitt/ 1957 CJZOA 47--5 1021 1024 odhe observa haematology, races cowan, imct: band 1969 JANSA 33--1 244 244 odhe plasma mineral indexes, nev rohwer, g1; lesp/ 1971 JANSA 34--5 896 896 odhe lipid, plas comp lev, neva lesperance, al; / 1972 JANSA 36--6 1201 1201 odhe wt, plasma minrl indx, fem lesperance,al; h/ 1973 JANSA 38--6 1331 1332 odhe blood compon, seas, wt, fe o'brien, jm; les/ 1974 JANSA 41--1 273 273 odhe effect temp on blood compo obrien, jm; alldr/ 1975 JOMAA 36--3 474 476 odhe erythr valu, alim canal ph browman, 1g; sears 1955 JOMAA 52--3 628 630 odhe tiss, organs, tota body ma hakonson, te; whic 1971 JOMAA 53--2 384 387 odhe total serum prot, populati anderson, ae; med/ 1972 190 odhe blood serum electrol, colo anderson, ae; med 1972 JWIDA 8---2 183 JWMAA 34--2 389 406 odhe erythrocytes, leukocy, col anderson, ae: med/ 1970 NAWTA 17--- 482 496 odhe rela hematology to conditi rosen, mn; bischof 1952 NAWTA 33--- 359 364 odhe blood parasites, texas dee robinson, rm; kut/ 1968 WDABB 4---3 78 odhe seriol surv, arbovir infec emmons, rw 80 1968 WLMOA 39--- 1 122 odhe carcas, bone, organ, gland anderson, ae; med/ 1974

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR CBPAB 43a-3 649 653 ceel blood chemis roosevelt elk weber,yb; bliss, 1972 JOMAA 49--4 762 764 ceel physiologic stud, rocky mt herin,ra 1968 JWMAA 39--3 617 620 ceel blood chemistry & hematolo pedersen,rj; pede 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR ANYAA 97--1 296 305 alal stud on blood, serum group braend,m 1962 CJZOA 53-10 1424 1426 alal serum corticoid, handl str franzmann,aw; fl/ 1975 HEREA 85--2 157 162 alal var, red cell enzy, scandi ryman,n; beckma/ 1977 JOMAA 51--2 403 405 alal charact, captive mich moos verme,lj 1970 JZAMD 8---1 27 37 alal serial bl chem, hemat franzmann,aw; bai 1977 NCANA 101-- 263 290 alal cerv, blood chem, nutritio leresche,re; sea/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ANIPA 22--1 105 114 rata effect hndlng, blood comp hyvarinen, h; hel/ 1976 BLUTA 32--6 439 442 rata hematol val, finnish reind timisjarvi j; re/ 1976 BRHLA 9---2 105 114 rata cryorheology of rein blood halikas,gc 1972 CBPAB 41a-2 437 438 rata fibrinogen concentration i halikas,gc; bower 1972 CBPAB 55a-2 187 193 rata serum enz, calves, nut eff bjarghov, rs; fje/ 1976 CBPAB 60a-4 383 rata seas chng ser levels, ions lund-larsen, tr / 1978 386 CJZOA 44--2 235 240 rata electroly, red cells, plas manery, jf; barlo/ 1966 CJZOA 46--5 1031 1036 rata hematologi studies, bar-gr mcewan,eh 1968 CJZOA 47--4 557 rata chan, blood const with age mcewan, eg; white/ 1969 562 CJZOA 50--1 107 116 rata seas chang, blood vol, wat cameron, rd; luick 1972 CNJMA 24--5 150 152 rata hematol val, barren gr car gibbs, hc 1960 GNKAA 12--1 56 65 rata [blood ser, gene poly tra] zhurkevich, nm; fo 1976 JOMAA 33--1 105 105 rata notes on lipids, wild mamm wilber,cg 1952 LBASA 21--6 817 824 rata biomedical research, reind dieterich, ra; lui 1971 ZEBFA 9---4 341 345 rata [hemoglobin types, ontoge] irzhak, li 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 55--2 448 455 anam hematol val, adult free-ra barrett, mw; chalm 1977 CJZOA 55--8 1252 1260 anam clinicochem val, adu free- barrett, mw: chalm 1977 RSPYA 21--3 297 306 anam resp functions, mamm blood dhindsa,ds; metc/ 1974 RSPYA 21--3 365 370 anam doga, cardiopulmonary para mckean,t; walker, 1974 WAEBA 575-- 1 anam pronghorn antelope carcass field, ra; smith, / 1972 6 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ABBGB 9---3 175 179 bibi doca, isozyme chr, cell cu carleer, j; pasto/ 1978 bibi two hemoglobin phenotypes harris,mj; wils/ 1973 BIGEB 9---1 1 11 CJZOA 57--9 1778 1784 bibi comp blood char plain, woo peden,dg; kraay,g 1979 EVOLA 12--1 102 110 bibi blood groups in amer bison owen,rd; stormon/ 1958 bibi doca, red cells, carb anhy sartore,g; storm/ 1969 GENTA 61--4 823 831 JWIDA 11--1 97 1975 100 bibi hematol, blood chem values marler, rj JWIDA 12--1 7 13 bibi hematol valu, 5 areas, u s mehrer, cf 1976 JWIDA 14--4 493 bibi serolog, hematol val, colo keith, eo; ellis,/ 1978 500 RSPYA 30--3 305 310 bibi blood respirato properties haines, h; chiche/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBPAB 40b-2 567 570 ovca ovca, ovmu transfe, hemogl nadler, cf; woolf/ 1971 CJZOA 51--5 479 482 ovca stress, lymphocyte stimula hudson, rj 1973 JAVWA 157-5 647 ovca physiol valu, cap, handlng franzmann, aw; tho 1970 650 JRMGA 25--4 292 1972 296 ovca serum phospho valu, forage hebert,dm JWIDA 6---1 67 68 ovca hematologic values, captiv woolf, a; kradel, d 1970 JWIDA 7---2 105 ovca physiol values, capti, wil franzmann, aw 1971 108 JWMAA 36--3 924 932 ovca envir sour, varia, phy val franzmann, aw 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovđa

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR FEPRA 35--3 480 480 oram ovca, angiotensi, conv enz zakheim,r; matti/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 39--4 554 559 obmo serologica evid, relations moody, pa 1958

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JANSA 35--6 1271 1274 dosh eff wat restric nutrnt dig asplund, jm; pfand 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 37--4 584 585 serol tech, ident blo prot tempelis,ch; rod/ 1973 JWMAA 40--3 517 522 ungl id hemoglobins, law enforc bunch,td; meadows 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBPAB 51a-1 21 26 ceni oxyg trnsport, phosphoryla ochiai,t; encki,y 1975

TOPIC 3. RESPIRATORY SYSTEM FUNCTIONS

Respiration includes both the physical exchange of oxygen and carbon dioxide between the lung surfaces and the atmosphere, called external respiration, and the chemical exchange of O_2 and CO_2 in cellular metabolism, called internal respiration. External respiration rates are rather easily measured, and changes in the gaseous composition of inhaled and exhaled air indicate the amounts of physical exchange of oxygen and carbon dioxide that has taken place. External respiration also includes changes in the vapor pressure of respired air which is part of the overall heat exchange and thermal energy balance of an organism. Internal respiration is necessary for tissue metabolism, and exothermic metabolic reactions are also involved in the thermal energy balance of an organism.

External respiration involves physical work, which can be accomplished only with the support of internal respiration and muscle contraction. Thus external and internal respiration are both essential and continuous functions for the maintenance of life itself, and both are part of the overall energy metabolism and heat exchange of organisms.

UNIT 3.1: EXTERNAL RESPIRATION

The mechanical movement of air from the atmosphere into the lungs and back to the atmosphere requires the expenditure of energy by the muscles involved in external respiration. The cost of breathing has been determined for man, and is 1.2% of the total oxygen consumption when at rest (Otis 1954). As activity and external respiration increase, the cost may double, and in very heavy respiration, may reach about four times the cost at rest, or nearly 5% of the total oxygen consumed. Margaria et al. (1960) determined a maximum of 3% of the total cost during exercise.

Respiration rates of wild ruminants are largely unknown. Generally younger animals have faster respiration rates than older animals of the same species. Larger animals, both of the same and different species, tend to have slower respiration rates than smaller ones. Respiration rates increase with increases in the amount of physical activity as more oxygen is needed to support higher rates of metabolism. The respiratory frequency of reindeer in lying posture was 23 and in standing posture 36 per minute (White and Yousef 1978).

Respiration rates are regulated by the CO_2 concentration in the blood; as the CO_2 concentration increases, the rate and volume of external respiration increases. It is interesting that, while increased activity results in an increased need for oxygen, it is not the need for oxygen but rather the increased CO_2 in the blood from increased internal resiration that stimulates pulmonary ventilation. This can be demonstated by increasing the CO_2 concentration of inhaled air, and observing an increase in the respiration rate and volume even without an actual increased energy expenditure.

A few measurements of external respiration rates have been made, usually as part of metabolism studies, and may be used to formulate a general pattern of changes in relation to age, size, and activity. Changes also occur with season as metabolic changes occur; measurements of external pulmonary ventilation rates should be made over the entire annual cycle. Superimposed on these predictable changes are the effects of transient stimuli. These changes occur in relation to fright responses, apprehension, etc. In general, transient changes in external respiration are expected to be related to changes in activity and metabolism due to overt responses.

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- Otis, A. B. 1954. The work of breathing. Physiol. Rev. 34:449-458.
- White, R. G. and M. K. Yousef. 1978. Energy expenditure of reindeer walking of roads and on tundra. Canadian J. Zool. 56(2):215-223.

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EXTERNAL RESPIRATION

SERIALS

CODEN	vo-nu	BEPA	ENPA A	ANIM	KEY	WORDS			- AUTHORS		YEAR
JAPYA	153	354	358		mect	na work,	breath,	mus er	margaria,r;	mili/	1960
PHREA	343	449	458		the	work of	breathi	ng	otis,ab		1954

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odvi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR CBCPA 61A-- 43 48 ceel oxygen util by elk calves robbins,ct; cohe/ 1977 JOMAA 49-4 762 764 ceel physiolo studies, rocky mt herin,ra 1968 JWMAA 43--2 445 453 ceel energy expenditure, calves robbins,ct; cohe/ 1979 RSPYA 29--2 225 230 ceel select oxygen transp param mckean,t; staube/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR RSPYA 30--3 305 310 bibi blood respirato properties haines, h; chiche/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovca

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR

ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 12--- 349 360 bibo physiological properties janusz,g 1967

UNIT 3.2: INTERNAL RESPIRATION

Internal respiration, or the exchange of gases between the lung surfaces and the blood is based on pressure differences between the blood and the lung surfaces, and the O_2 and CO_2 is exchanged by diffusion at rates dependent on the difference in pressure between capillary blood and the gas at the respiratory surface.

Blood increases in oxygen upon exposure to respiratory surfaces. Blood contains both plasma and cells, but the solubility of O_2 is low in plasma, so nearly all of the O_2 carried by the blood is associated with the red cells. These cells contain the pigment <u>hemoglobin</u>, which is chemically composed of a protein "globin" and four "heme" groups. The ironcontaining heme groups function in oxygen transport, and oxygen molecules (4) unite with the four iron atoms to form a fully-saturated <u>oxyhemoglobin</u> molecule. This is the basis for very efficient oxygen transport in the vascular system.

Muscles are active sites of oxidative metabolism, and they contain a particular kind of hemoglobin called <u>myoglobin</u> which has a greater oxygen affinity than hemoglobin. It is likely that the pigment myoglobin facilitates oxygen diffusions to the mitochondria in muscle cells (White, p. 195 in Gordon 1977).

LITERATURE CITED

Gordon, M. S. 1977. Animal physiology: principles and adaptations. Macmillan Publishing Co., Inc., N.Y. 699 pp.

REFERENCES, UNIT 3.2

INTERNAL RESPIRATION

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odvi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR RSPYA 29--2 225 230 ceel select oxygen transp param mckean,t; staube/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

1

alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR RSPYA 30--3 305 310 bibi blood respirato properties haines, h; chiche/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram
TOPIC 4. REPRODUCTIVE SYSTEM FUNCTIONS

Free-ranging animals exhibit definite reproductive cycles through each year of their lives. First, reproductive maturity is reached, which is dependent on both age and weight. Animals on poor range are expected to weigh les, and reproductive maturity is reached later. After reproductive maturity is reached, the period of breeding is regulated by the effects of light on the hormone balance, and the subsequent gestation and lactation periods are fixed in relation to the time of conception.

Spermatogenesis and ovulation are important functions of individual animals. The extent of differences in sperm and egg production between individual males and females, respectively, is not known because individuals cannot be evaluated under free-ranging conditions. A very high percentage of each sex is likely to be capable of breeding in wild ruminant populations.

After breeding, the male can be primarily concerned with maintenance while the female must meet the demands of gestation, parturition and lactation. These functions occupy about three-fourths of the year or more for the females, and include responsibilities not only for the production of a new individual but also must meet the high metaboliac cost of lactation and provide some measure of protection and behavioral security for the young. Further, the abundance of animals in a population depends on the successful completion of reproductive functions over successive annual cycles.

REFERENCES, TOPIC 4

REPRODUCTIVE SYSTEM FUNCTIONS

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR

aubo meth loen 321 mamm ecol of reprod in wild, do sadleir, rmfs 1969 edbo caun nyny 189 ovca reproduct in mammal, bok 6 austun, cr short, 1976

UNIT 4.1: REPRODUCTIVE CYCLES

Reproduction by wild ruminants does not occur randomly throughout the year, but is part of a biological chronology characteristic of each species and, in some cases, the locations of populations. Males are non-productive for much of the year; sperm are not produced by the inactive testes. Females are non-reproductive for only a short time each year; they are pregnant or lactating most of the time. The timing of the reproductive and non-reproductive periods through the year is quite fixed, with not too much variation between years. This is apparently due to the association between metabolic demands for reproduction and the availability of forage resources on a seasonal basis.

The male reproductive cycle is often represented by the amount of testicular development. White-tailed deer in Texas showed the lowest testicular weights in February-May (Robinson et al. 1965). Pronghorn testes weights (Montana) were at an annual low in January and February (O'Gara et al. 1971). Maximum testes weights and sperm production occur in late summer in pronghorn, resulting in the breeding of pronghorn females earlier than the breeding of deer. Pronghorn have a gestation period of 240 days compared to 200 days for deer, so the earlier breeding results in parturition at about the same time for the two species. The time of breeding is an important consideration in describing the biochronology of a species because breeding commits the females to defined periods of gestation The timing of these productive periods is an important and lactation. consideration in relation to the phenology of the range, especially the last one-fourth of gestation and the first half of lactation because these are such critical periods in the life of both dam and offspring. Later fall breeding combined with a longer gestation period would result in too short a growing period for the young prior to the beginning of their first winter. The importance of the phenology of the range in relation to timing of the arrival of spring was discussed and illustrated in Moen (1978).

Peak breeding of deer in Texas occured in December, about one month later than in New York, which Robinson et al. (1965) consider a normal variation due to latitude effects. Differences in breeding times from north to south are not very large.

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REPRODUCTIVE CYCLES

BOOKS

TYPEPUBLCITYPGESANIMKEYWORDS------AUTHORS/EDITORS--YEARaubocoupitny670mammpatternsmammalianreproducasdell, sa1964

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CAFGA 34--1 25 31 od-- br seas, produc, interstat chattin, je 1948

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJVRA 39--6 1053 1056 odvi antl, long bone mass, andr brown,rd; cowan,/ 1978 BIREB 16--3 340 343 odvi repro ster, fema seas chan plotka,ed; seal,/ 1977 odvi repr, proges, estrog, preg plotka, ed; seal, / 1977 BIREB 17--1 78 83 CJZOA 44--1 59 62 odvi breeding seasons, manitoba ransom, ab 1966 odvi seas var LH,FSH, tes, male mirarchi,re; how/ 1978 CJZOA 56--1 121 127 ENDOA 94--4 1034 1040 odvi annual testos rhyth, adult mcmillin, jm; sea/ 1974 JANSA 31--1 225 225 odvi sperm reserves of w-t deer lambiase, jt, jr; a 1970 JANSA 40--1 185 186 odvi andr lev, antl cy, bree se mirarchi, re; sca/ 1975 JANSA 42--1 271 272 odvi seas var, gonad char, male mirarchi, re; sca/ 1976 JOMAA 32--4 411 421 odvi analys reproduc pat, s tex iilege,d 1951 JOMAA 40--1 108 1959 113 odvi breed record, capt, alabam haugen, ao JOMAA 47--2 266 odvi endocrine glands, seas chan hoffman, ra; robin 1966 280 JOMAA 53--4 760 773 odvi ovar comp, repr phys, venez brokx, pa 1972 JRPFA 47--1 161 163 odvi chan estro, estradi, pregn harder, jd; woolf1 1976 JWMAA 10--3 249 odvi breeding season, new york cheatum, el; morto 1946 263 JWMAA 14--3 290 295 odvi breed rec, upper pen, mich haugen, ao; davenp 1950 JWMAA 28--1 171 173 odvi birth of white-tai d fawns michael, ed 1964 JWMAA 29--1 53 59 odvi reproduc cycle, male texas robinson, rm; tho/ 1965 JWMAA 29--1 74 79 odvi reproducti studies, penned verme, 1j 1965

odvi continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 30--4 843 845 odvi regional diff, fawning tim weber,aj 1966 JWMAA 31--1 114 123 odvi reprodtive biolo, manitoba ransom, ab 1967 JWMAA 33--3 708 odvi fertility, male w-t fawns follmann, eh; klim 1969 711 JWMAA 33--4 881 odvi repro pattern, nutri plane verme, lj 887 1969 JWMAA 36--3 868 875 odvi reproductive physiol, male lambiase, jt, jr; / 1972 JWMAA 38--2 183 196 odvi eff diethylstilbes, reprod harder, jd; peterl 1974 odvi initia, preg, lactating de scanlon, pf; murp/ 1976 JWMAA 40--2 373 374 JWMAA 40--4 792 795 odvi noneff, mechan birth contr matschke, gh 1976 JWMAA 41--1 87 91 odvi diethylstilbestr, contrace matschke,gh 1977 JWMAA 41--1 92 99 odvi ann chang, sperm prod org mirarchi, re; sca/ 1977 JWMAA 41--2 178 183 odvi androg levels, antl develo mirarchi, re; sca/ 1977 JWMAA 41--2 194 196 odvi antifertil act, syn proges matschke,gh 1977 JWMAA 41--4 715 719 odvi fact aff peak fawning, vir mcginnes, bs; down 1977 NFGJA 16--2 261 261 odvi twin fawns born 2 days apa hesselton, wt; van 1969 NFGJA 20--1 40 47 odvi breedi, parturit dates, ny jackson, lw; hesse 1973 PCGFA 9---- 128 odvi birth dates of alabama dee lueth, fx 1955 131 PCGFA 20--- 123 130 odvi breeding seas in louisiana roberson, jh, jr; d 1966 PCGFA 20--- 130 139 odvi delin of rut, breedin seas payne, rl; provos/ 1966 PCFGA 29--- 646 odvi oral accep, eff, diethylst matschke,gh 651 1975 PCGFA 32--- 335 338 odvi fawning dates, manag impli butts,gl; harmel/ 1978 POASA 56--- 24 25 odvi breeding season, e oklahom dunbar,mr 1976 TJSCA 26... 417 420 odvi breeding season in s texas harwell, wf; barro 1975 TNWSD 27--- 19 38 odvi photoperiodism, breeding mcdowell, rd 1970 VJSCA 24--3 112 112 odvi spermatozoan reserves in d lenker, dk; scanlo 1973 VJSCA 26--2 59 odvi plas androg lev, repro char mirarchi, re; sca/ 1975 59 VJSCA 26--2 60 60 odvi seas. age dif, male org wt russell, md: wess/ 1975 VJSCA 27--2 46 46 odvi plas progest lev, estr cyc kirkpatrick, r1; / 1976 WLSBA 3---4 152 156 odvi hormon implan, contr repro bell,rl; peterle, 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR APARD 1976- 208 215 odhe environ eff on reproductio sadleir,rmfs 1976 CAFGA 43--1 91 96 odhe breedin seas, herds, calif bischcoff,ai 1957 CJZOA 54-10 1617 1636 odhe horm reg, repro, antl cycl west,no; nordan,h 1976 CJZOA 54-10 1637 1656 odhe eff methallibure,hormon tr west,no; nordan,h 1976

odhe continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 54--1 302 303 odhe reproductio, b-t deer fawn thomas,dc; smith, 1973 JRPFA 44--2 261 odhe reprod pattern, female b-t thomas, dc; cowan, 1975 272 69 JWIDA 7---1 67 odhe bilateral testicular degen murphy, bd; clugst 1971 JWIDA 11--1 101 odhe testicular atrophy, calif demartini, jc; con 1975 106 JWMAA 14--4 457 469 odhe bree seas, prod, faw, utah robinette,w1; gas 1950 odhe preg fawn, quintupl mule d nellis, ch; prent/ 1976 JWMAA 40--4 795 796

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BEHAA 16--- 84 ceel charact of estrous, captiv morrison, ja 1960 92 JOENA 53-PR 48 49 ceel thyr calci, seas repro cha phillippo,m; lin/ 1972 JOMAA 36--1 145 145 ceel fetus in yearling cow elk saunders, jk jr 1955 JOMAA 47--2 332 334 ceel fetus resorption in elk haugen, ao 1966 JOMAA 51--4 812 ceel precoci antl dev, sex matu moran, rj **1970** 813 JRPFA 25--1 41 54 1971 ceel puberty, seas breedin male lincoln,ga JRPFA 27--3 427 438 ceel female reprodu cycl, scotl guiness, f; linco/ 1971 JWMAA 16--3 313 315 ceel age at sex maturity, male conoway, cf 1952 JWMAA 17--2 223 223 ceel pregnant yearling cow elk coffin, al; reming 1953 JWMAA 23--1 27 34 ceel breed seas, known-age embr morrison, ja: tra/ 1959 JWMAA 32--2 368 ceel exper studies, contr repro greer, kr; hawkin/ 1968 376 JZ00A 163-1 105 123 ceel seas reprodu changes, stag lincoln,ga 1971 JZ00A 172-3 363 ceel timing, reproduc, latitude fletcher, tj 367 1974 JZ00A 185-1 105 114 ceel calving times, red d, scot guiness, fe; gibs/ 1978 MAMLA 35--2 204 219 ceel ruru, season, births, n z caughley,g 1971 NAWTA 21--- 545 554 ceel postconcept ovulation, elk halazon, gc; buech 1956 NATUA 248-- 616 618 ceel restor lib, cast stag, est fletcher, tj; shor 1974 ZEJAA 1---1 69 75 ceel caca, dada [time of birth] rieck,w 1955 ceel 1 sided testicle shrinkage wurster,k; hofma/ 1975 ZEJAA 21--4 238 242

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARAVSPA 14--1 8191alal morph, ultrastr, spermatoz andersen,k1973JOMAA 37--2 300300alal late breeding, moose, alce moisan,g1956JWMAA 26--4 360365alal in gravelly, snowcrest mou peek,jm1962VILTA 6---3 1299alal reproductio, moose, sweden markgren,g1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CAFNA 90--4 449 463 rate annual antler cycle, newfo bergerud, at 1976 CJZOA 50--1 43 46 rata reprodu, female reind, car mcewan, eh; whiteh 1972 CJZOA 53--9 1213 1221 rata repro seas, carib, newfoun bergerud, at 1975 CJZOA 56--8 1684 1696 rata morphol, b-g caribou ovary dauphine,tc,jr 1978 rata antl shedd, parturi, reind espmark, y 1971 JWMAA 35--1 175 177 JWMAA 38--1 54 66 rata synchronous mating, b gr c dauphine, tc, jr; m 1974 NCANA 97--1 61 rata calving dates, carib, queb desmeules, p; sim 1970 66 PASCC 22--- 17 17 rata repro patter, reind, carib mcewan, eh; whiteh 1971 ZOLZA 46-12 1837 1841 rata [reproduc, wild, taimyr p] michurin, ln 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR WMBAA 16--- 17 23 bibi biol, manage, national par fuller, wa 1962

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 46--5 899 944 ovca social, physical maturatio geist,v 1968

SWNAA 22--- 153 ovca minimum breeding age, utah mccutchen, he 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR FUNAA 24... 96 100 obmo early matur, fecund, norwa alendal,e 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 19--4 417 429 oram two-year study, crazy moun lentfer, jw 1955

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JRPFA 21--1 1 8 dada reproductive cycle, male chapman, di; chapm 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR ATRLA 1---- 333 376 bibo reproduc of european bison jaczewski,z 1958 ATRLA 12--- 333 334 bibo breeding, zool garden, pol landowski,j; woli 1967 ATRLA 12--- 407 444 bibo reprod biol, reserve, free krasinski,z; racz 1967 ZSAEA 33--4 193 214 many [developm antlers, reprod] lau,d 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 52--3 537 544 anam ann test cycle, horn casti ogara, bw; moy, rf/ 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JRPFA 12--2 337 351 caca sexual cycle seas breed ma short, rv; mann,t 1966

CHAPTER 6, WORKSHEET 4.1a

Testes development as percent of body weight in male white-tailed (odvi) through the annual cycle

The reproductive data in Robinson et al. (1965) on male deer collected through the year are ideal for representation as a sine wave through the annual cycle. The circle diagram shown in the Figure 2 in the published paper illustrates a pattern throughout the year that can be easily converted to a symmetric sine wave, with the maximum occurring in November and the minimum in May. Thus only a primary phase correction (see CHAPTER 1) is necessary.

Complete the following table using data given by Robinson and adding the columns indicated by the underscore. The first line is an example.

Deer Number	Date Collected	Age (years)	JDAY	Total Weight (pounds)	LWKG	Testes Weight (gms)	Testes Percent of total Weight
N-4	9-22-62	3.5	264	88	40	31.5	0.078
N-97	_	-		_	_	_	_
				_	_	_	· _
_	-	_	_	· _	_	_	. —
-				_		_	_
_		·		_	_	<u> </u>	
-	_	_				·	-
_	_	_		_		· · <u> </u>	
	_ •	_		_			
	_	-					_

Rewrite the procedures described in CHAPTER 1 for deriving sine wave equations with a primary phase correction, and derive an equation for testes weight as a percent of live weight (TPLW). Plot the results on the graph on the next page. The primary phase correction is:

The equation is:



LITERATURE CITED

Robinson, R. M., J. W. Thomas, and R. G. Marburger. 1965. The reproductive cycle of male white-tailed deer in central Texas. J. Wildl. Manage. 29(1):53-59.

UNIT 4.2: SPERMATOGENESIS AND OVULATION

Testes are compact organs with specialized cells for the production of spermatozoa. Testicular weights, epididymal weights, testicular spermatozoa numbers, and epididymal spermatozoa numbers were all highly caculated with plasma endrogen levels in white-tailed deer (Mirachi et al. 1976). These reproductive characteristics of the male, are apparently related to the light regime, and coincide with the reproductive condition of the female, of course.

When spermatogenesis occurs, the spermatozoa arise from specialized sex cells called spermatogonia. They are released and move by way of the epididymis and vas deferens to the ejaculatory duct, from which they are expelled at the time of mating. Spermatogenesis occurs almost exclusively during the period of lowest body temperature (McCauley 1971:395). The testes of wild ruminants are suspended in the scrotum, which has a lower temperature than the abdominal cavity.

Ovulation occurs as undeveloped sex cells, or ova, mature and are released from the ovary, enter the fallopian tube and move through it to the uterus. If sperm are present, the ovum may be fertilized and then implanted in the uterine wall where it develops.

The female enters a period of estrus, or "heat," when ovulation occurs and she is ready to receive the male. Estrus occurs again in 3-4 weeks if conception has not occurred. The frequency of unsuccessful matings during the first estrus is not known for wild ruminants because of their secretiveness and daily behavior patterns. The general impression one gets from reproductive data on wild ruminants is that they reproduce very successfully and consistently when range conditions are favorable and herd health is good.

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Mirachi, R. E., P. F. Scanlon, and R. L. Kirkpatrick. 1976. Seasonal variation in gonadal characteristics of male white-tailed deer. J. Anim. Sci. 42(1):271-272.

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SPERMATOGENESIS AND OVULATION

BOOKS

TYPEPUBLCITYPGESANIMKEYWORDS------AUTHORS/EDITORS--YEARaubocoupitny670mammpatternsmammalianreproduasdell,sa1964

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS AUTHORS		YEAR
AJANA	132-2	189	2 05	odvi	ultrastr corpus lute, preg sinha,aa se	a1,u/	1971
BIREB	171	78	83	odvi	repr, proges, estrog, preg plotka ed; s	ea1,/	1977
CIRIB	62	1053	1056	odvi	collec semen. electro ejac bierschwal, c	j: m/	1 9 68
COVEA	393	282	291	odvi	corpora lutea, ovul incide cheatum,el		1949
JANSA	311	225	225	odvi	sperm reserves of w-t deer lambiase, jt,	jr; a	1970
JANSA	421	271	272	odvi	seas var, gonad char, male mirarchi,re;	sca/	1976
JAVMA	157-5	627	632	odvi	char semen coll, elec ejac bierschwal,c	j; m/	1970
JOMAA	323	267	280	odvi	notes on fecunidty, maine palmer,rs		1951
JOMAA	401	108	113	odvi	breed record, capt, alabam haugen, ao		1959
JOMAA	482	321	321	odvi	polyovulation in wt-tailed hesselton.wt		1967
JOMAA	534	760	773	odvi	ovar comp, repr phys, venez brokx, pa		1972
JWMAA	121	78	86	odvi	produc, yield, george rsrv o'roke,ec; h	amers	1948
JWMAA	152	73	80	odvi	produc, mortalit, coralled severinghaus	.cw	1951
JWMAA	291	74	79	odvi	reproducti studies. penned verme.li	,	1965
JWMAA	293	487	492	odvi	corpora lutea variation of trauger.dl:	hauge	1965
JWMAA	311	114	123	odvi	reprodtive biolo, manitoba ransom, ab	0	1967
JWMAA	333	708	711	odvi	fertility, male w-t fawns follmann.eh;	klim	1969
JWMAA	334	881	887	odvi	repro pattern, nutri plane verme, lj		1969
JWMAA	352	369	374	odvi	accessory corp lute, ovari mansell, wd		1971
JWMAA	363	868	875	odvi	reproductive physiol, male lambiase, jt,	jr; /	1972
JWMAA	382	183	19 6	odvi	eff diethylstilbestrol rep harder, jd; p	eterl	1974
JWMAA	411	92	99	odvi	ann chang, sperm prod, org mirarchi, re;	sca/	1977
JWMAA	412	87	91	odvi	microencapsulatd diethylst matschke,gh		1977
JWMAA	412	194	196	odvi	antifertil act, syn proges matschke,gh		1977
JWMAA	414	731	735	odvi	ferti control, steroi impl matschke, gh		1977
NFGJA	111	13	27	odvi	product, growth, adirondac severinghau	s,cw;	1964

odvi continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR PCFGA 29--- 646 651 odvi oral accep, eff, diethylst matschke,gh 1975 VJSCA 24--3 112 112 odvi ovula, pregnan, lactat dee scanlon, pf; murp/ 1973 V.JSCA 24--3 112 112 odvi spermatozoan reserves in d lenker, dk; scanlo 1973 VJSCA 27--2 46 46 odvi plas progest lev, estr cyc kirkpatrick,rl; / 1976 WLSBA 3---4 152 156 odvi hormon implan, contr repro bell,rl; peterle, 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR APARD 1976- 208 215 odhe environ eff on reproductio sadleir, rmfs 1976 CAFGA 44--3 253 259 odhe productiv, herds californi bischoff, ai 1958 JOMAA 54--1 302 odhe reproductio, b-t deer fawn thomas, dc; smith, 1973 303 JWIDA 7---1 67 69 odhe bilateral testicular degen murphy, bd; clugst 1971 JWIDA 11--1 101 odhe testicular atrophy, calif demartini, jc; con 1975 106 JWMAA 19--1 115 136 odhe fertility, utah mule deer robinette, w1; ga/ 1955 JWMAA 19--4 503 odhe rocky mount, high repr rte jensen,w; robinet 1955 503 JWMAA 21--1 62 65 odhe ovarian anal, reprod perfo golley, fb 1957 MRLTA 50--1 12 12 odhe record, multiple ovulation fowle, ke 1969 NAWTA 9---- 156 161 odhe productivity, central utah robinette,wl; ols 1944 NAWTA 15--- 589 596 odhe productiv, mule d, colorad tolman, cd 1950 THGNB 3---3 101 106 odhe early pubert, female b-t d mueller,cc; sadle 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR 92 BEHAA 16 84 1960 ceel charact of estrus, captive morrison, ja CIRIB 8---4 994 997 ceel freezi red d semen, polan jaczewski,z; mor/ 1976 JOMAA 47--1 152 153 ceel occ of acces corpora lutea douglas, mjw 1966 JRPFA 25--1 41 54 ceel puberty, seas breedin male lincoln,ga 1971 JRPFA 27--3 427 438 ceel female reprodu cycl, scotl guiness, f; linco/ 1971 1953 JWMAA 17--2 177 184 ceel reproduction, yellowstone kittans, wh JWMAA 24--3 297 307 ceel ovarian char, breed histor morrison, ja 1960 JWMAA 32--2 368 376 ceel exper studies, contr repro greer, kr; hawkin/ 1968

ceel continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JZ00A 163-1 105 123 ceel seas reprodu changes, stag lincoln,ga 1971 NAWTA 20--- 560 567 ceel increas natal, lowered pop buechner, hk; swan 1955 NAWTA 21--- 545 ceel postconcept ovulation, elk halazon,gc; buech 1956 554 ZEJAA 4---3 105 130 ceel [reproductive phenomena] valentincic, si 1958 ZEJAA 21--4 238 242 ceel 1 sided testicle shrinkage wurster,k; hofma/ 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AVSPA 14--1 81 91 alal morph, ultrastr, spermatoz andersen,k 1973 JWMAA 22--3 261 268 alal reproductn in a population edwards, ry; ritce 1958 JWMAA 23--4 381 401 alal reprod & produc, newfoundl pimlott, dh 1959 alal reproductio, moose, sweden markgren,g VILTA 6---3 1 299 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 56--8 1684 1696 rata morphol, b-g caribou ovary dauphine,tc,jr 1978 JOMAA 52--2 479 479 rata twinning in caribou mcewan,eh 1971 JOMAA 54--3 781 781 rata twinning in reindeer, nwt nowosad, rf 1973 JWMAA 38--1 54 66 rata synchronous mating, b gr c dauphine, tc, jr; m 1974 JZ00A 164-4 419 424 rata collec, exam, reinde semen dott, hm; utsi, mnp 1971 JZ00A 170-4 505 508 rata artific insemina, reindeer dott, hm; utsi, mnp 1973 PASCC 22--- 17 rata repro patter, reind, carib mcewan, eh; whiteh 1971 17

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

anam

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 9---2 155 156 ovca non-breeding in bighorn sh pulling, avs 1945 JWMAA 30--1 207 209 ovca twinning in bighorn sheep spalding, dj 1966 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR FUNAA 24... 96 100 obmo early matur, fecund, norwa alendal,e 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 19--4 417 429 oram two-year study, crazy moun lentfer,jw 1955

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR ATRLA 1.... 333 376 bibo reproduc of european bison jaczewski,z 1958 ATRLA 12--- 407 444 bibo reprod biol, reserve, free krasinski,z racz 1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AMNTA 114-1 101 116 ungu maternal reproductv effort robbins, ct; robbi 1979

UNIT 4.3 GESTATION AND PARTURITION

Gestation occurs when the fertilized ovum, called a zygote, is implanted on the endometrium of the uterus. The zygote develops, becoming an embryo, and extraembryonic tissues develop to become the placenta. The placental membranes function in the exchange of nutrients, gases, and wastes between the fetal and maternal circulatory systems.

Fetal development is relatively slow in the first one-third, increases some in the second one-third, and increases rapidly in the final one-third of pregnancy; growth rates were discussed in CHAPTER 1.

Parturition in wild ruminants seems to be a rather routine event that is accomplished rather quickly, based on the few observations of parturition reported in the literature. Most wild ruminants tend to give birth at times and places that make observation difficult.

Breeding as a result of natural selection does not result in the physical mismatches that can occur through selective breeding of domestic ruminants. A large-breed ram and a small-breed ewe, for example, may result in difficulties during parturition due to fetal size. Difficulties at birth are probably quite infrequent in natural populations, although actual rates of mortality during parturition are unknown.

REFERENCES, UNIT 4.3

GESTATION AND PARTURITION

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CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS AUTHORS Y	EAR
JOANA	941	1	33	cerv	aspects of placentation hamilton,wj; har/ 1	.960
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS Y	'EAR
AJANA AJANA AJANA	126-2 127-4 132-2	201 369 189	241 395 205	odvi odvi odvi	morphogen, fetal membranes sinha,aa; seal,u/ 1 ultrastr amnion, amni plaq sinha,aa; seal,u/ 1 ultrastr corpus lute, preg sinha,aa; seal,u/ 1	.969 .970 .971
BIREB	171	78	83	odvi	repr, proges, estrog, preg plotka,ed; seal,/ 1	.977
CNJNA	542	259	259	odvi	doca, cotyled attach, uter scanlon, pf 1	974

odvi continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JRPFA 47--1 161 163 odvi chan estro, estradi, pregn harder, jd; woolf1 1976 JWIDA 9---4 356 358 odvi multipl anomali, w-t fetus wobeser,g; runge, 1973 odvi congen anom, neonat, alber barrett, mw; chalm 1975 JWIDA 11--4 497 501 JWMAA 26--4 409 411 odvi x-ray in determi pregnancy verme, 1k; fay, 1d/ 1962 JWMAA 27--1 142 143 odvi technique for preser uteri haugen, ao 1963 JWMAA 28--1 171 odvi birth of white-tai d fawns michael, ed 173 1964 JWMAA 29--1 53 odvi reproduc cycle, male texas robinson, rm; tho/ 1965 59 JWMAA 29--3 487 492 odvi corpora lutea variation of trauger, dl; hauge 1965 JWMAA 30--2 414 417 odvi sexing embryo by chromatin segelquist, ca 1966 JWMAA 31--1 114 odvi reprodtive biolo, manitoba ransom, ag 123 1967 JWMAA 33--4 881 887 odvi repro pattern, nutri plane verme, 1j 1969 JWMAA 37--3 423 odvi support dev, field laparot scanlon, pf; lenke 1973 424 JWMAA 39--4 684 691 odvi uterin comp, growth, pregn robbins, ct; moen, 1975 JWMAA 40--2 373 374 odvi initia, preg, lactating de scanlon, pf; murp/ 1976 NFGJA 16--2 261 261 odvi twin fawns born 2 days apa hesselton, wt; van 1969 NFGJA 18--1 42 51 odvi reprod anomali, female, ny hesselton,wt; jac 1971 OJSCA 78-AP 14 14 odvi possibl superfeta, spontan lamvermeyer, bl; m 1978 PIAIA 71--- 241 247 odvi struc, cervic regi, uterus morris, je 1964 VJSCA 23--3 116 116 odvi a laparotomy technique w-t scanlon, pf; lenke 1972 VJSCA 23--3 116 116 odvi aspects of early pregnancy scanlon, pf; murp/ 1972 odvi ovula, pregnan, lactat dee scanlon, pf; murp/ 1973 VJSCA 24--3 112 112

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ACATA 84--1 118 128 odhe anam, morpho, cervix uteri kanagawa,h; hafez 1973 ANREA 122-- 335 340 odhe quadruplets in mule deer sears, hs browman 1955 CJZOA 48--1 123 132 odhe odvi, develo, fetal period ommundsen, p; cowa 1970 odhe gesta per, breed, fawn beh golley,fb JOMAA 38--1 116 120 1957 JWMAA 37--3 312 326 odhe effect nutr chan on captiv robinette, w1; ba/ 1973 odhe preg fawn, quintupl mule d nellis, ch; prent/ 1976 JWMAA 40--4 795 796

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 47--6 1418 1419 ceel sexual dimorphi in fetuses retfalvi,1 1969 JOMAA 36--1 145 145 ceel fetus in yearling cow elk saunders, jk jr 1955 JOMAA 47--2 332 ceel fetus resorption in elk haugen, ao 1966 334 JWMAA 17--2 177 ceel reproduction, yellowstone kittans, wh 184 1953 JWMAA 17--2 223 223 ceel pregnant yearling cow elk coffin, al; reming 1953 JWMAA 23--1 27 34 ceel breed seas, known-age embr morrison, ja; tra/ 1959 JWMAA 31--1 145 149 ceel determ preg, rectal palpat greer, kr; hawkins 1967 JWMAA 40--2 330 ceel nutri, hesta, succes repro thorne, et dean/ 1976 335 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 26--4 360 365 alal in gravelly, snowcrest mou peek,jm 1962

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR CAFNA 90--4 498 499 rata twin fetuses, woodland car shoesmith,mw 1976 JOMAA 49--4 778 778 rata placental remnants, rumens miller,fl: parker 1968 JWMAA 25--2 205 205 rata sex determination of calve bergerud,at 1961 JWMAA 28--3 477 480 rata field meth, parturiti rate bergerud,at 1964

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 30--1 207 209 ovca twinning in bighorn sheep spalding,dj 1966 JWMAA 43--4 970 973 ovca deter pregncy, progestin, ramsay,ma; sadlei 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS-				AUTHORS	YEAR
FUNAA	24	101	103	obmo	gest	tation	period	of	muskox	alendal,e	1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARJWMAA 19--4 417 429 oram two-year study, crazy moun lentfer, jw1955

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBCPA 43A-- 673 679 mamm gestation period, body wt kihlstrom, je 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AMNTA 114-1 101 116 ungu matern repro effort, fetal robbins,ct; robbi 1979

UNIT 4.4: LACTATION

The production of milk is a distinguishing feature of mammals. It is a very necessary process as the neonates are not able to either, forage or digest forage; milk is the sole supply of nutrients immediately after birth. Further, colostrum, the first milk produced after parturition is highly nutritious and contains more dry matter, more protein, and more vitamins than milk produced a few days after parturition and through the rest of the lactation period. The colostrum of ruminants also contain antibodies essential for the young until they develop an active immunity to diseases. The low resistance due to a lack of antibodies at birth in ruminants is due to the lack of placental transmission from mother to fetus. The effects of these important mother-young relationships in the first few days of life may be realized in population dynamics as changing environmental conditions affect the successful establishment of both the nutritional and social relationships.

The amount of milk produced usually increases for a few weeks after parturition and then slowly decreases. The increase coincides with the increased demand for nutrients by the rapidly growing young before the rumen is developed well enough to be fully functional.

Milk is released in response to the nursing activities of the young. They may suckle several times a day, with a decline in the frequency of nursing in the last two-thirds of lactation. This decline coincides with increasing dependence on forage and decreasing milk production. Milk production ceases when the young no longer stimulate the dam by nursing, usually 3-5 months after parturition in wild ruminants.

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TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR

aubo isup amia 291 doca secretion of milk, 4th ed. esped,w; smith, vr 1952 phpa 584 handbook of biologica data spector, ws, ed. edbo saco 1956 aubo coup itny 670 mamm patterns mammalian reprodu asdell, sa 1964 edbo nyny 516 mamm mammary gland, dev & maint larson, bl; smith, 1974 acpr mamm biosyn & secr milk; disease larson, bl; smith, 1974 edbo acpr nyny 458 edbo nyny 425 mamm nutrit and biochem of milk larson, bl; smith, 1974 acpr

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 39--2 355 360 odvi milk consumpt, weight gain robbins, ct; moen, 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARJWMAA 44--2 472 478 odhe milk yield, black-tailed d sadleir, rmfs1980

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JRPFA 37--1 67 84 ceel comp, yield, milk, red dee arman,p; kay,rnb/ 1974 JWMAA 40--2 330 335 ceel nutri, gesta, succes repro thorne,et; dean,/ 1976 SZSLA 41--- 297 312 ceel physiol effe lacta on moth hanwell,a; peake 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR ALLKA 55 23 28 alal struc rib, thorax indic mil anghi,c 1968 JWIDA 12--2 202 207 alal milk, hair, element relati franzmann, aw; f1/ 1976 PNSUA 27--2 129 138 alal compar nut in preg, lactat payne, pr; wheele/ 1968 VILTA 6---3 1 299 alal reproductio, moose, sweden markgren,g 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 49--4 443 447 rata measu, milk intake, calves mcewan,eh; whiteh 1971 CJZOA 54--1 55 64 rata glucos metab, lactat reind white,rg; luick,j 1976 SZSLA 41--- 297 312 rata physiol effe lacta on moth hanwell,a; peake 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOANA 94--1 1 33 cerv aspects of placentation hamilton, wj; har/ 1960 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR b1b1 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS---- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

1977

EVOLA 31--1 177 199 mamm signif lacta, evol of mamm pond, cm

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TOPIC 5. EXCRETORY SYSTEM FUNCTIONS

Excretion of unused forage components and waste products of metabolism is an important function since regularity is necessary to avoid build-up of potentially toxic substances.

Urine and feces are the two main kinds of materials excreted from the body of wild ruminants. Gas production also occurs, and waste materials are sloughed off the skin and hair. These waste products should be considered when evaluating the metabolic costs of maintenance; they are discussed in later chapters.

Urine and feces are discharged directly from the excretory system in measurable quantities that can be related to water and forage intake. The difference between ingestion and excretion represents some level of metabolic efficiency. There are many details to consider when evaluating efficiencies, however, since the components of excretory products come from both ingestion and metabolic sources.

Few research studies have been completed in the excretory functions of wild ruminants. The basic functions behind urine and feces excretion are discussed in UNITS 5.1 and 5.2. Some of the more practical uses of fecal excretion characteristics is in estimating populations, but this is discussed in CHAPTER 19.

UNIT 5.1: URINARY FUNCTIONS

Water and salt balances are maintained as a result of the actions of the kidneys in regulating the amount and concentration of the urine. It is the only organ capable of regulating the movements of water and salts between the environmental medium and the fluid compartments of the body (McCauley 1971:158). Water loss also occurs in the feces, but this is quite constant, and through the skin. Neither of these two pathways are capable of regulating water balance, however, fecal water losses are fairly constant, and the losses through the skin vary in relation to thermal energy balance rather than water balance. Thus dehydration occurs when temperatures are excessive and water losses through the skin high.

The functional unit in the kidney is the nephron. It is richly supplied with blood, and differences in blood pressure within the glomerulus result in fluid, called glomerular filtrate, collecting in the cavity of Bowman's capsule. Reabsorption by active transport then occurs, resulting in the reclaiming of useful materials such as sugars and amino acids, and waste products are allowed to pass into the rumen. These and other details of renal function are described in McCauley (1971:153-168). A schematic drawing of kidney function is shown below.



Bowman's capsule

Note that usea may be reabsorbed rather than eliminated by the ruminant, resulting in the conservation of nitrogen as an aid in protein metabolism when the range is low in protein.

The recycling of urea is an example of resource conservation by free-ranging animals, illustrating how the resource economy may be maintained over time, thereby maintaining a higher level of productivity than would be the case if resource transactions were made on a short-term basis only.

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URINARY FUNCTIONS

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 155-7 1085 1085 odhe urine-collect device, male richmond,m; pill/ 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR NZJSA 13--4 663 668 ceel kidney wt, kidne fat index batcheler,cl; cl/ 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BIJOA 155-3 549 566 alal ceel, chymotrypsin, pancre lindsay,rm; steve 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 39--2 379 386 rata kidney wt fluct, fat index dauphine,tc,jr 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ANREA 169-2 343 343 ovca observ kidney, desert bigh horst,r; langwort 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JANSA 35--6 1271 1274 dosh eff wat restric nutrat dig asplund, jm; pfand 1972

UNIT 5.2: FECAL FUNCTIONS

The feces of ruminants contain water, undigested forage residues, cells that have been abraded from the gastrointestinal tract, bacteria, products of rumen fermentation, and other products of physiological processes such as bile acids, pigments, mucin, and inorganic salts. Large amounts of feces are defecated by ruminants because so much of the forage ingested is not digested. The more lignified the forage material is, the less it can be digested. Fecal samples may be used as a technique for estimating dietary composition by identifying fragments.

The water content of ruminant feces varies seasonally. Succulent spring forage is high in water content, and feces have little or no form. At other times, especially in the winter, the feces are quite dry and, in many species, formed into small pellets which are defected as pellet groups.

The excretion of rather dry pellet groups, composed largely of indigestible forage, has been used by biologists as a field method for estimating populations. If a known member of pellet groups are defecated per animal each day, then the number of pellet groups divided by the number defecated per day is an estimate of the number of animal-days of use. Such estimates may not be very precise due to sampling errors in counting pellet groups in the field and variations in the number of pellet groups defecated per day. This technique is discussed further in CHAPTER 19.

REFERENCES, UNIT 5.2

FECAL FUNCTIONS

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
JWMAA	333	506	510	ođ	qual ident forage remnants	zyznar,e; urness,	1969
					х. - С С С С С С С С		
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
JRMGA	301	61	63	odvi	chrom oxid indic fecal out	ruggiero, 1f; whel	1977
JRMGA	322	93	97	odvi	infl brush control on diet	quinton,da; hore/	1979
JWMAA	261	50	55	odvi	rain, count of pellet grou	wallmo,oc; jacks/	1962
VJSCA	233	116	116	odvi	dosh chrmicoxide fecal out	sanders,ot skee/	1972
CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
JRMGA	302	116	118	odhe	food, wld hors, doca, colo	hansen,rm; clark/	1977
				odhe	continued on the next page		

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR 444 1964 JWMAA 28--3 435 odhe defecation rates of mule d smith, ad JWMAA 31--1 190 191 odhe anam, id fecal gr, pH anal howard, vw, jr 1967 JWMAA 32--4 961 962 odhe fecal ph values, dom sheep nagy, jg; gilbert, 1968 JWMAA 43--2 563 564 odhe number pellets per defecat strong, 11; freddy 1979 JWMAA 32--4 961 962 odhe fecal ph values, dom sheep nagy jg; gilbert, 1968 SWNAA 13--2 159 166 odhe food plants, habitat, okla clark, tw 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 29--2 406 407 ceel determinat defecation rate neff,dj; wallmo,/ 1965 JWMAA 41--1 76 80 ceel foods of ungulates, colora hansen,rm; clark, 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 40--2 374 375 alal dail wint pell gr, bed, al franzmann,aw; ar/ 1976 NCANA 95--5 1153 1157 alal [numb pellet-gro each day] desmeules,p 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

rata

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR GRBNA 38--2 222 224 anam sim bet prong, odhe fec pe johnson,mk; maccr 1978 JRMGA 32--4 275 279 anam fec, rum, util meths, diet smith,ad; shandru 1979 JRMGA 32--5 365 368 anam livest, foods, dese steppe johnson,mk 1979 JWMAA 40--3 469 478 anam diets, forag avail, colora schwartz,cc; nagy 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 42--3 581 590 bibi diet, s1 rvr herd, nw terr reynolds, hw; han/ 1978 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 37--3 363 366 ovca food hab, plant frag, fece todd, jw; hansen,r 1973 JWMAA 39--1 108 111 ovca food of, southern colorado todd, jw 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CWOPA 35--1 1 19 obmo rata, diets, canadi arctic parker,gr 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEARJZOOA 185-- 270273 dosh caca, comparison wint diet henry, bam1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 43--3 796 798 ungu fecal ph compar, 3 species peek,jm; keay,ja 1979

TOPIC 6. SKELETAL SYSTEM FUNCTIONS

The skeleton of vertebrates provides support for the entire body, more or less rigid places for the attachment of muscles, and protection for some of the vital organs. Joints or articulations are present in two main kinds: immovable joints, as in the skull, and movable ones, as in the limbs. The skeleton plus the musculature provides the basic form of the animal's body which results in animals of grace and beauty, attractive to nearly all persons.

The skeleton has important seasonal functions as it is a storage reservoir for animals for mobilization during periods of rapid antler growth Skeletal remains are also useful for determining mortality of sex and age groups.

UNIT 6.1: THE SKELETON AS SUPPORT

One of the main functions of the skeleton is body support, which is attained as a result of the rigidity of the bones and the muscle tone and contractions that result in various postures. Skeletal support charcteristics are of importance when evaluating details of locomotion, and the anatomical characteristics that affect relationships between animal and range, such as the effects of snow and forage distributions and availability. Support functions themselves are not analyzed further in this UNIT; anatomical measurements were discussed in CHAPTER 1, and the effects of range conditions on animals in CHAPTER 17.

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THE SKELETON AS SUPPORT

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CATRB 12--4 323 330 cerv cyclic bone remodeling dee hillman,jr; davi/ 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 50--2 302 310 odvi alal, stuctur adapta, snow kelsall, jp 1969

CODEN	vo-nu	BEPA	ENPA	ANIM	EY WORDS			AUTHORS	YEAR
CJZOA	414	629	636	odhe	ge determ,	ossif,	long bo	lewall,ef; cowan,	1963
CAFGA CAFGA	414 421	327 15	346 21	odhe odhe	osh, dogo, ivi, pelv g	skeleta girdl, r	l diffe el, sex	hildebrand,m taber,rd	1955 1956
JOMAA	452	226	235	odhe	ang-rel gro	o dif, s	k ratio	klein,dr	1964

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 37--1 129 129 ceel healing, fractured leg bon gilbert, pf; hill, 1956 JWMAA 30--2 369 374 ceel bone char assoc with aging gilbert, pf; hill, 1956

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 50--2 302 310 alal odvi,structur adapta, snow kelsall,jp 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEARATICA 19--2 111113rata functio brow tine, caribou pruitt,wo1967UABPA 18--- 141rata mechanics, energy, crateri thing,h1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR AMNTA 113-- 103 122 mamm scal. skel mass, body mass prange,hd; ander/ 1979
UNIT 6.2: THE SKELETON AS A STORAGE RESERVOIR

The skeleton serves as an important storage reservoir of nutrients that can be mobilized when nutrient intake is not sufficient to meet current metabolic demands. Two particularly good examples of this situation are at the time of antler growth and when energy demands are higher than metabolizable energy intake in the diet.

Minerals. The accumulation of minerals in the skeletal system for mobilization during antler growth is probably necessary for such rapid rates to be possible. Phosphorous and calcium are mobilized from the ribs and long bones for deposition in the antlers of white-tailed deer and this basic physiological process is very likely characteristic of all of the ruminants that shed their antlers each year.

Marrow. Bone tissue is very much alive and active, though the calcified tissue is rigid and often encloses more active, spongy tissue. The marrow inside of the shafts of the long bones contains blood at all times, and a rather high fat content when range conditioNs are good enough to result in a positive energy balance by the animal. The fat contents of the femur is often used as an indicator of the physical condition of an animal, and its stability permits one to estimate the season of death when dead animals are found in the spring. Animals not found during the hunt can be distinguished from those that died as a result of poor nutrition in the winter.

REFERENCES, UNIT 6.2

THE SKELETON AS A STORAGE RESERVOIR

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JWMAA JWMAA	94 292	319 397	322 398	odvi odvi	symptoms of malnutrition harris,d kidney, marrow fat, condit ransom,ab	1945 1965
NFGJA	211	67	72	odvi	physical condition of whit monson, ra; stone/	1974
NYCOA	35	19	22	odvi	bone marrw index of malnut cheatum,el	1949
PCGFA	26	57	68	odvi	var fat levl, mandib cavit nichols,rg; pelt	1972
PSEBA	129	733	737	odvi	calcium strontium age antl cowan, rl; hartso/	1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 45--2 252 259 odhe density studies, body fat whicker, fw 1964 JOMAA 52--3 628 630 odhe tiss, organ, tota body mas hakonson, te; whic 1971 JWMAA 36--2 579 594 odhe indice, carc fat, colo pop anderson, ae; med/ 1972 JWMAA 41--1 81 86 odhe exp starva, recovery, does decalesta,ds; na/ 1977 WAEBA 589-- 1 6 odhe the mule deer carcass field, ra; smith, / 1973 WLMOA 39--- 1 122 odhe carcas, bone, organ, gland anderson, ae; med/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS---- YEAR JSFAA 22--1 29 33 ceel fatty-acid compos, adipose garton, ga; duncan 1971 JWMAA 30--1 135 140 ceel measurements, weight relat blood, da; lovaas, 1966 JWMAA 32--4 747 751 ceel fat content, femur marrow greer, kr 1968 MAMLA 35--3 369 383 ceel demog, fat res, bod sz, nz caughley,g 1971 WAEBA 594-- 1 8 ceel the elk carcass field, ra; smith, / 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AZOFA 12--2 148 155 alal fatty-acid compos, org fat tanhuanpaa,e; pul 1975 JWMAA 40--2 336 339 alal marrow fat, mortali, alask franzmann,aw; arn 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBCPA 30--1 187 191 rata fatty acid comp bone marow meng, ms; west, gc/ 1969 CBPAB 56--3 337 341 rata liver, bone, bone marrow bjarghov, rs; jac/ 1977 FUNAA 23... 106 107 rata fat deposits, svalbard dee oritsland, na 1971 JWMAA 34--4 904 907 rata wt, dried marrow femur fat neilans,ka 1970 NJZOA 24--4 407 417 rata morph, fat stor, org weigh krog, j; wika, m; / 1976 PASCC 22--- 14 14 rata water flux, climate, nutri cameron, rd; luic/ 1971 TNWSD 28--- 91 108 rata phys var, condit, b g cari dauphine, tc, jr 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JANSA 33--1 309 309 anam carbonyl analysis of fat booren,a; field,/ 1971 JDSCA 38--- 1344 1344 anam major chemi compos, bovine reid,jt; wellin/ 1955 JOMAA 56--3 583 589 anam seas tren in fat lev, colo bear,gd 1971 WAEBA 575-- 1 6 anam pronghorn antelope carcass field,ra; smith,/ 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 26--2 305 308 bibi the lipids in bison bison wilbur,dg; gorski 1955

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBCPA 50b-4 599 601 ovda fatty acid comp bone marrw west,gc; shaw,dl 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 49--8 1159 1162 many comp, adipose tiss triglyc garton,ga; dunca/ 1971

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TOPIC 7. MUSCULAR SYSTEM FUNCTIONS

Movements of vertebrate animals are made possible by the contraction of the muscles spanning skeletal articulations. Speedy locomotion is a very important attribute of wild ruminants as they often depend on short bursts of speed to escape danger.

Muscular contraction is an exothermic process, resulting in the release of heat energy. The heat energy must be dissipated or else the temperature of the body tissue will rise. Dissipation can become a problem on warm autumn days when the winter coat has grown enough to provide good insulation. Then, heat energy cannot escape efficiently, and warm autumn days can be thermally stressful for animals that must run as the emerging winter coat provides good insulation from heat loss. I have observed white-tailed deer with tongues out for cooling on several occasions under such conditions.

Muscles can also be a source of heat production without body movement; shivering is a form of thermogenesis that may be important for animals in extreme cold. Thus muscles function in more ways than for locomotion. Free-ranging animals, well adapted for their nautral environments, integrate all of their life functions into a way of being, a continuum in space and time, that is best understood by studying the animal as a whole in relation to the functions of its different parts or systems.

UNIT 7.1: LOCOMOTION

Locomotion occurs when muscles spanning skeletal articulations contract. The details of locomotion have been studied for several domestic and some wild species. The mechanics of locomotion by race horses, for example, is currently being studied in Sweden and at Cornell University, USA. Details of locomotion have not been critically evaluated for North American wild ruminants however.

One of the major characteristics of locomotion that has a distinct practical value is the mechanics of weight-loading when an animal is moving. Static weight loads, determined by relating body mass to hoof area, are of less value than dynamic weight loads when different species of animals are being compared, or different activities such as walking or running. More research in this rather specialized area of investigation would greatly increase our understanding of the effects of snow on animals in the northern regions.

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LOCOMOTION

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR UABPA 18--- 1 41 rata behav, mechan energ, crate thing h 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovca

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEARSCAMA 202-5 148 157 mamm how animals runhildebrand, 1960

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UNIT 7.2: THERMOGENESIS

Thermogenesis is another term for heat production. It is a term applied to increases in heat production as a response to cold, implying that thermogenesis is a specific response in addition to the heat production that is a continuous characteristic of life.

Voluntary activity is a thermogenic response, and may be employed by ruminants to "keep warm." Severinghaus and Cheatum (1956:146) describe cases where white-tailed deer in the Adirondacks were observed to be walking slowly along heavily-used trails at midnight when temperatures approached -35°C, presumably to keep warm by excercising. While heat production increases will result from increased movement, it is not a good long-term adaptation because of the energy costs incurred at a time when food resources are declining in quality and may not be readily accessible, while body reserves are decreasing.

Muscular thermogenesis may occur as a result of shivering, without an increase in activity. Shivering is an involuntary muscular response, which includes an initial increase in muscle tone followed by several muscular tremors per second. Andersson (1970:1125-6) states that shivering may increase oxygen consumption by 400 percent, and that it is more effective from a thermodynamic point of view than voluntary muscle contractions. Thus thermogenics may be an important part of an animals' response to cold, but a difficult one to interpret in the field.

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

UNIT 7.3: INVOLUNTARY MUSCULAR CONTRACTIONS

Involuntary muscular contractions occur continuously as a part of the maintenance of life. The heart beats without voluntary control, the diaphragm contracts and external respiration occurs, the rumen and rest of the gastrointestinal tract experience involuntary contractions that keep food materials moving, and there are other involuntary contractions of muscles associated with different system functions. All of these muscular contractions cost the animal something, but the costs are a part of maintenance. There is essentially no literature on this subject for wild ruminants, except that in references describing the functions of various body systems.

REFERENCES, UNIT 7.3

INVOLUNTARY MUSCULAR CONTRACTIONS

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY	WORDS	AUTHORS	YEAR
				anam				
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS	AUTHORS	YEAR
				bibi				
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS	AUTHORS	YEAR
				ovca				
CODEN	VO-NU	BEPA	ENPA	ANIM	KEY	WORDS	AUTHORS	YEAR
				ovda				
CODEN	VO-NII	BEPA	ένρα	ANTM	KEY	WORDS	AUTHORS	YEAR
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CODEN	vo-nu	BEPA	ENPA	ANIM	КЕҮ	WORDS	AUTHORS	YEAR
				oram		-		

TOPIC 8. NERVOUS SYSTEM FUNCTIONS

The nervous system is a most elaborate and intricate control system that coordinates the maintenance, activity, and productive functions of each individual. Many other systems interact with the nervous system. The muscular system, for example, is coordinated to a high degree by the nervous system, resulting in smooth and graceful body movements. The endocrine system functions with the nervous system to a large extent. Klemm (1970:849) points out that the endocrine system is not only controlled by the nervous system but hormones also act on the nervous system in a feedback way.

The overall function of the nervous system is described by Klemm (1970:851) in a simplified way as an input \rightarrow output system mediated by the processing of the information, with the additional possibility of retention of some of the information in the memory. The functions can be illustrated like this.



Inputs may be processed in the brain before output responses are made. These are called voluntary responses. Some responses are made before the central nervous system has processed the input information. An animal may be startled, run a short distance without processing the reasons for running, and then stop to process the information provided by the input. The source of the input may be of no consequence--a branch may fall and scare a deer--or it may be of considerable importance to the individual--a twig may be snapped by an approaching predator. When incidents are processed and consistent biological responses formulated, the animal is exhibiting a memory that may be drawn on for later processing. Such information becomes part of the "historical environment" discussed in CHAPTER 3 of PART II.

Some responses do not include the nervous system; sensory receptors that signal distention of the rumen are part of the continuous system that causes the output--rumenoreticular contractions--to occur without conscious response by the organism. These are involuntary responses, and are part of the maintenance processes of life. The sensory capabilities of an animal are of interest to ecologists because these functions link animal to environment. These functions are commonly referred to as the senses, and include sight, hearing, taste, touch, smell, thermal senses, and pain. These senses alone do not define the functional environment (See CHAPTER 3 in PART II), but they establish the perceived operational environment. If an individual cannot see, for example, then none of the wavelengths of the electromagnetic spectrum usually detected visually by an animal of that species affect that individual, and its environment is altered accordingly.

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NERVOUS SYSTEM FUNCTIONS

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edbo	else	nyny 9	33 many	acoustic behavior of anima b	ousnel,rg,ed	1963
aubo	phli	nyny 2	40 many	the senses of animals m	natthews, lh knig	1963
aubo	macm	nyny 1	13 many	sensory mechanisms of	case,j	1966
aubo	hi11	loen 7	60 many	the chemical senses m	noncrieff.rw	1967
aubo	ccth	spil 2	00 many	molecular basis of odor a	amoore,je	1 9 70
aubo	tapl	nyny 1	83 many	animal senses b	ourton, r	1970
edbo	apcc	nyny 4	12 many	comm, chem sig: advan chem	johnston,jw,jr; /	1970
edbo	iucn	mosw 9	04 ungu	behav, relat to management g	geist, v, ed; walth	1971
proc	acpr	nyny 2	31 rata	ecochemi studies, reindeer b	pertmar,g	1975

UNIT 8.1: VISION

The eye, the organ of vision, is an extremely important part of the nervous system of diurnal animals. The eye of wild ruminants has but one lens, (it is a simple rather than a compound eye), and the lens focuses the image on the retina on the back of the eye where the stimuli are converted by the receptors to nerve impulses that are transmitted to the brain for processing.

Differences in visual functions of wild ruminant species have been observed but not evaluated physiologically. What structural and functional capabilities does the pronghorn have that enables it to see for great distances that other wild ruminants do not have? Comparative studies have not been done, so definitive answers to such questions are not available.

REFERENCES, UNIT 8.1

VISION

BOOKS

TYPE	PUBL	CITY	PGES	ANIM	KEY WO	RDS				AUTHORS/EDITORS	YEAR
aubo	ccth	spil	250	many	animal	visi	: what	anim	see	smythe, rh	1961
aubo	chha	loen	132	vert	vision	in	vertel	brates	3	tansley,k	1965

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

AJVRA 39--4 699 702 odvi cone, rod photo receptors witzel,da; sprin/ 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR cee1 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AVCSA 18--2 159 167 alal lens lesions, elk, sweden kronevi,t; holmb/ 1977 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR rata CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ыы CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram

UNIT 8.2: HEARING

Most ruminants detect and interpret sounds in the natural environment with a high level of capability. Further, individuals emit sounds that have particular meanings to other members of the species. Elk, for example, use "bugling" as an integral part of breeding behavior. This sound is emitted by the vocal cords. White-tailed deer emit a snorting sound when they are uncertain of the kind of stimulus being received. This sound is made by forcing air out through the nostrils vocal cords do not seem to be involved.

The organs of hearing are located in the inner ear. These organs convert vibrations of different frequencies and wave-lengths to nerve impluses that are transmitted to the brain for processing. The impulses received are interpreted according to pitch, which is dependent on the frequencies of the vibrations, and intensity, or loudness, which is dependent on the frequencies and amplitudes of the vibrations. Wild ruminants respond to differences in both of these as they process such information in the ecological context. Close but harmless sounds may cause a response, for example, while sounds that are distant but associated with potentially harmful events (distant gunshots, for example) may not cause a response.

Little is known about the actual sensitivity distributions of wild animals to different sound characteristics. Direct measurements of inner ear characteristics are difficult if not impossible to make in wild ruminants, and animals trained to respond in certain ways to different sound stimuli have not been used in wild ruminant research. Hearing capabilities must be guessed at or evaluated on the basis of limited observations made in the field.

One interesting bit of research was completed at the Wildlife Ecology Laboratory at Cornell when young whitetail fawns that were being trained for telemetry experiments were exposed to a systematic playing of recorded wolf howls (Moen et al. 1978). The fawns exhibited a patterned heart rate response to the frequencies and loudness of the howls, indicating some kind of genetically programmed response. Heart rate measurements in response to a series of snowmobile activities have also been made; the deer were very quick to detect the sound of the snowmobile. Further details are given in CHAPTER 17. Additional observations of heart rate responses to common sounds such as airplanes, tractors, and other noises of society have also been made while completing heart rate studies. Some of these transient responses are described in Moen and Chevalier (1977). The conclusion must be that deer, at least, are very sensitive to sounds and do a considerable amount of information processing without exhibiting overt behavioral responses.

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HEARING

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR BPURD 1---- 387 397 rata intrasp commu, mother-calf ericson, ca 1975 408 1975 BPURD 1---- 398 rata acoustic commun, review of lent pc BPURD 1---- 423 435 rata socializat, calving ground miller, fl; ander/ 1975 ZETIA 29--1 42 81 rata mother-young rel, ontog be espmark,y 1975 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 50--3 647 648 anam th blowsound of pronghorns waring,gh 1969 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR SWNAA 9---3 118 145 anim biol acoust, sound spectro davis, li 1964

UNIT 8.3: OLFACTION

Olfaction, or the sense of smell, appears to be an important function associated with the selection of forage by at least some of the wild ruminants, and the avoidance of predators and in the reproductive behavior of all of the wild ruminants. Olfaction is made possible by specialized epithelial cells lining the nasal cavities. These specialized cells receive olfactory stimuli, and transmit information via olfactory nerves to the olfactory lobes of the brain for processing.

The olfactory areas are larger in the keen-scented animals than in those with a lesser sense of smell. The actual mechanism of olfactory perception is subject to further measurement and debate. Kare (1970:1173) briefly discusses theories of olfaction. Similarities in molecular configurations between the stimulus and the receptor may be one explanation. Molecular vibrations of the stimulus are thought to play a part in some olfactory relationships. Solubilities of adsorbed molecules may also play a part in olfaction, as well as enzyme effects and other chemical relations at the olfactory surface. Rather than seeking a single explanation for olfaction, the different contributions of all of these possible mechanisms under different ecological situations should be considered.

An interesting similarity exists between the apparent genetically programmed responses of young white-tailed deer to wolf howls using the sense of hearing described in the previous unit (UNIT 8.2) and the results of Muller-Schwarze (1972) on the responses of young black-tailed deer to predator odors. These black-tailed deer exhibited negative responses to the droppings of both North American and African predators, even though they had had no previous encounters with such predators. Muller-Schwarze concluded that the deer possessed a genetically determined negative response to odors of predators. Such conclusions, and those described in UNIT 8.2 on hearing responses to recorded wolf howls, are of real ecological interest and potentially of great significance in management. The role of odors in the behavior and ecology of wild ruminants is apparently great, and may be even greater than we humans, with our limited olfactory capabilities, consider it to be. The scent environment is very dynamic (see CHAPTER 3), and olfaction is very important in behavioral relationships (see CHAPTERS 4 and 5).

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OLFACTION

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ceel

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alal

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

bibi

AMZOA 7---3 421 429 mamm olfaction in mammals moulton, dg 1967

UNIT 8.4: TASTE

Taste is an important sensation associated with food. It is also closely associated with the sense of smell, and the two can hardly be separated except by experimental elimination of one of these senses.

The taste receptors are called taste buds, and are found in large numbers on the tongue. Cattle have large numbers on the tip of the tongue and even larger numbers on the back of the tongue (Kare 1970:1192). This distribution may also be characteristic of wild ruminants.

Taste and/or smell apparently function in food discrimination. Preference tests have shown that black-tailed deer have distinct preferences for different solutions (Crawford and Church 1971). Forage preferences of white-tailed deer discussed in PART IV indicate that they select the more digestible forages before the less digestible ones, showing that taste and smell have effects on the diet quality.

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TASTE

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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UNIT 8.5: TOUCH

No references to studies of the sense of touch in wild ruminants have been found. The role of touch in their daily lives may be quite important, however. The tongue of such selective browsers as deer appears to have a highly developed sense of touch, with the tongue and lips having important roles in prehension.

The sense of touch is also important in mother-young relationships. The licking of the newborn by the dam and the nursing responses of the young almost within minutes after birth likely involve the sense of touch to a high degree.

The roles of touch in the lives of wild ruminants should be considered when evaluating both physical and biological relationships to their environnments. Descriptions of these roles should be made, even if there is little or no experimental evidence of the functions or importances of the roles at this time.

REFERENCES, UNIT 8.5

TOUCH

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				odvi				
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UNIT 8.6: THERMAL SENSES

Sensors that detect differences in the thermal energy at the skin surface or other sites of temperature sensitivity are important in the regulation of the thermal energy balance of free-ranging animals in their natural environments. These sensors function in thermoregulatory behavior, as well as in the internal thermogenic responses possible when heat energy must be dissipated or conserved.

The physiological functions of thermal sensors have not been reported on for wild ruminants. They presumably have a rather highly developed repertoire of possible responses as a result of thermal sensing since they are quite well-adapted to cold climates. The sensors in most direct contact with the environment must be located on the exposed skin surfaces. The nose, for example, seems to be sensitive to differences in thermal energy loads. Whitetail fawns at the Wildlife Ecology Laboratory, for example, have been observed to rest with their heads in the shade and their bodies exposed to the sun on hot days, suggesting that the head--most likely the nose--contains the sensors.

The roles of thermal sensors in relation to the transfer of heat energy from the skin to the environment are discussed in CHAPTER 16. Thermoregulatory behavior, piloerection, and other thermally-induced responses provide some clues to the functions of thermal sensors.

REFERENCES, UNIT 8.6

THERMAL SENSES

SERIALS

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CLOSING COMMENTS

CHAPTER 6: SYSTEMS PHYSIOLOGY, contains descriptions, sometimes brief, of the functions of different systems of the body. These descriptions of system functions are in addition to descriptions of system characteristics in CHAPTER 1. There would be some advantages to discussing system characteristics and functions together, but then the discussions of a part of the animals body becomes so lengthy that relationships to other parts and the workings of the integrated whole cannot be easily worked into the overall discussion. The present organization of the material reflects a functional approach to the whole organism.

A limited number of WORKSHEETS have been included in CHAPTER 6. Many of the descriptions of system functions do not lend themselves well to mathematical representation, and data are lacking on many functions.

The next chapter (CHAPTER 7: ENERGY METABOLISM) describes one of the fundamental characteristic of life. All system functions are dependent on the metabolism of energy, and knowledge of the amounts of energy involved in ecological metabolism is absolutely essential when calculating the impacts of populations on range resources and carrying capacity.

> Aaron N. Moen February 23, 1981
GLOSSARY OF SYMBOLS USED - CHAPTER SIX

ADEP = Apparent digestible energy in percent

AGDA = Age in days

BHRM = Bedded heart rate per minute

FHRM = Foraging heart rate per minute

HERA = Heart rate

JDAY = Julian day

LWKG = Live weight in kilograms

MEPP = Methane production as percent of gross energy in the food

RHRM = Running herat rate per minute

SHRM = Standing heart rate per minute

TPLW = Testes weight as a percent of live weight

WHRM = Walking heart rate per minute

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AABIA Annals of Applied Biology ABBIA Archives of Biochemistry and Biophysics ABBGB Animal Blood Groups & Biochemical Genetics ACATA Acta Anatomica Anatomia Histologia Embryologia AHEMA American Journal of Anatomy AJANA A.JBSA Australian Journal of Biological Sciences AJMEA American Journal of Medicine AJPHA American Journal of Physiology American Journal of Veterinary Research AJVRA Arkansas Academy of Science Proceedings AKASA Allattani Kozlemenyek ALLKA AMNTA American Naturalist AMZOA American Zoologist Anatomischer Anzieger ANANA ANBEA Animal Behaviour Animal Production ANIPA ANREA Anatomical Record Annals of the New York Academy of Sciences ANYAA APARD American Association Zoo Veterinarian Annual Proceedings Applied Microbiology APMBA ARPHA Annual Review of Physiology APSSA Acta Physiologica Scandinavica Supplementum ATICA Arctic ATRLA Acta Theriologica AVCSA Acta Veterinaria Scandinavica **AVS PA** Acta Veteinaria Scandinavica Supplementum AZFEA Acta Zoologica Fennica AZOFA Annales Zoologici Fennici BEHAA Behaviour BIGEB **Biochemical Genetics** BIJOA **Biochemical Journal** BIREB Biology of Reproduction British Journal of Nutrition BJNUA BLOOA Blood BLUTA Blut BPURD Biological Papers of the University of Alaska Special Report BRHLA Biorheology BUCDA Bulletin of the Georgia Academy of Sciences California Fish and Game CAFGA CAFNA Canadian Field Naturalist CATRB Calcified Tissue Research CBCPA Comparative Biochemistry and Physiology CBPAB Comparative Biochemistry and Physiology A Comparative Physiology CIRIB Proceedings International Congress on Animal Reproduction and

Artificial Insemination

Canadian Journal of Biochemistry CJBIA CJCMA Canadian Journal of Comparative Medicine CJZOA Canadian Journal of Zoology Clinical Chemistry CLCHA Canadian Journal of Comparative Medicine and Veterinary Science CNJMA Canadian Journal of Animal Science CNJNA CNVJA Canadian Veterinary Journal COVEA Cornell Veterinarian Chesapeake Science CPSCA Canadian Wildlife Service Occassional Paper CWOPA ECOLA Ecology European Journal of Biochemistry EJBCA ENDOA Endocrinology EVOLA Evolution FEPRA Federation Proceedings FOSCA Forest Science FUNAA Fauna GENTA Genetics GNKAA Genetika GRBNA Great Basin Naturalist HEREA Hereditas HLTPA Health Physics JANSA Journal of Animal Science JAPYA Journal of Applied Physiology Journal of Agricultural Science JASIA JAVMA Journal of the American Veterinary Medical Association JB CHA Journal of Biological Chemistry JCECD Journal of Chemical Ecology JCOQA Journal of the Colorado-Wyoming Academy of Sciences JDSCA Journal of Dairy Science Journal of Evolutionary Biochemistry and Physiology JEBPA JICRB Journal of Interdisciplinary Cycle Research JOANA Journal of Anatomy JOENA Journal of Endocrinology JOMAA Journal of Mammalogy JONUA Journal of Nutrition JOPAA Journal of Parasitology JPROA Journal of Protozoology JRMGA Journal of Range Management JRPFA Journal of Reproduction and Fertility JSFAA Journal of the Science of Food and Agriculture JWIDA Journal of Wildlife Diseases JWMAA Journal of Wildlife Management JZAMD Journal of Zoo Animal Medicine JZOOA Journal of Zoology

KPSUA Khimiya Prirodnykh Soedinii

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LAANA Lantbrukshogskolans annaler LBANA Laboratory Animals LBASA Laboratory Animal Science MAMLA Mammalia MGQPA Minnesota Department of Natural Resources Game Research Project Quarterly Progress Report Murrelet, The MRLTA Nutrition Abstracts and Reviews NARFA NATUA Nature North American Wildlife and Natural Resources Conference, NAWTA Transactions of the, NCANA Naturaliste Canadien, Le NFGJA New York Fish and Game Journal NJZOA Norwegian Journal of Zoology New York State Conservationist NYCOA New Zealand Journal of Science NZJSA OJSCA Ohio Journal of Science PAANA Proceedings of the Australian Society of Animal Production Pennsylvaina State University College of Agriculture PAARA Agricultural Experiment Station Progress Report PASCC Proceedings of the Alaskan Scientific Conference PCBSA Proceedings of the Canadian Federation of Biological Societies Proceedings of the Southeastern Association of Game and Fish PCGFA Commissioners PCZOA Proceedings of the International Congress of Zoology PHREA Physiological Reviews PIAIA Proceedings of the Iowa Academy of Science PNSUA Proceedings of the Nutrition Society POASA Proceedings of the Oklahoma Academy of Science Proceedings of the Society for Experimental Biology and Medicine PSEBA QJEPA Quarterly Journal of Experimental Physiology and Cognate Medical Sciences RSPYA Respiration Physiology SCAMA Scientific American SCIEA Science SOVEA Southwestern Veterinarian Southwestern Naturalist SWNAA SZSLA Symposia of the Zoological Society of London TAMSA Transactions of the American Microscopical Society THGNB Theriogenology TJSCA Texas Journal of Science TNWSD Transactions of the Northeast Section, The Wildlife Society

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UABPA UASPA UCPZA	Biological Papers of the University of Alaska Proceedings of the Utah Academy of Sciences, Arts and Letters University of California Publications in Zoology
	Unomine Academiterrel Exceptions Chetics Pulletia
WALDA	wyoming Agricultural Experiment Station Bulletin
WDABB	Bulletin of the Wildlife Disease Association
WLDHA	Waldhygiene
WLMOA	Wildlife Monographs
WLSBA	Wildlife Society Bulletin
WMBAA	Wildlife Management Bulletin
XAARA	U S Department of Agriculture, Agricultural Research Service
XAMPA	U S D A Miscellaneous Publication
XARRA	U S Forest Service Research Note RM

ZEBFA Zhurnal Evolyutsionnoi Biokhimii i Fiziologii

ZEJAA Zeitschrift fuer Jagdwissenschaft

ZETIA Zeitschrift fuer Tierpsychologie

ZOLZA Zoologicheskii Zhurnal

ZOOLA Zoologica

ZSAEA Zeitschrift fuer Saeugetierkunde

acpr	Academic Press	New York	nyny
amph	American Physiolocial Society	Washington, D. C.	wadc
anth	Antheneum	New York	nyny
apcc	Appleton-Century-Crofts	New York	nyny
butt	Butterworth	London	loen
butt	Butterworth	Washington, D. C.	wadc
caun	Cambridge University Press	New York	nyny
ccth	Charles C. Thomas	Springfield, IL	spil
chha	Chapman & Hall	Appleton, WI	apwi
coup	Cornell University Press	Ithaca, NY	itny
cupr	Cambridge Univ. Press	Cambridge, England	caen
dcch	C. D. Church, Oregon State	Corvallis, OR	coor
edar	Edward Arnold	London	loen
else	Elsevier	New York	nyny
grst	Grune and Stratton	New York	nyny
haro	Harper and Row	New York	nyny
hein	Heinemann	London,England	loen
hill	Hill	London, England	loen
hrwi	Holt, Rhinehart & Winston, Inc.	New York	nyny
iepu	Intext Education Publishers	New York	nyny
isup	Iowa State University Press	Ames, IO	amia
iucn	IUCN	Morges, Switzerland	mosw
jwwa	J. W. Walsh	Portland, ME	pome
1efe	Lea and Febiger	Philadelphia, PA	phpa
libr	Little, Brown	Boston, MA	noms
macm	MacMillan Company	New York	nyny
meth	Methuen & Co., Ltd.	London	loen
mhbc	McGraw-Hill Book Co., Inc.	New York	nyny
moco	C. V. Mosley Co.	St. Louis, MO	salo
nhfg	New Hampshire Fish & Game Department	Concord, NH	conh
orpr	Oriel Press	Newcastle Upon Tyne, Eng.	nute
orst	Oregon State University	Corvallis, OR	coor

pepr phli prha	Pergamon Press Philosophical Library Prentice-Hall, Inc.	Oxford, England New York Englewood Cliffs, NJ	oxen nyny ecnj
saco	Saunders Publishing Co.	Philadelphia,PA	phpa
tapl	Taplinger	New York	nyny
uchp unep	Univ. of Chicago Press University of New England Publishing Unit	Chicago, IL Armidale, Australia	chil arau
vare	Van Nostrand-Reinhold	New York	nyny
wbsc whfr wiwi	W. B. Saunders Company W. H. Freeman Company Williams and Wilkins	Philadelphia San Francisco, CA Baltimore, MD	phpa sfca bama

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2.la	Heart rates of white-tailed deer fawns (odvi) in relation to age and activity
2.1b	Heart rates of adult white-tailed deer (odvi) in relation to activity and time of year
4 . 1a	Testes development as percent of body weight in male white-tailed deer (odvi) through

THE BIOLOGY AND MANAGEMENT OF WILD RUMINANTS

CHAPTER SEVEN

ENERGY METABOLISM OF WILD RUMINANTS

by

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Published by

CornerBrook Press Box 106 Lansing, N.Y. 14882

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Library of Congress Catalog Card Number 80-70984

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CHAPTER 7. ENERGY METABOLISM

Metabolism, the biochemical processes characteristic of life, includes both assimilation (anabolism) and breakdown of protoplasm (catabolism). When the rate of assimilation equals the rate of breakdown, metabolism is at the maintenance level. If the rate of assimilation exceeds the rate of breakdown, growth or weight gains occur, and if breakdown exceeds assimilation, weight losses occur.

Anabolism occurs at different rates in different tissues, depending on the age, time of year, and activity of the animal. A young growing animal has a higher rate of metabolism per unit body weight than an older mature one. Young growing animals deposit more protein and skeletal tissue, and older, more mature animals deposit more fat tissue. Anabolism is greater in the summer and fall than in the winter and spring. In fact, weight losses usually occur in the winter as part of an adaptive survival strategy. Active animals have higher rates of metabolism than inactive ones, and activity levels are often reduced in the winter as another adaptive survival strategy.

Catabolism, the breakdown of tissue, is the reverse of anabolism. It is a part of maintenance. Some tissues are broken down and their energy, protein, and minerals become available to the animal. Body fat is an energy reserve that may be mobilized when ingested nutrients are not sufficient to meet the energy needs of the animal. Male Cervids partition resources into antlers in spring and summer, mobilizing minerals from some of their bones for deposition in the antlers. Pregnant females partition nutrients into their own maternal tissue and into fetal and reproductive tissue. Lactating females assimilate nutrients into milk, sometimes mobilizing their own tissues to do so. These are important capabilities when forage resources are depleted or unavailable.

Ruminants also have metabolic adaptations for the conservation of resources. Lower energy metabolism in the winter when range resources are more limited and higher metabolism in the summer when range resources are less limited is a seasonal conservation adaptation. This is an important metabolic adaptation for wild ruminants living on ranges with seasonally fluctuating resources, and may be critical for survival when range resources are especially limited, as in severe winters.

Measurements of energy metabolism have been made for many years. The French scientist, Lavoisier completed a very basic experiment in metabolism by placing mice in a container surrounded by ice and measuring the rate of ice-melt. He found that the energy required to melt the ice was related to the heat production of the mice, providing an early estimate of metabolism. Techniques have been refined considerably since that early experiment. While metabolic chambers are still used to directly measure the heat given off by an animal, indirect measurements and estimates are often made by gas analyses, and the amounts of oxygen consumed and carbon dioxide released used to estimate metabolic rates.

Several terms are used in the literature to describe metabolism measurements. Basal metabolism has been defined as the minimum energy cost when an animal is at rest in a thermoneutral environment and in a postabsorptive nutritive condition (Brody 1945). The definition of basal metabolism implies standard conditions for measurement rather than minimum metabolism for life. The resting animal is not active, nor is it sleeping. Thermoneutrality is the condition of being neither too hot nor too cold while in the metabolism chamber. Then, there are no thermogenic responses; the animal makes no attempt to keep from getting too hot (by panting, for example) or too cold (by shivering, for example). The post-absorptive condition eliminates the metabolic contributions of rumen microflora to the total heat production of the animal. These microflora produce their own metabolic heat as they break down the ingesta in the rumen. This heat is not a part of the host's metabolic heat production.

The energy required to maintain life during basal metabolism tests is used to meet the costs of circulation, excretion, secretion, respiration, and the maintenance of muscle tone. Basal metabolism is a laboratory measurement that is useful for comparative purposes, but it is not applicable to the field situation since free ranging animals seldom meet the conditions prescribed for a metabolism test.

The term <u>fasting metabolism</u> applies to metabolism tests conducted on animals in a post-absorptive condition and in a thermoneutral environment, but not corrected for activity. Results are given in this way when corrections for activity during the test cannot be made.

Ecological metabolism is an expression of the total cost of maintenance, activity, and production processes of free-ranging animals (Moen 1973:123-124). The cost varies because the rates of these processes vary. Some of the variability is rhythmic due to seasonal patterns in weather and thermal exchange, the timing or reproductive functions, activity patterns, and other biological functions. Some of the variability is due to the effects of transients, such as quick changes in the level of energy expenditure for activity due to a flight response from a predator. The metabolic expenditures for each of these processes accumulate over a 24-hour period, resulting in the ecological metabolism per day (ELMD).

One very useful expression for evaluating rates of energy metabolism is base-line metabolism. This expression is a mathematical base-line, a common denominator used when dividing ecological metabolism per day (ELMD) to determine a multiple of base-line metabolism (MBLM) for comparative purposes, numerically equal to (70)(MEWK) KCAL per day and (294)(MEWK) KJOU per day (MEWK = $W_{kg}^{0.75}$). While the numerical value approximates the average basal metabolism of a large number of species, base-line metabolism should be considered as a mathematical base-line only and not as an average basal metabolism value. Average basal metabolism of white-tailed deer varies seasonally (Moen 1973), and averages are not appropriate representations of metabolic rhythms that vary in a patterned way through the year. Time-related ecological metabolism should be expressed as a function of time; the use of averages for long time periods results in the loss of too much information of ecological significance.

The ratio of ELMD to BLMD is called the <u>multiple of base-line metabol-</u> ism (MBLM). This ratio is weight-independent: the effects of weight appear in both the numerator and the denominator (MBLM = ELMD/BLMD). MBLM is especially useful when comparing ELMD for different species. When ELMD is expressed as a multiple of base-line metabolism, interspecies comparisons can be easily made since differences in size have been eliminated by the use of weight in both the numerator and denominator. MBLM may be used to establish a reasonable biological framework for expressing ELMD for a variety of species; calculations indicate that MBLM values approaching 5 are unrealistically high, and below 1.3 or so, low. Details for different species are discussed next.

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ENERGY METABOLISM

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TOPIC 1. FACTORS AFFECTING ENERGY METABOLISM

Nothing is free. The maintenance of basic life processes, the completion of activities, and the production of new tissues are all energyrequiring processes. The energy costs of these processes can be estimated because the energy transfers involved are predictible processes. Equations can be derived that provide good first approximations of the costs of these processes in different ecological circumstances.

Each individual must meet its own requirements for the basic life functions--respiration, circulation, daily activities that provide food necessary to support these functions, maintenance of body tissues, production of new tissues to replace that metabolized--that combine to establish a maintenance requirement of each individual. Since metabolism is an energyusing process, the energy used must be replaced each day in order to sustain life. If the energy requirement is not being met, the animal is in a negative energy balance, and if the amount of energy in ingested nutrients exceeds the energy required, the animal is in a positive energy balance.

The energy flow through an organism includes several pathways of use and dissipation illustrated below. The numbers below each of the terms are relative values illustrating the efficiencies of an animal given by Crampton and Harris (1969:145) at each step.



The efficiencies of energy use have a direct bearing on the amounts of food required. Further, the nutritional requirements are constantly changing as a result of physiological and environmental variables. Since so little is known about the metabolic efficiencies and maintenance, activity, and productive functions of wild ruminants, it is difficult to assess the effects of the different variables in the lives of free-ranging animals in their natural environments. Nevertheless, some patterns may be recognized from data on both domestic animals and wild ruminants, providing insights into relationships between wild ruminants and their natural environments.

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UNIT 1.1: BIOLOGICAL FACTORS AFFECTING ENERGY METABOLISM

Some of the biological factors affecting energy metabolism are historically determined, and some are variables due to circumstances. The former include factors that are determined by genetic and by "programmed" physiological changes. The latter include interspecies and intraspecies relationships present due to current circumstances.

GENETIC

It is well known that animals exhibit individual variation due to genetic differences. The more obvious characteristics, such as coat color, body size, etc., are often the only characteristics considered by field biologists when studying particular populations. Since energy metabolism is so vital to the survival of the individual, genetic factors which affect the energetic efficiencies of animals may have a significant bearing on their survival, especially when conditions of stress begin to appear.

SEX

The two sexes have different metabolic rates at different times of the year; differences in metabolic rates of the sexes are related to their different biological roles. Males are very active during the breeding season, spending a lot of energy for activity. Females are very active during the lactation period, especially in foraging activity when they must supply nutrients for milk production.

AGE

It is well known that the metabolism per unit weight of young animals is higher than that of adults; the metabolic rate of an adult, relative to weight, is usually less than one-half that of a newborn. Lambs reach <u>summit</u> <u>metabolism</u>, the maximum heat production possible without voluntary muscular activity, within one-half hour after birth, and maximum values may be 15 times greater than fetal levels and 5 times greater than "basal" levels (Alexander 1962a). He noted that there was no increase after ingestion of milk, and heat production declined slowly with advancing age, prolonged exposure to cold, and continued fasting.

The physiological capabilities of newborn wild ruminants are quite well-developed as the young are active shortly after birth. Some species, such as caribou, goats, sheep, and muskox, sometimes give birth to young in rather severe weather conditions, and only well-developed offspring can survive. Newborn caribou are able to follow their dam within hours after birth, indicating a high level of physiological development, with survival very much dependent on the nutritional ties between the neonate and the dam.

HAIR CHARACTERISTICS

Since warm-blooded animals maintain a specific internal body temperature (with some variation of course), hair coats with lower insulation values may necessitate higher levels of heat production by the animal. If two animals with equal surface areas were exposed to identical thermal environments, the one with the better insulation would lose less heat. If heat production were identical for the two animals, the one with the better coat would have a more positive heat balance. Brody (1954) attributed the ability of cattle to withstand temperatures to -40° F to their coat insulation and highly developed vascular control of the thermal conductivity of peripheral tissues. Wild ruminants are very dependent on hair characteristics for insulation from the cold, and have seasonal molts to accomodate seasonal changes in weather.

RELATION TO SURFACE AREA

Interpretations of the basal metabolic rate have been made by physiologists for many years, and numerous disagreements have arisen over its relationship to surface area and body weight. The "surface area law" was formulated because it could be shown that heat loss from any object is proportional to surface area, and since heat production must be proportional to heat loss if homeothermy is to be maintained, it was concluded that heat production must be proportional to surface area. This was demonstrated for dogs and rabbits, and the concept was widely applied to other species as well (Brody 1964:356).

However, one factor in the relationship between surface area and heat production is the variability in the surface area of a single animal. A standing animal has a greater surface area than the same animal lying down and curled up. Thus the surface area of an individual is variable, and living organisms may be constantly changing their surface area, heat production, evaporative heat loss, muscular activity, diet, hair insulation through pilo-erection, and other characteristics to maintain a homeothermic condition. If one considers the complexity of heat exchange by radiation, conduction, convection, and evaporation, it becomes immediately obvious that surface area is only one of several important parameters operating in heat exchange. A simple relationship observed empirically between surface area and heat production can hardly be called a "law" in view of the many other associated variables.

If, however, one considers two animals of the same species with identical thermal characteristics of the coats and equal rates of heat production per unit of metabolic tissue, the larger animal with its more favorable surface area:weight ratio should be able to withstand more cold stress than a smaller one. It is possible to calculate the critical surface area (Moen 1968) of an animal. The critical surface area is the surface area:weight relationship which is just sufficient to result in a net thermal exchange of zero. Larger animals have relatively less surface area than smaller ones, and they also have more metabolic tissue and greater metabolic potentials. Heavier lambs were able to maintain homeothermy under conditions of higher heat loss than lighter ones (Alexander 1962b), and this may be due not only to their more favorable size and surface area ratio but also their larger amounts of metabolic resources.

RELATION TO BODY WEIGHT

A large animal has a higher absolute metabolism than a small one. The heat production per unit weight of a small animal is greater than the heat production per unit weight of a large animal, however. The relationship is illustrated below.



Weight

Benedict's mouse to elephant curve (Benedict 1938; see also Dukes 1955:625 and Moen 1973:116), widely known in physiological circles, is a quantified display of this relationship; variations between species were found to be quite low and the smooth and logical shape of the entire graph striking. Deviations from the curve may be real or they may be artifacts, and perhaps are a combination of the two.

The total mass of an animal includes some tissue that is not actively participating in the metabolic processes but is the final product of metabolic processes. The hair, horns, and antlers, once growth is completed, are examples. Ingesta in the gastro-intestinal tract is another example; it adds to body weight but is not part of the animal's metabolic tissue.

There has been considerable discussion of the mathematical relationship between heat production and body weight. The formula for this relationship is:

$Q = k W^{b}$

where Q is the heat production in calories per day, k is a constant, W is the weight in kilograms, and b is an exponent that has generated most of the controversy. Physiologists have been searching for an exponent which will result in a constant value of k. If this could be found, the basal heat production of any animal could be determined by simple multiplication after measuring the weight of the animal.

Chapter 7 - Page 9

Published values of b have ranged from 0.66 to 0.75. Much of the controversy is in the rounding off of the value. Brody (1945) suggests that b should be 0.7 and Kleiber (1961) uses 0.75. The National Research Council (1966) adopted the exponent b = 0.75. Fasting homeotherms under standard conditions produce about 70 kcal of heat per $kg^{0.75}$ (Kleiber 1961). Measurements of four deer in winter coat resulted in an average of 75.6 Kcal per $kg^{0.75}$ per 24 hours, and measurements of three deer in summer coat had an average of 84.4 Kcal per kg^{0.75} (recalculated from Silver 1968. see Since winter and summer differences are part of the Moen 1973:117). rhythmic seasonal variations in metabolism of deer, I no longer calculate an average basal metabolism for deer, choosing rather to use $70W_{K\rho}^{0.75}$ as a and representing ecological metabolism mathematical base-line with time-dependent equations.

REPRODUCTIVE CONDITION

Three stages in the reproductive cycle--breeding, gestation, and lactation--have definite effects on the energy metabolism. It is well known that male wild ruminants expend large amounts of energy in increased activity during the breeding season and lose considerable weight. During gestation, the female must expend additional energy to maintain the uterus, to meet increased demands on the circulatory, respiratory, and excretory systems, to handle the endocrine influences on her own metabolism, and provide for fetal growth (Brody 1945). The energy metabolism per unit weight is not higher for pregnant females until the last 1/3 of pregnancy when fetal growth is much accelerated, however (Morrison 1948). During rapid fetal growth, elevated metabolism is expected since growth demands additional energy for synthesis of fetal tissue. The high rate of fetal metabolism is followed by a declining but still high neonate metabolism, after which metabolic rates decline to more stable and rhythmic adult levels.

Lactation places a high physiological demand on the female as milk is synthesized from the nutrients in ingested food. The heat production of cattle during heavy lactation is approximately 100% above the non-lactating level, with the increase directly proportional to milk production. The higher metabolic rate is associated with the higher food consumption and milk prodution rather than with muscular activity (Brody 1945:820). Calculations of the cost of lactation for white-tailed deer indicate that it is the most costly sustained biological process there is (Moen 1973 and Robbins and Moen 1975).

PATHOGENS AND PARASITES

Whenever one organism supports the life processes of another organism living upon or within it, the energy requirements of the host animal must go up unless the two species exhibit a kind of mutualism (Odum 1959) in which the growth and survival of both populations is benefited. The relationship between a ruminant and the rumen flora and fauna is an example of mutualism. Pathogens and parasites, however, increase the catabolic mechanisms of an animal (Jones 1963 in Thayer), resulting in increased energy demands.

HEAT INCREMENTS

Heat production increases when an animal becomes more active since muscle contractions are exothermic. Digestion of food in the gastrointestinal tract also results in an increment in heat production of an animal. It is defined as an extra heat increment incident to the total nutritive process, including the energy cost of excretion of the end products (Brody 1945:59). It is sometimes called an energy waste associated with the use of food, but the extra heat energy may be of value during cold weather as it helps keep the animal warm.

What heat increments are characteristic of animals on different diets? Blaxter (1962:141) presents a table showing the differences in heat production of sheep and steers on fasting, maintenance, and full-feed diets. For sheep the ratio of the maintenance to the fasting diet is 1.47/1, and of the full feed diet to the fasting diet 1.88/1. The ratio for steers on a maintenance diet compared to a fasting diet was 1.38/1. Such increments are of interest to the field biologist when analyzing the energy balances of animals on ranges with different food supplies, topographies, and weather conditions.

ACTIVITY

Activity, or the movement of mass through distance, is work. Every animal must expend energy when moving its mass horizontally and vertically for whatever distances are involved. Range and wildlife managers ought to be interested in the amounts of energy required for these movements since topographic relief, distances to water and other range characteristics may have effects on the efficiencies of the animals and on population ecology.

The speed with which an animal moves also has an effect on the energy cost of movement. In sheep, the energy cost of horizontal locomotion increased but the cost of ascending vertical gradients decreased with increasing speed (Clapperton 1961). It is interesting to speculate on the energy costs involved in rapid bursts of speed exhibited by wild ruminants during escape behavior.

SOCIAL AND PSYCHOLOGICAL EFFECTS

Numerous sociological and psychological stressors influence energy requirements. Pfander (1963:117) lists such things as confinement vs. natural range, herd vs. individual response, numbers per group, space per animal, noise level, and other disturbances as factors that affect energy requirements of domestic species.

One unknown factor in basal metabolism measurements is the effect of confinement on the psychology and metabolism of the animal. Well-trained animals appear more calm, of course, but visible signs of excitement need not be present to have elevated metabolism. A male white-tailed deer that was rescued from an ice-covered lake (Moen 1967) showed body temperature variations that were not accompanied by visible signs of excitement when resting in the garage at the laboratory. The deer showed a steady increase in body temperature from a low of 26.4° C to a high of 39.5° C as it was warmed up, after which it returned to 38° C. My presence in the room, however, caused the temperature to rise quickly to 38.5° C, but the animal showed no signs of fear. When I left the room, body temperature declined to 38° C again. The docile nature of the deer when in the garage changed instantly when it saw the trees through the open garage door as we prepared to release him. After thrashing around in its nylon mesh cage, the deer was finally released and ran directly into the woods.

RHYTHMIC CHANGES IN THE BASAL METABOLIC RATE

Marked changes in the rates of metabolism during daily and seasonal periods have been observed. Nocturnal animals have a lower BMR during the daytime (Benedict 1938), and times of muscular activity and metabolic rates are related (Brody 1945). Seasonal variations also occur. A depressed winter metabolism in white-tailed deer was first reported by Silver (In Siegler 1968:191), but the number of deer studied was not sufficient to warrant a definite conclusion about seasonal lability. Several years of research at the Wildlife Ecology Laboratory have shown definite seasonal metabolic rhythms in ecological metabolism (Moen 1978).

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJAEA 11--1 85 96 dosh seas var wool gr, heat tol wodzicka,m 1960 AJAEA 13--1 82 dosh temp reg, metab, newborn 1 alexander,g 99 1962 AJAEA 13--1 144 164 dosh energ metab, starved newbo alexander,g 1962 AJAEA 15--6 969 973 dosh ener cost, expend, feed ac graham, nmcc 1964 BJNUA 16--4 615 626 dosh fasting metab, adult wethe blaxter,kl 1962 COVEA 57--1 43 53 dosh erythrocyte loss, restitut georgi, jr; whitlo 1967 PAANA 5---1 138 140 dosh effect lactat, wool growth corbett, jl 1964 PNSUA 20- xxxi xxxii dosh energy expenditur, walking clapperton, jl 1961

SCIEA 145-- 1318 dosh hematocr val, hemoglo type evans, jv; whitloc 1964

CODEN	vo-nu	BEPA	ENPA ANI	M KEY WORDS AUTHORS	YEAR
АЈРНА	219-4	1104	1107	<pre>scali, energ cost, running taylor,cr; schmi/</pre>	1 97 0
AMNTA	99	373	376	nat mort, reprod, min meta wilson,ta	1965
AMZOA	81	71	81	thermodynam consid, nutrit wiegert,rg	1968
ATRLA	24-12	125	136	metab, body size, homeothe poczopko,p	1979
				continued on the next page	

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CODEN	vo-nu	BEPA	ENPA ANIM	KEY WORDS AUTHORS	YEAR
CIWPA	503	1	215	vital energ: comp bas meta benedict,fg	1938
FEPRA	294	1524	1532	ener metab, bod size, scal schmidt-nielsen,k	1970
JANSA	433	692	704	fasting heat prod, body wt thonney,ml; touc/	1976
JCCPA	31	281	291	oxyge consump, basal condi morrison,pr	1 9 48
NUMEB	211	88	104	energet fat metabo, growth van es,ajh	1977
OJSCA	746	370	380	energetics of endotherms mcnab,bk	1974
PECTD	12	85	101	bioenerg params, wild rumi drozdz,a; weiner/	1975
PHREA	274	511	541	body size, metabolic rate kleiber,m	1947

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Davydov, A. F. On the processes of muscular activity in reindeer during procurement of feeds from under the snow. Russian original from Sbornik "opyt izucheniya regulyatsii fiziologicheskikh funkstii v estestvennykh uslovyakh sushchestvovania organizmov," Vol. 6:35-40. 1963. Translation available from U.S. Dept. Commerce, Clearinghouse for Federal Scientific and Technical Information, Springfield, Va.

CHAPTER 7, WORKSHEET 1.1a

Resting heat production of black-tailed deer

Measurements of the resting heat production of male and female blacktailed deer fawns have been made by Nordan et al. (1970). The weight and heat production measurements reported in TABLE II (p. 278) make it possible to calculate multiples of base-line metabolism (MBLM) at each weight. The data and calculations are given below.

	LWKG	REHP	LWKG ^{0.75}	BLM	MBLM
Males:	5	382	3.34	234	1.63
	10	880	5.62	394	2.24
	15	1180	7 - 62	534	2.21
	20	1460	9.46	662	2.21
	25	1722	11.18	783	2.20
Females:	5	498	3.34	234	2.13
	10	837	5.62	394	2.12
	15	1076	7.62	534	2.01
	20	1408	9.46	662	2.13
	25	1669	11.18	783	2.13

Note how similar the MBLM values are throughout this range of body weights.

Keep in mind these are <u>resting</u> heat production values, and will be different from <u>ecological metabolism</u>. The MBLM for the ecological metabolism of a growing fawn will be higher (see CHAPTER 7, Page 2 and also TOPIC 6).

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Nordan, H. C., I. McT. Cowan, and A. J. Wood. 1970. The feed intake and heat production of the young black-tailed deer (Odocoileus hemionus columbianus). Can. J. Zool. 48(2):275-282. ١

CHAPTER 7, WORKSHEET 1.1b

Energy metabolism of pronghorn

The total heat production and fasting heat production of female pronghorns from 2 months to 18 months are given by Wesley et al. (1973). Calculations of MBLM are easily made from the age, weight, and heat production data presented in Table 2 (p. 567). The following table is derived from their data.

Month A of Year	ge Class (months)	LWKG	MEWK*	Total Heat Production	MBLM	Fasting Heat Production	MBLM
July	2.0	8.9	5.15	783	2.17	603	1.67
January	7.5	26.1	11.55	1316	1.63	843	1.04
May	12.0	26.8	11.78	1425	1.73	884	1.07
November	18.0	37.8	15.24	1707	1.60	1220	1.14

*MEWK = metabolic weight in kg = $LWKG^{0.75}$

Note how MBLM for total heat production varies from 1.60 to 2.17. Variations after the age of 2.0 months may be a reflection of a seasonal rhythm, but there are too few data through the annual cycle to confirm that.

Evaluate these MBLMS in relation to the seasonal rhythms of white-tailed deer discussed in TOPIC 6.

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CHAPTER 7, WORKSHEET 1.1c

Behavioral effects on the energy expenditures of white-tailed deer

The effects of different amounts of physical activity in six different behavioral regimes on energy metabolism have been demonstrated in a sequence of calculations in Moen (1976). The least activity, with 75% of the time spent bedded, 10% standing, 10% foraging, 5% walking, and no running, resulted in graphed values of energy expenditure as low as 1700 kcal per day for a 60-kg deer. The most activity, with 25% of the time spent bedded, 30% standing, 12% walking, 30% foraging, and 3% running, resulted in graphed values approaching 3000 kcal per day for a 10-kg deer. These two expenses, and various combinations of effects between them, are plotted in Figure 3, page 198 of Moen (1976).

Determine multiples of base-line metabolism for the energy expenditures given in the paragraph above with the following formula:

MBLM = Energy expenditure/(70)(MEWK)

where MBLM is the multiple of base-line metabolism, and MEWK is metabolic weight in kg, and (70)(MEWK) is base-line metabolism. Both of these are discussed on Chapter 7 - Page 2.

Other values of MBLM may be determined from Figure 3 in Moen (1976) also. Note that the 1976 publication includes discussions of multiples of basal metabolism rather than <u>base-line</u> metabolism. This change in terminology is due to clarification of concepts as a result of analyses described in Moen (1978).

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UNIT 1.2: ENVIRONMENTAL FACTORS AFFECTING ENERGY METABOLISM

The concept of thermoneutrality--the thermal range in which an animal does not need to respond physiologically to maintain normal body temperature --is well known. The terms "upper critical temperature" and "lower critical temperature," deeply entrenched in the literature, are meant to represent the limits of the thermoneutral range on a thermometric scale. Many more parameters are involved in heat exchange in the natural world than temperature alone, however. Further, temperature measurements are on an ordinal scale, and do not represent the actual amount of energy exchanged in a particular system. I have suggested that the concept of a critical thermal environment (Moen 1968) replace the "critical temperature" idea used previously. Critical thermal environment is a much more appropriate term in view of the many factors involved in heat exchange.

A critical temperature may be determined if all thermal parameters except temperature are held constant in a particular system. In the same manner, however, a critical wind velocity can be determined if temperature and all other thermal parameters are held constant. Similarly, critical values for body weights, evaporation rates, postures, heat production, and many other parameters can also be calculated.

The concept of thermal neutrality is physiologically sound. The term critical thermal environment is comprehensive and applies to the concept of thermoneutrality.

Another environmental factor affecting energy metabolism is snow. The depth and density characteristics of snow can have a major effect on energy metabolism as animals of different sizes and weights sink to different depths.

Surface characteristics also affect the metabolic cost of walking. Less energy expenditure was measured on reindeer walking on roads than on tundra (White and Yousef 1978), indicating that the more firm road surface was easier to walk on than the softer tundra surface.

The topography affects energy metabolism as the cost of walking on the level is less than the cost of walking upslope. The cost of walking downslope is less than the cost on an upslope (Robbins et al. 1977), but the compensation is not equal to the upslope costs.

Environmental stimuli, such as mechanical noises, wind, predators, and other stimuli, may get the attention of an animal and increase the energy metabolism. Many of these transients are of short duration, such as a jet plane passing overhead, and may contribute little to the total daily cost of living. Others, such as a predator, may demand a higah rate of energy metabolism for escape. Heart rate increases are discussed in Moen and Chevalier (1977); corresponding changes in energy metabolism are unknown, however.

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BISNA	18–11	1041	1043	odvi	critical thermal environme	moen,an	1968
CJZOA CJZOA	536 565	679 1207	685 1210	odvi odvi	ambien temp effect, physio heart rates w-t-d wolf how	holter,jb; urban/ moen,an; dellaf/	1975 1978
ECOLA	571	192	198	odvi	energy conservat in winter	moen,an	1976
JOMAA	484	655	656	odvi	hypotherm, water-chill dee	moen,an	1967
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
JWMAA	333	482	490	odvi	diges ener, win, mich does	ullrey, de; youat/	1969
JWMAA	344	863	000		den an antista a ser ad alta da ser		
		005	869	odvi	alges metab ener mich does	ullrey,de; youat/	1970
JWMAA	351	37	869 46	odvi odvi	effect falli temp, heat pr	ullrey,de; youat/ silver,h; holter/	1970 1971
JWMAA JWMAA	351 424	37 715	869 46 738	odvi odvi odvi	effect falli temp, heat pr seas chan, heart rate, act	ullrey,de; youat/ silver,h; holter/ moen,an	1970 1971 1978
JWMAA JWMAA	351 424	37 715	869 46 738	odvi odvi odvi	diges metab ener mich does effect falli temp, heat pr seas chan, heart rate, act	ullrey,de; youat/ silver,h; holter/ moen,an	1970 1971 1978

CJZOA 48--2 275 282 odhe feed intake, heat producti nordan,hc; cowan/ 1970 ECMOA 2---1 1 46 odhe seasonal migration, mule d russel,cp 1932 JWMAA 43--1 162 169 odhe energ requi, fawns, winter baker,dl; johnso/ 1979

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBPAB 60A-3 251 256 ceel energ, nitr metab, cold en simpson, am; webs/ 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 50--2 302 310 alal od, structur adaptat, snow kelsall,jp 1969 QSFRA 3---- 51 73 alal influence of snow on behav des meules,p 1964

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR APSCA 105-3 268 273 rata spitzberg, wint-dorman ung ringberg,t 1979 CJZOA 39--7 845 rata clima, metab, therm, infan hart, js; heroux,/ 1961 856 rata energy metab, barren groun mcewan, eh CJZOA 48--2 391 392 1970 IJBMA 12--1 21 27 rata act wint cari, snow, alask henshaw, j 1968 PHZOA 30--2 93 105 rata melt points, anim fat, col irving,1; schmid/ 1957

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 31--1 159 164 anam mort, severe wint, montana martinka,cj 1967 JWMAA 37--4 563 573 anam energy metabolism, prongho wesley,de; knox,/ 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 56-11 2388 2393 ovca winter bioenerget, rocky m chappel,rw; hudso 1978 JWMAA 35--3 488 494 ovca variat of rectal temperatu franzmann,aw; heb 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

Chapter 7 - Page 19

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARJASIA 44--1 566doca wint nutr depres, grow, re joubert,dm1954JASIA 58--1109120doca sig extremities, therm reg whitton,gc1962RSPYA 4---3353362doca oxy cost, therm-hyperven hales, jrs; findla 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR 407 AJAEA 11--3 402 dosh regula bod temp, hot envir brook, ah; short, b 1960 AJAEA 13--1 82 99 dosh wind, h2o, metab rate, lam alexander,g 1961 dosh heat excha, lamb hot envir alexander,g; will 1962 AJAEA 13--1 122 143 AJAEA 13--1 144 164 dosh energ metab, starv newborn alexander,g 1962 AJAEA 15--6 982 dosh ambi temp, heat prod, preg graham,nm 1964 988 BJNUA 21--3 769 785 dosh conti meas, heart ra, ener webster, ajf 1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CNJNA 43--1 39 46 dosh low envir temp, phys respo hess,ea 1963 JASIA 52--1 13 24 dosh ener metab, clos clipped s graham,nm; wainm/ 1959 JPHYA 198-2 251 276 dosh shiv, nonshiv thermogenesi alexander,g; will 1968 SCIEA 145-- 1318 1318 dosh hematocr val, hemoglo type evans,jv; whitloc 1964

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR ANYAA 53--3 600 607 mamm role, piliary sys, therm e herrington, lp 1951 BISNA 18-11 1041 1043 critical thermal environme moen, an 1968 FEPRA 28--3 1053 1058 intersp diff, cold adaptat jansky,1; bartun/ 1969 FEPRA 29--4 1541 1552 many rhythmic var in ener metab aschoff, jj; pohl/ 1970 JAPYA 7---4 355 364 skin temp, arcti, heat reg irving,1; krog, j 1955 NATUA 234-- 483 many hier watr, eng, desrt macfarlane,wv; h/ 1971 484 NUMEB 21--1 88 104 energet fat metabo, growth van es,ajh 1977 SSEBA 18--- 31 48 many insulat, metab adapt, cold hart, js 1964 ZOBIA 2--- 887 calor val equiv, bod wt ch dargol'ts, vg 899 1973

OTHER PUBLICATIONS

Davydov, A. F. On the processes of muscular activity in reindeer during procurement of feeds from under the snow. Russian original from Sbornik "opyt izucheniya regulyatsii fiziologicheskikh funkstii v estestvennykh uslovyakh sushchestvovania organizmov," Vol. 6:35-40. 1963. Translation available from U.S. Dept. Commerce, Clearinghouse for Federal Scientific and Technical Information, Springfield, Va.

CHAPTER 7, WORKSHEET 1.2a

Chamber temperature effects on MBLM of the white-tailed deer

The energy expenditure values for white-tailed deer in a temperaturecontrolled chamber given by Holter et al. (1975) may be converted to MBLM by dividing the y-axis values in Figure 3 (p. 682) by 70. In the summer, MBLM was about 3.0 as the ambient temperature in the chamber was lowered to near 0°C. This is a very high relative rate of metabolism as a response to cold; productive processes require MBLM's of 2.5 to 4.0 (Moen 1978). In the winter, MBLM reached a low of about 1.3 at about 10°C, and a high of about 1.8 at about -20°C. These values are within the range of values for the winter period in the annual metabolic cycle discussed in Moen (1978). No indications of a rise in energy metabolism have been observed in the deer at Cornell's Wildlife Ecology Laboratory when air temperatures have dropped from about 10°C to -20°C. These animals have been free to choose their options in response to natural weather patterns.

Air temperatures to -20° C and cooler have always been accompanied by further reductions in heart rates and energy conservation behavioral responses by these deer. Different results in the New Hampshire chamber are likely due to the fact that the number of options are very limited in a chamber, and that the freedom to select and move to locations of the deer's choice cannot be exercised.

Equations for energy expenditures during different seasons are not given in Holter et al. (1975), but values may be estimated from plotted data using a grid overlay. Complete the estimations and compare MBLM's determined with the seasonal rhythms in Moen (1978).

LITERATURE CITED

Holter, J. B., W. E. Urban, Jr., H. H. Hayes, H. Silver, and H. R. Skutt. 1975. Ambient temperature effects on physiological traits of white-tailed deer. Can. J. Zool. 53(6):679-685.

Moen, A. N. 1978. Seasonal changes in heart rates, activity, metabolism and forage intake of white-tailed deer. J. Wildl. Manage. 42(4):715-738.

CHAPTER 7, WORKSHEET 1.2b

The effects of walking speed, percent slope, and snow depths on the energy expenditures of white-tailed deer

The effects of various combinations of walking speed, percent slope, and snow depths on energy expenditures have been graphed in Moen (1976, Figure 3, p. 194). The energy costs of different combinations may be read on the y-axis and MBLM determined for a 60-kg deer. For example, a deer in Behavioral Regime 6, walking 90 cm per second on a 0% slope in about 18 cm of snow is predicted to have an energy expenditure of 2400 kcal per day. The multiple of base-line metabolism is:

$$MBLM = 2400/(70)(60^{0.75}) = 1.59$$

Determine the energy costs of different combinations and summarize in the space below. Note that the multiples of <u>basal</u> metabolism given in Moen (1976) are considered in this book to be multiples of <u>base-line</u> metabolism (see WORKSHEET 1.1c, Chapter 7 - Page 18c).

LITERATURE CITED

Moen, A. N. 1976. Energy conservation by white-tailed deer in the winter. Ecology 57(1):192-198.

CHAPTER 7, WORKSHEET 1.2c

Energy exchanges of red deer (ceel) at three chamber temperatures

Energy exchanges of young red deer at three air temperatures and three levels of feeding are given in Table 4 (p. 253) of Simpson et al. (1978). The values are presented in MJ per day, which can be converted to kcal with the following equation:

kcal per day = (MJ/4.184)1000

Multiples of base metabolism are determined by dividing kcal per day by (70)(MEWK). Using these equations, the following MBLMs have been determined:

			Level of Fee	eding
		Low	Medium	High
Metabolizable energy intake	(MBLM)	1.10	1.84	2.33
Heat production (MBLM) at	16°	1.57	1.58	1.70
	8°	2.05	2.35	2.49
	4°	1 86	2 38	2.20

These multiples indicate a rise in heat production at each level of feeding as the temperature dropped from 16 to 8°, and at only one level of feeding as the temperature dropped from 8 to 4° .

The MBLM calculations are illustrated here to call attention to the utility of MBLM as a parameter for comparisons between levels of feeding, chamber conditions, weights, and species. Verify these calculations based on the original data in Simpson et al. (1978) in order to become thoroughly familiar with the patterns of MBLM in different situations.

LITERATURE CITED

Simpson, A. M., A. J. F. Webster, J. S. Smith, and C. A. Simpson. 1978. Energy and nitrogen metabolism of red deer (<u>Cervus elaphus</u>) in cold environments: a comparison with cattle and sheep. <u>Comp. Biochem.</u> Physiol. 60:251-256.

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CHAPTER 7, WORKSHEET 1.2d

Effects of weather on energy metabolism of infant caribou

The effects of different combinations of air temperature, wind and precipitation on the metabolism of infant-caribou was studied by Hart et al. (1961). Results are presented in Table III (p. 850), including a column for heat production in kcal per hour/kg^{0.75}. These data may be converted to MBLMs by simply multiplying the HP value by 24 and dividing by 70. Values of MBLM range from about 1.4 for a dry calf in the lab at 0°C to over 10 for a calf exposed to wind and precipitation.

Complete the calculations of MBLM for each of the test conditions given in the published paper and evaluate them in relation to the experimental variables. Examples are MBLM in relation to air temperature (the first five results in Table III) and MBLM in relation to wind velocity (the last six results in Table III). Multiple regression analyses may be useful here too, except that the effects of wind on MBLM may not be linear. This is discussed further in PART V.

Note that Hart et al. indicate that 25 kcal per hour/kg^{0.75} is a "lethal level." This is (25)(24)/70 = 8.6 = MBLM, which is a higher rate of metabolism than that of very high producing dairy cows! Infant caribou can hardly be expected to maintain such a metabolic level for very long.

LITERATURE CITED

Hart, J. S., O. Heroux, W. H. Cattle and C. A. Mills. 1961. The influence of climate on metabolic and thermal responses of infant caribou. Can. J. Zool. 39(7):845-856.

TOPIC 2. MEASUREMENTS OF ENERGY METABOLISM

Measurements of the metabolic rates of several species of wild ruminants have been made with a variety of methods at different times of the year. The results show variations in relation to several parameters of ecological importance. The time of the year is an important consideration for evaluating seasonal rhythms, and the day, week, or at least the month of the year in which the tests were conducted should always be given in the results. If the time of year cannot be determined, the results are relatively worthless.

Weights of the animals should also be given, of course. Metabolic expenditures can then be determined per unit weight, or per unit metabolic weight. Sex, age, and reproductive characteristics are also important characteristics for interpreting the results of metabolism tests.

The topics and units that follow give the results of metabolism tests and calculations for a variety of wild ruminants. Short-term chamber measurements are included, though it is difficult to derive equations for ecological metabolism over the entire annual cycle from such studies. Calculations by summation, estimations from vital signs and in relation to base-line metabolism provide estimates throughout the year.

UNIT 2.1: CHAMBER MEASUREMENTS

Direct measurements of metabolism involve the measurement of the heat production of the animal after the manner of Lavoisier in the latter part of the eighteenth century. His early methods have been replaced by very refined calorimetry chambers, however. Professor Armsby built one early in the 20th century at the Pennsylvania State University; it is still in existence but not currently being used. The basic principle is the same as the crude ice-box calorimeter, but circulating water removes heat from the chamber and calculations are based on the flow rate, temperature changes, and other thermal parameters of the water. Direct calorimetry chambers are not used extensively now because of the high cost of construction and the availability of simpler and less expensive oxygen consumption methods.

The calculation of heat production from oxygen consumption is based on the relationship between the volumes of oxygen consumed and heat production. The relationship is illustrated by equation (1) from Brody (1945).

This equation shows that the oxidation of one mol of hexose $(C_6H_{12}O_6)$ requires 6 mols of O_2 , with the resulting production of six mols of CO_2 , 6 mols of H_2O_1 , and 678 calories of heat energy.

 $C_{6}H_{12}O_{6} + 6 O_{2} = 6 CO_{2} + 6 H_{2}O + 678 Cal$ (1) (180 g.) (134.4 lit.) (134.4 lit.) The heat production (678 calories) divided by the consumption of 134.4 liters of oxygen or the production of 134.4 liters CO_2 yields 5.047 calories (5.047 = 678/134.4). Thus 5.047 calories of heat are produced for each liter of O_2 consumed and each liter of CO_2 produced in carbohydrate oxidation. The oxidation of mixed fat results in the release of 4.69 calories per liter oxygen consumed, and 6.6 calories per liter carbon dioxide produced. For the oxidation of mixed protein, 4.82 calories are released per liter O_2 consumed, and 5.88 calories per liter CO_2 produced. The figures above indicate that the numbers of calories produced by oxidation varies with the carbohydrate, fat, and protein content of the food, so the proportion of carbohydrate, fat, and protein in the food should be known before calculating the heat production from either oxygen consumption or carbon dioxide production.

The amount of protein oxidized is determined from the urinary nitrogen excreted. This assumes that there is a constant quantity of nitrogen in the protein. If 16% of the protein is nitrogen, the protein catabolized is estimated by multiplying the urinary N by the ratio of total protein to nitrogen in the protein (100/16 = 6.25) (Equation 2)

Urinary N x
$$6.25$$
 = protein catabolized (2)

The example above is true for several animal proteins. Cereal proteins contain 17-18% N, so the conversion factor is 5.8 to 5.9. In practice, the assumption that protein contains 16% nitrogen is sufficient. The relative amounts of fat and carbohydrate oxidized are determined from the non-protein respiratory coefficient (R.Q.). The R.Q. is the ratio of mols or volumes of CO₂ produced to mols or volumes of O₂ consumed. Thus in equation (1), the R.Q. is equal to 1.0 because 6 $CO_2/6$ O₂ = 1.00; only carbohydrates had been consumed.

The R.Q. for mixed fats is 0.71, and for mixed protein, 0.81. These are averages only, as fatty acids vary in their R.Q. (short-chain fatty acids have an R.Q. nearer 0.8) and each protein and amino acid has its distinctive R.Q. The relative amount of heat produced by the oxidation of carbohydrates and fats can be computed from the R.Q., and these are presented in tabular form in Brody (1945:308-310), the source of information for the material discussed in the previous paragraphs.

The respiratory quotient does not always have the simplicity of interpretation implied in the foregoing discussions. In ruminants, large quantities of CO₂ are produced by the rumen bacteria, and this cannot be distinguished from the CO₂ resulting from internal respiration. The measurement of oxygen consumption and its multiplication by the caloric equivalent of oxygen is a satisfactory method for estimating heat production by the ruminant. Note in Table 1 that the calories per liter of oxygen cosumed ranged from 4.686 to 5.047. The mean value is about 4.86, and this occurs at an R.Q. of 0.85. The R.Q. of protein is about 0.82, so if one is satisfied with an approximation of mean values, the rate of heat production can be calculated by multiplying the liters of oxygen consumed by 4.82 to 4.85 without correcting for protein metabolism. This method is often used by comparative physiologists (Hoar 1966).

BASAL METABOLISM

The tabular format shown below includes columns for the information necessary for interpreting published results of basal metabolism tests. The first three items identify the conditions for a basal test. Completion of the blanks results in KCAL and KJOU being expressed on an absolute basis, and MBLM provides for comparisons of the results on a relative basis.

				Time	Weight		24-h	our	
		_		or	11		Dasal me	Laborism	
Reference	Species	Sex	Age	Year	Kg	IFMW	KCAL	KJOU	MBLM
-	_	-	-	-	-	-	-	-	·
_	-	-	-	-	-		_	_	-
-	-	-	-	-	_	-	-	-	-
-	-	_		-	_	_	-	-	_
-	-	-	_	-	_	_	-	_	-
_	_	_	_	-	_	_	_	-	_
	_								
-	-	-	-	-	-		-	-	-

FORMAT FOR COMPARING BASAL METABOLISM TESTS

Basal metabolism tests on white-tailed deer showed seasonal variations, with the lowest metabolism occurring when the deer were in winter coat and the highest when they were in summer coat (Silver 1968). Seasonal variations in basal metabolism may be expressed with a sine wave in the same manner that seasonal variations in weight were, showing seasonal metabolic rhythms as an animal goes from a winter low to a summer high in a gradual way. There is no general formula available yet for the expression of seasonal rhythms in metabolism. Results of curve fitting the data for white-tailed deer are given in the WORKSHEETS.

FASTING METABOLISM

Fasting metabolism measurements are expected to show seasonal variations, just as basal metabolism tests did. The differences between the two tests is that basal tests are for animals at rest, and fasting tests are uncorrected for activity. Both tests are conducted on post-absorptive animals in thermoneutral conditions.

A tabular format for comparing the results of fasting metabolism tests is shown on the next page.

· · · · · · · · · · · · · · · · · · ·				Time	Weight		24-1	hour	
				of	in		fasting m	etabolism	L
Reference	Species	Sex	Age	Year	Kg	IFMW	KCAL	KJOU	MBLM
_	-	_	-	_	_	-	· _	-	_
-	-	-	-	-	-	-	-	-	-
_	-	-	-	-	-	-	-	-	-
-	-	-	-	-	-	_	-	-	-
-	-	-	-	-	-	-	-	-	-
-	-	-	-	-	-	-	-	-	-
-	-	-	-	-	-	-	-	-	-

FORMAT FOR COMPARING RESULTS OF FASTING METABOLISM TESTS

LITERATURE CITED

Brody, S. 1945. Bioenergetics and growth. Reinhold Publ. Co., New York. 1023 pp.

Hoar, W. S. 1966. General and comparative physiology. Prentice-Hall, Inc., Englewood Cliffs, New Jersey. 815 pp.

REFERENCES, UNIT 2.1

CHAMBER MEASUREMENTS

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 53--6 679 685 odvi ambien temp effect, physic holter, jb; urban/ 1975 odvi energy metabolism, red dee brockway, jm; mal/ 1968 JPHYA 194-1 22 24 JWMAA 23--4 434 438 odvi basal metab, a pilot study silver, h; colovo/ 1959 JWMAA 33--1 204 208 odvi cage, metab, radioiso stud cowan, rl hartso/ 1969 JWMAA 33--3 490 odvi fasting metabolism silver, h; colovo/ 1969 4**9**8 JWMAA 35--1 37 odvi effect falli temp, heat pr silver, h; holter/ 1971 46

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JPHYA 194-1 22 24 ceel energy metabolism, red dee brockway, jm; malo 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 39--7 845 856 rata clima, metab, therm, infan hart,js; heroux,/ 1961 CJZOA 48--2 391 392 rata energy metab, barren groun mcewan eh 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 34--4 908 912 anam energ flux, water kinetics welsey,de; knox,/ 1970 JWMAA 37--4 563 573 anam energy metabolism, prongho wesley,de; knox,/ 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

obmo

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS				AUTHOR	RS-		YEAR
AJPHA	178-3	515	516	oram	meta	abolism	of	mountaí	goat	krog, h	n;	monson,m	1954

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEARAJAEA 13--1 144164dosh energy meta, new-born lamb alexander,g1962AJAEA 18--1 127136dosh fastng metab rate reln wt, graham,nm1967BJNUA 16--4 615626dosh fast metab, adul wether sh blaxter,k11962

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 22--1 3 24 caca energy metabolism, roe dee weiner, j 1977

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CHAPTER 7, WORKSHEET 2.1a

Basal metabolism of white-tailed deer

Measured basal metabolic rates of white-tailed deer show differences between winter and summer coat. These are summarized in Moen (1973:117) based on data in Silver (1968). The average multiple of base-line metabolism for the winter coat is 75.6 and for the summer coat, 84.4.

How can these limited data be used to represent a seasonal rhythm in basal metabolism of deer? Suppose the summer peak was reached on August 15, which is JDAY 227, and the winter low six months later, which is JDAY 45. The sine wave is again employed to determine a smooth transition from maximum to minimum, and a phase correction is needed to make JDAY 227 equivalent to 90° when sin 90 = +1.

The primary phase correction is 90 - (227)(0.9863) = -134 = PRPC. Therefore:

 $\sin[(227)(0.9863)]-134 = +1.0$, and $\sin[(45)(0.9863)]-134 = -1.0$

The equation for predicting the seasonal rhythm in basal metabolism per day (BAMD) is:

$$BAMD = [(84.4 + 75.6)/2] + \{sin [(JDAY)(0.9863)] - 134\} \{ [84.4 - 75.6] /2 \}$$

This simplified deviation of BAMD illustrates another use for the sine wave equation for smoothly correcting observed biological values over an annual cycle. The deviation is illustrative, and not meant to prove that basal metabolism values through the year are exactly these values.

Complete the calculations and plot the results below.

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LITERATURE CITED

- Moen, A. N. 1973. Wildlife ecology. W. H. Freeman & Co., San Francisco. 458 pp.
- Silver, H. 1968. Deer nutrition studies. In The white-tailed deer of New Hampshire, ed. H. R. Siegler. Survey Report No. 10, Concord: New Hampshire Fish and Game Department, pp. 182-196.

CHAPTER 7, WORKSHEET 2.1b

Fasting metabolism of white-tailed deer

The fasting metabolism of white-tailed deer has been measured by Silver et al. (1969) and the data recalculated and compiled in Moen (1973:118). Average heat production values in kcal per $W_{kg}^{0.75}$ are as follows:

Age	Sex	<u>Coat</u>	Heat production	MBLM
Adult	male	winter	95.6	1.37
Adult	male	summer	141.4	2.02
Adult	female	winter	97.5	1.39
Adult	female	summer	146.3	2.09
	yearlings	summer	130.6	1.87
	fawns	winter	90.1	1.29

The differences in MBLM between sexes are very slight. The values 1.38 = MBLM in winter coat and 2.06 = MBLM in summer coat could be used for both sexes. Using the procedures described in WORKSHEET 2.1a, derive a sine wave equation for fasting metabolism expressed as a multiple of base-line metabolism and plot the results below.



LITERATURE CITED

Moen, A. N. 1973. Wildlife ecology. W. H. Freeman & Co., San Francisco. 458 pp.

Silver, H. 1968. Deer nutrition studies. In The white-tailed deer of New Hampshire, ed. H. R. Siegler. Survey Report No. 10, Concord: New Hampshire Fish and Game Department, pp. 182-196.

CHAPTER 7, WORKSHEET 2.1c

Metabolism of a mountain goat

The metabolism of a $1 \ 1/2$ year-old male mountain goat was measured in February and March in a temperature-controlled chamber by Krog and Monson (1954).

The measured values are:

Weight	Ambient Temperature	Total Heat Production (kcal per 24 hour)	Kcal per LWKG ^{0.75}	MBLM
32 kg	20 to -20	1027		
	-30	1304		-
	-50	2362		

Complete the "kcal per $W_{kg}^{0.75}$ " column by dividing total heat production by $32^{0.75}$, and the MBLM column by dividing the previous answer by $(70)(32^{0.75})$. Note the MBLM values for later comparisons with seasonal rhythms in different species discussed in TOPIC 6 of this CHAPTER.

LITERATURE CITED

Krog, H. and M. Monson. 1954. Notes on the metabolism of a mountain goat. Am. J. of Physiol. 178:515-516.

UNIT 2.2: OUTDOOR MEASUREMENTS

It is difficult to get good direct measurements of the heat production of wild ruminants in chambers because of the extensive training required and the potentials for psychological effects, and it is impossible to measure heat production directly when they are ranging freely of course. How may the "cost of living" of free-ranging animals be quantified? Since it is impossible to measure ecological metabolism directly over 24-hour periods throughout the annual cycle, it must be calculated based on biological evidence for general patterns and magnitudes of metabolic rates of specific activity and productive functions.

Outdoor measurements may be made in several ways, including the use of respiratory masks, tracheal cannulae, calculations of the cost of movement of mass over distance, and by converting vital signs to estimates of metabolic rates. Closed circuit spirographic-masks, consisting of an oxygen spirometer which measures the rate of oxygen consumption, are one of the least expensive methods for measuring enerby expenditures, although not all animals are willing to wear the masks. Closed-circuit systems involve rebreathing of the same air, except that CO2 is absorbed by porous soda lime after passing through one-way valves. Open-circuit mask methods for animals involve the measurements of CO₂ increases larger and 02 decreases of directly inspired air. Gas analyzers are used to determine the amounts of oxygen consumed, and these measurements are converted to estimates of heat production.

The basic equations for determining metabolism using heart rates and activity patterns and heart rate to metabolism conversions have been described, and can be assembled into a sequence of calculations throughout the year to predict ecological metabolism throughout the year. While these calculations can be repeated for every day, 365 times for a year, that is unnecessary as 15-day intervals are sufficiently close to result in a smooth curve over the annual cycle.

REFERENCES, UNIT 2.2

OUTDOOR MEASUREMENTS

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY	WORDS	AUTHORS	YE/	AR
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JWMAA 20--3 221 232 odvi nutrient req, growth, antl french, ce; mcewa/ 1956 JWMAA 27--2 185 195 odvi rumen ferment, ener relati short, hl; 1963 JWMAA 33--3 482 490 odvi diges ener, win, mich does ullrey, de; youat/ 1969 JWMAA 34--4 863 odvi diges metab ener mich does ullrey, de; youat/ 1970 869 JWMAA 37--3 301 311 odvi nutrition requiremen, fawn thompson, cb; hol/ 1973 JWMAA 42--4 715 738 odvi seas rhyth, heart rat, met moen,an 1978 JWMAA 43--4 880 odvi predict energ, nitro reten holter, jb; urban/ 1979 888

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CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS				AUTHORS		YEAR
CJZOA	482	275	282	odhe	feed	l intake,	heat	prod,	yo	nordan,hc;	cowan/	1970
JWMAA	431	162	169	odhe	ene	rg requir	em fav	vn, wi	nte	baker,dl;	johnso/	1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBPAB 59A-1 95 100 ceel effic of utili diet energy sipson, am; webst/ 1978 CBPAB 61A-- 43 48 ceel oxyg use, calves, locomotn cohen, y; robbins/ 1978

JWMAA 43--2 445 453 ceel energy expenditure, calves robbins, ct; cohe/ 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BPURD 1---- 335 339 rata meth meas ener exp, unrstr young, ba; mcewan, 1975

CJZOA 56--2 215 223 rata energ expnd, walk, rd, tund white, rg; yousef, 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR PAANA 7---- 335 341 dosh co2, index ener expen, 1mb white,rg; leng,ra 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARPNUSA 37--1 1319altern meth, larg anim cal brockway, jm1978

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CHAPTER 7, WORKSHEET 2.2a

Caloric value of oxygen at different respiratory quotients

Calculations of heat production based on oxygen consumed are made by multiplying the liters of oxygen consumed by the caloric value of O_2 at the respiratory quotient (R. Q. = REQO) characteristic of the diet of the animals at the time of measurement.

The abbreviated table below, based on data in Brody (1945:310) illustrates differences in the kcal per liter of oxygen at REQOs ranging from 0.70 to 1.00.

REQO	Kcal per liter 0 ₂
0.70	4.686
0.75	4.729
0.80	4.801
0.85	4.863
0.90	4.924
0.95	4.985
1.00	5.047

The table in Brody gives the kcal per liter 0_2 for R. Q. from 0.70 to 1.00 at 0.01 intervals. While the table covers more than half a page vertically, curve-fitting with linear regression reduces the tabular information to a simple equation. Thus, the kcal per liter of 0_2 (KCLO) at different respiratory quotients (REQO) can be calculated:

KCLO = 3.818 + 1.229 REQO; $R^2 = 1.00$

LITERATURE CITED

Brody, S. 1964. Bioenergetics and growth. Hafner Publishing Co., Inc. New York. 1023 pp.

Chapter 7 - Page 32aa

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TOPIC 3. ESTIMATES OF ENERGY COSTS FOR PRODUCTION

Production results in the formation of new individuals (reproduction) and the deposition of new tissue by the individual (growth). Both of these productive functions are essential for the growth of the population. The growth rates of individuals often become indicators of the reproductive potentials of populations. Populations with smaller than average individuals will likely have lower than average reproductive rates because when requirements for a high rate of growth are not met, resources are not available for a high rate of reproduction. In white-tailed deer, for example, first conception becomes probable at a weight of about 40 kg. If a fawn does not attain this weight by its first winter, it is unlikely to re-. produce until it is a yearling.

Inadequate nutrition during the gestation period may result in abortion, resorption of fetal tissue, or reduced fetal growth during the final stages of gestation. When fetal growth is reduced, the young weigh less at birth and have decreased chances of survival.

Inadequate maternal nutrition may impair lactation, possibly reducing the growth rate of the nursing young. Lactation is the single greatest sustained energy and protein requirement of mammals, and the major factor contributing to the high metabolic rate in the summer. The timing of this high cost is ecologically sound, however, as offspring are produced in late spring, just in time for the nursing dam to have access to a range with new growth as a source of nutrients for the costly process of lactogenesis.

While forage resources are more abundant and often quite digestible in the summer, this does not suggest that nutrients are unlimited on a green and growing range. Wild ruminants forage rather selectively, and will not consume any plant just because it is present. In fact, they often exhibit a high degree of selection not only for a particular plant species, but also for plant parts such as flowers. Ingestion of selected plant parts contributes to a higher quality diet (Whittemore and Moen 1980).

When nutrients are ingested in sufficient quantities to result in the production of new tissue, the nutrient balance is positive. The new tissue may be milk, which is made available to the dependent offspring whose growth may be at the expense of the body tissue of the lactating female. Weight curves have been calculated (CHAPTER 1) showing that annual minimum weights of lactating females occur at about the time of peak lactation.

LITERATURE CITED

Whittemore, S. and A. N. Moen. 1980. Composition and digestibilities of various forages of the white-tailed deer. Can. J. Anim. Sci. 60:189-192.

UNIT 3.1: THE ENERGY COST OF BODY GROWTH

Growth of wild ruminants that have not yet reached their full adult size occurs in the spring, summer and fall. Unbred yearlings may grow from spring through fall. Nursing young grow in the summer, and reproducing females will grow in late summer and fall as lactation demands are reduced. Antler and horn growth occurs in mature males from spring to early fall. The amount of growth is partially related to body weight and growth, so it is also related to range conditions, and partially related to the genetic potentials of the individual.

The cost of body growth is equal to the amounts of nutrients deposited plus the cost of synthesizing the tissue. These costs are directly dependent on tissue composition. Suppose that an animal gained 10 kg over a 100-day period. Suppose that 8 kg of this increase was water, 1.5 kg fat, and 0.5 kg protein. The energy increment in the animal's body is equal to 1.5 kg fat times 9,500 kcal per kg = 14,250 plus 0.5 kg protein times 5,500 kcal = 2750 kcal, totaling 17,000 kcal of tissue energy. This is 170 kcal per day.

One very important parameter must also be considered, and that is the efficiency of growth. If the efficiency were 50% in the example above, then the energy cost of tissue deposition would be equal to twice the energy in the tissue and the total cost would be $2 \times 170 = 340$ kcal. Efficiency of growth is related to physiologic age. But body composition is related to body weight and age, so the older and larger mammal, gaining less but depositing more fat, is depositing tissue with a higher caloric density. Thus while efficiency of growth goes down, changes in efficiency are reflected in changes in body composition. Examples illustrating how the cost of growth can be calculated on the basis of chemical composition and the cost of tissue synthesis are given in a WORKSHEET.

REFERENCES, UNIT 3.1

THE ENERGY COST OF BODY GROWTH

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JWMAA	203	221	232	odvi	nutrit req, growth, antler french, ce; mcewe/	1956
JWMAA	373	301	311	odvi	energ requiremnts of fawns thompson, cb; hol/ 1	1973
JWMAA	392	355	360	odvi	milk consumpt, weight gain robbins, ct; moen,	1975
JWMAA	394	684	691	odvi	uterin comp, grow, pregnan robbins, ct; moen,	1975
NAWTA	22	119	132	odvi	nutrient requirements of mcewen,lc; frenc/ 3	1957
PAARA	209-1	1	11	odvi	eff feed restric, antl dev long,ta; cowan,rl	1959

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR NAWTA 22--- 179 186 odhe feed requirem, grow, maint cowan, imct; wood/ 1957 CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR NATUA 215-- 1134 1136 ceel comparitive nutr, pregnanc payne, pr; wheele/ 1967 CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR PNUSA 27--2 129 138 alal compar nut in preg, lactat payne, pr; wheele/ 1968 . CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR CJZOA 48--2 391 392 rata energ metab, bar gro carib mcewan, eh 1970 CJZOA 48--5 905 913 rata seas chan, energ, nit inta mcewan, eh; whiteh 1970 CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR

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CHAPTER 7, WORKSHEET 3.1a

The energy cost of body growth

Experiments designed to measure the cost of body growth are not as easily completed on wild ruminants as on domestic ruminants. Estimates of the cost of growth may be made by determining change in weight over a specified time period, changes in body composition in relation to weight, and the caloric equivalents of the tissue involved. The steps are:

- 1. Subtract final weight from initial weight.
- 2. Determine body composition at final and initial weights and the amounts of each of the following:
 - a. Waterb. Fatc. Proteind. Ash
- 3. Multiply the change in the fat and protein components by their approximate combustion values (Brody 1945:35).

Fat = 9.5 kcal per gm Protein = 5.7 kcal per gm

- Add the energy contents of the fat and protein gain together and divide by the number of days between the initial and final weight used in Step 1.
- 5. Multiply the answer in Step 4 by the reciprocal of the efficiency (50% efficiency: 1/0.50 = 2; 33% efficiency: 1/0.33 = 3).

Specific equations for the energy cost of growth may be compiled from the weight equations and chemical composition equations in PART I.

LITERATURE CITED

Brody, S. 1964. Bioenergetics and growth. Hafner Publishing Co., Inc., New York. 1023 pp.

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CHAPTER 7, WORKSHEET 3.1b

Total energy content of white-tailed deer as a function of ingesta-free weight

The total energy content of white-tailed deer in relation to ingesta-free weight has been determined by Robbins et al. (1974), simplifying calculations of the cost of growth. Only slight differences were observed in males and females so a single equation may be used to estimate total caloric content in kcal (TCCK).

 $TCCK = e^{1.3720} \ln IFWK - 0.5876$

Using this equation and the seasonal weight equations in CHAPTER 1, the caloric content can be quickly determined after initial and final weights have been determined. Set up some weight differences based on the calculations in CHAPTER 1, UNIT 1.4, determine the daily change in energy content, and estimate the energy cost of these changes. Label and plot the results below. Differences in efficiencies may be represented as a family of curves.

LITERATURE CITED

Robbins, C. T., A. N. Moen and J. T. Reid. 1974. Body composition of white-tailed deer. J. Anim. Sci. 38(4):871-876.

UNIT 3.2: THE ENERGY COST OF HAIR GROWTH

The cost of hair growth may be determined in a manner similar to that for body growth. Time-periods of hair growth, amounts of hair, and chemical composition of the hair need to be known when making these calculations. These parameters were discussed in CHAPTER 2; calculations are illustrated in the WORKSHEETS.

REFERENCES, TOPIC 3.2

THE ENERGY COST OF HAIR GROWTH

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 38--4 871 876 odvi body composition of white- robbins,ct; moen, 1974

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 CODEN VO-NU BEPA ENPA ANIM KEY WORDS-------- AUTHORS-------- YEAR

 JANSA 45---4 826 831 doca hair indic calor-prot stat haaland,g1; mats/ 1977

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CHAPTER 7, WORKSHEET 3.2a

The energy cost of hair growth of white-tailed deer

The energy cost of hair growth may be estimated by determining the weight of the hair coat, chemical composition, and caloric values of the hair to give the energy content of the hair coat. The daily cost may be estimated by multiplying the energy content by the reciprocal of the efficiency of hair growth and dividing by the numbers of days during which the hair growth occurred. The formula may be written as:

 $ECHD = \{ [(HAWG)(FACO)(9.74) + (HAWG)(PRCO)(5.61)] 1/PEHG \} / DHGD \}$

where ECHD = energy cost of hair growth per day

HAWG = hair weight in grams

FACO = fat content expressed as a decimal fraction = 0.098

- PRCO = protein content expressed as a decimal fraction = 0.890

DHGD = duration of hair growth in days

The caloric values and fat and protein contents are taken from Robbins et al. (1974) HAWG is determined from WORKSHEET 1.3b in CHAPTER 2. PEHG is a variable, and DHGD is dependent on the biochronology of the species or population group being considered.

Calculate and plot HAWG and ECHD on the next page, labeling the y-axis with the appropriate units of measurement. The effects of different physiological efficiencies of hair growth may be expressed with a family of curves.

How much does variation in PEHG contribute to variation in ecological metabolism? This question may be answered after all of the maintenance, activity, and production costs have been considered and ecological metabolism per day (ELMD) determined. Completion of the WORKSHEETS in this CHAPTER will enable the user to evaluate the cost of any process in relation to the total.

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Robbins, C. T., A. N. Moen and J. T. Reid. 1974. Body composition of white-tailed deer. J. Anim. Sci. 38(4):871-876.
UNIT 3.3: THE ENERGY COST OF GESTATION

Accelerating requirements for gestation must be met in the last onethird of pregnancy. This acceleration generally coincides with the period of renewed plant growth, although differences in the lengths of winters result in different times of renewed plant growth from year to year. A latearriving spring may have considerable effect on the condition of pregnant females during the last one-third of pregnancy (see Moen 1978). Conception occurs in the fall, and its timing is not related to the termination of winter several months later, of course. High conception rates may be followed by reduced birth rates if winter conditions are extended.

Increases in the requirements for gestation are related to increases in fetal growth and in the energy content of the uterus and associated reproductive tissues. The trends are illustrated below; curve-fitting of actual data may result in logarithmic, exponential, or power curves. Costs for associated uterine tissues increase in the same way, and are added to the costs of fetal growth to determine the total energy cost of gestation.



Calculations of the cost of gestation are made in the WORKSHEETS, with equations given for white-tailed deer and proportionalities described for other species.

LITERATURE CITED

Moen, A. N. 1978. Seasonal changes in heart rates, activity, metabolism, and forage intake of white-tailed deer. J. Wildl. Manage. 42(4):715-738.

REFERENCES, UNIT 3.3

THE ENERGY COST OF GESTATION

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AMNAA 43--3 650 666 odvi fetal develop, white-taile armstrong, ra 1950 JOMAA 47--2 266 odvi endoc glan, seas, sex, age hoffman, ra; robin 1966 280 JWMAA 33--3 482 490 odvi dig energ req, wintr, does ullrey, de; youat/ 1969 JWMAA 34--2 383 388 odvi morphol develop, aging, fe short, c 1970 JWMAA 34--4 863 odvi dig, met ener, wintr, does ullrey, de; youat/ 1970 869 JWMAA 39--4 684 odvi uterin comp, grow, pregnan robbins, ct; moen, 1975 691 JWMAA 42--4 715 738 odvi seas chan, heart rate, act moen, an 1978 NAWTA 28--- 431 443 odvi nutrit, growth, fetal, faw verme, lj 1963 NAWTA 22--- 119 132 odvi nutrient requirements of w mcewen, lc; frenc/ 1957 VJSCA 23--3 116 116 odvi aspects of early pregnancy scanlon, pf; murp/ 1972 VJSCA 24--3 112 odvi ovula, pregnan, lactat dee scanlon, pf; murp/ 1973 112 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--1 123 132 odhe development, fetal period ommundsen,p; cowa 1970 JWMAA 23--3 295 304 odhe embryo, fetal development hudson, p; browman 1959 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 47--6 1418 1419 ceel sexual dimorphism, fetuses retfalvi,1 1969 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR PNUSA 27--2 129 138 alal compar nut in preg, lactat payne, pr; wheele/ 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--5 905 913 rata seas chan, energ, nit inta mcewan,eh; whiteh 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 46--3 524 525 ovca fetal measure, milk charac forrester,dj; sen 1965 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AMNTA 114-1 101 116 ungu matern repro effort, fetal robbins, ct; robbi 1979

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CHAPTER 7, WORKSHEET 3.3a

The energy cost of gestation in white-tailed deer

The energy cost of gestation is equal to the energy deposited in the gravid uterus each day times the efficiency of the physiologic processes involved. The energy content of the gravid uterus has been determined by Robbins and Moen (1975), and may be used to calculate the energy cost of gestation with the following equation.

 $ECGD = (e^{0.2803} + 0.0282 \text{ DIGE})(1/PEGE)$

where ECGD = energy cost of gestation per day, DIGE = days into gestation, and PEGE = physiological efficiency of gestation

Plot ECGD below, expressing PEGE as a family of curves again.



LITERATURE CITED

Robbins, C. T. and A. N. Moen. 1975. Uterine composition and growth in pregnant white-tailed deer. J. Wildl. Manage. 39(4):684-691.

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CHAPTER 7, WORKSHEET 3.3b

The energy costs of gestation in several species of wild ruminants

The energy costs of gestation have been estimated for several species of wild ruminants in Robbins and Moen (1975) by considering the costs for these species to be proportional to the costs for deer. Fetal weights and lengths of the gestation period vary, of course.

The equations for estimating ECGD for different species, modified modified from those in Robbins and Moen (1975), are listed below.

Species	Equation	LEGP
ceel:	$ECGD = (e^{0.0193} DIGE - 1.7938)(1/PEGE)$	260
alal:	$ECGD = (e^{0.0204} DIGE - 1.7358)(1/PEGE)$	245
rata:	$ECGD = (e^{0.0226} DIGE - 1.6198)(1/PEGE)$	220
anam:	$ECGD = (e^{0.0209} DIGE - 1.7140)(1/PEGE)$	240
bibi:	$ECGD = (e^{0.0173} DIGE - 1.9036)(1/PEGE)$	290
ovca:	$ECGD = (e^{0.0334} DIGE - 1.2443)(1/PEGE)$	150
oram:	$ECGD = (e^{0.0278} DIGE - 1.4281)(1/PEGE)$	180

Complete the numbering of the x and y axes and plot the energy costs of gestation on the grid on the next page. Note again that different physiological efficiencies may be expressed a a family of curves.

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LITERATURE CITED

Robbins, C. T. and A. N. Moen. 1975. Uterine composition and growth in pregnant white-tailed deer. J. Wildl. Manage. 39(4):684-691.

Chapter 7 - Page 44bb

ECGD

CHAPTER 7, WORKSHEET 3.3c

The energy costs of gestation in relation to days into gestation and fetal weight at birth

The two previous WORKSHEETS have included equations for selected species with averge fetal weights and gestation periods built into the equations. An alternative equation in Robbins and Moen (1975) may be used to provide the flexibility needed when applying the proportional costs to any species. The formula for calculating the energy contents of the uterus and contents (kcal) per fetal weight at parturition (kg) is:

$$ECGD = \left\{ [e^{(2.1548 + 5.01 (HIGR DIGE/LEGP)]} - [e^{(2.1548 + 5.01 (LOWR DIGE/LEGP)}] \right\} (BIWK)/$$

$$(HIGR DIGE - LOWR DIGE)(1/PEGE)$$
where ECGD = energy cost of gestation per day
HIGR DIGE = higher number of days into gestation
LOUP DIGE = heaver number of days into gestation

LOWR DIGE = lower number of days into gestation LEGP = length of the gestation period BIWK = birth weight in kg PEGE = physiological efficiency of gestation

Make the calculations for selected species with different birth weights and evaluate the daily costs associated with gestation. The results may be plotted below.



Chapter 7 - Page 44c

LITERATURE CITED

Robbins, C. T. and A. N. Moen. 1975. Uterine composition and growth in pregnant white-tailed deer. J. Wildl. Manage. 39(4):684-691.

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UNIT 3.4: THE ENERGY COST OF MILK PRODUCTION

Milk production has been mentioned several times already as a very costly biological process. One might come to this conclusion simply by observing the condition of lactating females; they are often in the worst condition of the year during the lactation period. The weight pattern shows minimum weights during the lactation period, and it is well known that if milk production of dairy cattle is to remain high, a high quality ration must be fed in large quantities.

If milk production is the most costly sustained biological process there is, then it is imperative that ways be found to quantify the cost. Direct measurements of milk production might be made by actually milking the animal, but this is difficult because wild ruminants are not inclined to stand while being milked. Further, milk production is partly dependent on the demand for milk, and this varies with the suckling young as they grow and their diet includes increasing amounts of forage. Since the demand of the young for milk cannot be duplicated by hand-milking the female, estimates of milk production obtained in this way will not be accurate.

Animals in captivity might be weighed before and after nursing, and either the dam's weight loss or the sucklings' weight gain measured. Scales sufficiently large to weigh the dam before and after nursing are not accurate enough to weigh the difference in her weights, however. The suckling animal often urinates and defecates while nursing--the dam usually licks the anal region to stimulate release--so accurate measurements of milk intake are difficult to make. Furthermore, it is difficult to be present to weigh the animals each time the young nurse, which could be several times a day.

The only practical way to determine milk production of wild ruminants is by mathematical means. There is a fundamental relationship that can be considered the basis for mathematical calculations, and that is that the milk production of the females of a species will be adequate to supply that portion of the nutrients derived from milk by the young of that species to meet their total metabolic costs for growth and activity. The amount of nutrients supplied by the milk varies as the rumen develops and the young become increasingly more dependent on forage for nutrients. This mathematical approach, then, requires a knowledge of three things, including the rate of rumen development so that the ratio of milk nutrients: forage nutrients is known, the chemical composition of the milk, and the ecological metabolism of the nursing young. These were discussed in earlier chapters, and they can be assembled here in WORKSHEETS to calculate the cost of milk production.

REFERENCES, UNIT 3.4

THE ENERGY COST OF MILK PRODUCTION

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JRPFA 37--1 67 84 cerv composi milk, cerv species arman,p; kay,rnb/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 24--4 439 441 odvi rearing, breedi fawns, cap murphy,da 1960 JWMAA 25--1 66 70 odvi deer milk, substitute milk silver,h 1961 JWMAA 29--1 79 odvi comp milk, bld nursng doe, f youatt, wg; verme/ 1965 84 JWMAA 37--3 301 odvi energ requiremnts of fawns thompson, cb; hol/ 1973 311 JWMAA 39--2 355 odvi milk consumpt, weight gain robbins, ct; moen, 1975 360

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CAFGA 37--2 217 218 odhe composit, deer milk, calif hagen, hl 1951 JOMAA 36--3 473 474 odhe mule deer milk browman, 1g; sears 1955 JOMAA 58--3 420 423 odhe changes nutri compos, milk mueller,cc; sadle 1977 JWMAA 20--2 212 214 odhe postnat grow, comp of milk kitts,wd; cowan,/ 1956 JWMAA 44--2 472 odhe milk yield, black-tailed d sadleir, rmfs 478 1980

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR JRPFA 37--1 67 84 ceel comp, yield of milk, red d arman,p; kay,rnb/ 1974 JWMAA 40--2 330 335 ceel nutrition, gestat, reprodu thorne,et; dean,/ 1976 ZTTFA 31--5 227 238 ceel comp, milk, red deer, pt 1 brueggemann,j; d/ 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--2 213 215 alal gros comp milk, fat, minrl cook, hw; rausch,/ 1970 JWIDA 12--2 202 207 alal milk, hair, element relati franzmann, aw; fl/ 1976 NCANA 101-1 227 262 alal review of energy requireme gasaway, wc; coady 1974 PNUSA 27--2 129 138 alal compar nut in preg, lactat payne, pr; wheele/ 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 45--6 1101 1106 rata milk, gros comp, fat, prot hatcher,vb; mcew/ 1967 CJZOA 49--4 443 447 rata measurement milk intake, r mcewan,eh; white/ 1971 JDSCA 57-11 1325 1333 rata milk comp chang, grazing r luick,jr; white,/ 1974 ZOBIA 32--6 746 750 rata alal, electroph, milk prot shubin,pn; turub/ 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CAFGA 37--2 217 218 anam odhe, compos milk, compars hagen,hl 1951 JWMAA 34--4 908 912 anam enrgy flux, water kinetics wesley,de; knox,/ 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 36--2 305 308 bibi the lipids in bison bison wilbur,cg; gorski 1955

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CAFGA 41--2 131 143 ovca rearing lambs in captivity deming,ov 1955 CJZOA 43--5 885 888 ovca milk, gross comp, fat cons chen,ech; blood,/ 1965

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--4 629 633 ovda milk, stage lact, composit cook, hw; perarso/ 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CJZOA 34--6 569 571 obmo gross composition of milk tener, js 1956 CJZOA 48--6 1345 1347 obmo gros comp, ftty acid, minrl baker, be; cook, h/ 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CJZOA 47--1 5 8 oram gross compos, fat acid con lauer, bh; blood, / 1969 CJZOA 47--2 185 187 oram miner const, milk, arctic luer, bh; baker, b 1969

CHAPTER 7, WORKSHEET 3.4a

The energy cost of milk production in white-tailed deer

The energy cost of milk production may be estimated if the quantity of milk being produced is known. Estimates of the latter may be made with an equation in Moen (1973:355), modified to include new terms and parameters. The formula is:

MPGD = [(MBLM)(70)(MEWK)] [(PNDM)(1/NECM)/GENM]

where MPGD = milk production in gms per day MBLM = multiple of base-line metabolism MEWK = metabolic weight in kg PNDM = percent nutrients derived from milk = [1.136 - 0.045 LWKG] NECM = net energy coefficient for milk GENM = gross energy in the milk

The formula can be used only if numerical values for each of the paramaters are available. The first one needed, MBLM, has not been discussed in the ecological context yet (see TOPIC 6). The MBLM of resting fawns was usually between 2.10 and 2.25 (see CHAPTER 7, WORKSHEET 1.1a). The MBLM for ecological metabolism of suckling fawns is discussed in CHAPTER 7, UNIT 6.1. In the absence of a derived MBLM, MBLM = 2.5 may be used as a first approximation.

MEWK is the metabolic weight, or $IFWK^{0.75}$, where IFWK is the ingesta-free-weight of the fawns. Growth and weights were discussed in CHAPTER 1.

PNDM is an expression for determining the percent nutrients derived from milk, a function of rumen development. Derivation of this equation is discussed in Moen (1973:342).

NECM is an expression of gross to net energy efficiency, including both the digestible energy and metabolizable energy coefficients.

GENM, the gross energy in the milk, may be measured directly or determined from the chemical composition of the milk, especially fat percent (FATP). FATP may be calculated with an equation derived from data in Youatt et al. (1965).

FATP = $1.036 e^{(-1.0117)(DILA)}$; $R^2 = 0.995$

where FATP = fat percent and DILA = days into lactation.

Using the equation in CHAPTER 2, WORKSHEET 2.2b, p. 46b, GENM may be estimated from FATP. MPGD may then be calculated by substituting these numbers and equations in the formula near the top of this WORKSHEET. A grid is provided on the next page for plotting the results.

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TOPIC 4. ESTIMATES OF THE ENERGY COSTS OF ACTIVITIES

Every activity that an animal participates in has an energy cost associated with it. The movement of any mass for any distance involves work, and this is as true of wild ruminants as it is of man-made machines. Work is also involved in maintaining posture; muscles develop a tonus that permits animals to hold their heads erect, their ears in an alert position, stand, and other postures. While the holding of these postures my not involve motion, work is still being performed as the body is held in place against the force of gravity.

The energy costs of activities such as bedding, standing, walking, foraging, and running can be quantified with the use of the principles of physics involving the movement of mass over distance, and they may be calculated from oxygen consumption measurements. There are differences in efficiencies at different rates of movement, but such details have been analyzed for only two species of wild ruminants.

The energy costs of five different activities are discussed in the units that follow, with worksheets for calculating the costs for use in time and energy budget calculations.

REFERENCES, TOPIC 4

ESTIMATES OF THE ENERGY COSTS OF ACTIVITIES

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS------ AUTHORS/EDITORS-- YEAR

aubo acpr loen 545 vert scale effects in locomotio pedley, tj, ed 1977

UNIT 4.1: THE ENERGY COST OF BEDDING

The energy costs of bedding may be approximated with the measurements of resting metabolism in relation to basal metabolism. This ratio provides a first approximation that can be conveniently applied to any animal, especially when evaluated in relation to base-line metabolism. There are differences due to posture, such as an erect head posture or a curled-up posture, but these details have not yet been discerned by experimentation.

The energy cost of bedding is a part of basal metabolism measurements. Animals then are at rest, and in a post-absorptive state in thermoneutral conditions. The measured heat production under basal conditions includes no heat of fermentation or heat of nutrient metabolism due to digestive processes, or extra heat production due to thermogenesis.

Resting metabolism of reindeer (rata) has been measured by White and Yousef (1978). Resting metabolism of two reindeer used in walking experiments varied markedly. A group of four adult females taken from late summer grazing had a resting metabolism of 125 LWKG^{0.75} per day.

It is very important to realize that the resting metabolism of free-ranging animals also includes the costs of productive processes in progress. Thus the measured cost while resting is not the costs of resting alone.

Basal metabolism references and worksheets are given in UNIT 2.1, with WORKSHEET 4.1a in this UNIT illustrating how the activity costs of bedding can be calculated as a multiple of base-line metabolism.

LITERATURE CITED

White, R. G. and M. K. Yousef. 1978. Energy expenditure in reindeer walking on roads and on tundra. Can. J. Zool. 56(2):215-223.

REFERENCES, UNIT 4.1

THE ENERGY COST OF BEDDING

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY	WOE	RDS				AUTHORS	YEAR
ECOLA	571	192	198	odvi	ener	gy	cons	ervat	io,	winter	moen,an	1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JPHYA 194-1 22 248 ceel energy metabolis, red deer brockway, jm; malo 1967 JWMAA 43--2 445 453 ceel energy expendi, elk calves robbins, ct; cohe/ 1979 567 ceel postur, activ, metab, cold gates,c; hudson,r 1979 JWMAA 43--2 564 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR alal CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--2 391 392 rata energ metab, jar gro carib mcewan, eh 1970 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJPHA 178-3 515 516 oram metabolism of mountai goat krog,h; monson,m 1954 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS---- YEAR JTBIA 49--2 345 362 mamm model, est metab, rest, ac wunder, ba 1975

CHAPTER 7, WORKSHEET 4.1a

Estimates of the energy cost of bedding

The energy cost of bedding may be considered equal to basal metabolism, because basal metabolism test results are expressed for animals at rest. The heat production of the free-ranging animal is expected to be higher, of course, since the heat of fermentation and heat of nutrient metabolism are present.

Calculations of the cost of bedding are easily made by multiplying base-line metabolism by the appropriate MBLM. If basal metabolism is considered equal to be the interspecific mean of 70 $W^{0.75}$, then the energy cost of bedding per day (ECBD) is:

$ECBD = BMBL (70)(IFWK^{0.75})(FTBD)$

where ECBD = energy cost of bedding per day, BMBL = bedded multiple of base-line metabolism, IFWK = ingesta-free weight in kg, and FTBD = fraction of time bedded per day.

If basal metabolism is greater than the interspecific mean, it is still convenient to express ECBD as a multiple of base-line metabolism. Using the average value of basal/base-line ratio for deer, MBLM = 80/70 =1.1 (see CHAPTER 7, UNIT 2.1). The equation for seasonal rhythms in basal metabolism expressed as MBLM may also be used; this is given in CHAPTER 7, UNIT 6.1.

Note that the fraction of time bedded per day (FTBD) is given in the equation above The intitial MBLM calculation is for a 24-hour period so FTBD, based on time budgets discussed in PART II, is used to allocate the portion of the day spent in bedded activity.

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UNIT 4.2: THE ENERGY COST OF STANDING

The energy costs of standing have been measured for several species, both ruminants and non-ruminants. The cost of standing above lying is about 9% in man, cattle, and sheep (Crampton and Harris 1969:151). An increase of 9.7% in oxygen consumption was observed in a reindeer when standing compared to lying (White and Yousef 1978). Elk calves had standing metabolic rates of 137 to 161 LWKG^{0.75} per day (Robbins et al. 1979). These are more than 9% higher than the expected basal rate, but it must be emphasized that the energy costs measured while standing also include the costs of productive processes. The total measured cost while standing is not only the cost of standing.

The energy costs of standing may be expressed as multiples of base-line metabolism. The multiple for man, cattle, and sheep (9% increase) would be 1.1. The costs of standing for domestic ruminants under controlled experimental conditions may be less than the costs for wild ruminants since standing by the latter may be accompanied by a state of alertness that contributes to the metabolic cost while standing. Some of the cost estimates for standing discussed by Robbins et al. (1979:451-452) seem unreasonably high; increase of more than 60% must be due to some experimental effects. The cost of standing should be considered as a physical cost, separate from the costs of alertness while standing.

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- White, R. G. and M. K. Yousef. 1978. Energy expenditure in reindeer walking on roads and on tundra. Can. J. Zool. 56(2):215-223.

REFERENCES, UNIT 4.2

THE ENERGY COST OF STANDING

SERIALS

CODEN	VO-NU	BEPA	ENPA	AN IM	KEY WORDS AUTHORS	YEAR
ECOLA	571	1 92	198	odvi	energy conservatio, winter moen,an	1976
JWMAA JWMAA	342 424	431 715	439 738	odvi odvi	wint feed patterns, penned ozaga,jj; verme,l seas chan, heart rate, act moen,an	1970 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JPHYA 194-1 22 248 ceel energy metabolis, red deer brockway,jm; malo 1967 JWMAA 43--2 445 453 ceel energy expendi, elk calves robbins,ct; cohe/ 1979 JWMAA 43--2 564 567 ceel postur, activ, metab, cold gates,c; hudson,r 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--2 391 392 rata energ metab, bar gro carib mcewan,eh 1970 CJZOA 56--2 215 223 rata energy expend, walk, tundr white,rg; yousef, 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CHAPTER 7 - WORKSHEET 4.2a

Estimating the energy cost of or while standing

Estimates of the energy cost of standing may be made by using MBLM for standing as the measure of the cost of standing. If this MBLM is applied to free-ranging animals, it should either contain concomitant costs of production, or the value of MBLM should be supplemented by additional costs of production. The important point is that a distinction must be made when calculating the costs of standing or while standing.

The costs of standing may be expressed with MBLM = 1.1 or slightly higher. The costs while standing, based on measurements of wild ruminants on ad libitum diets, were as high as 2.3, based on data (161/70 = 2.3) in Robbins et al. (1979:447). If the former is used, production costs are to be added. If the latter, they are a part of the direct measurement.

Another necessary parameter when calculating the cost of standing is the fraction of time spent standing each day (FTSD). This is necessary because the base-line metabolism approach is on a 24-hour basis. Thus, the formula for determining the energy cost of standing per day is:

 $ECSD = (SMBL)(70)(IFWK^{0.75})(FTSD)$

where ECSD = energy cost of standing per day,

SMBL = standing multiple of base-line,

IFWK = ingesta-free weight in kg, and

FTSD = fraction of time standing per day.

Since there are variables in the formula dependent on the species, weights, time of year, and other factors unique to the situation being evalauted, equations are left for you, the user, to derive. An unlabeled grid is provided for plotting results.

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LITERATURE CITED

Robbins, C. T., Y. Cohen, and B. B. Davitt. 1979. Energy expenditure by elk calves. J. Wildl. Manage. 43(2):445-453.

Chapter 7 - Page 54aa

UNIT 4.3: THE ENERGY COST OF WALKING

Walking, defined as locomotion resulting when no more than two of the hooves are off the ground at one time, involves the movement of mass over distance at a fairly slow rate of speed. Walking on the level is composed of movement in one plane (the horizontal), and walking uphill involves movement in two planes (horizontal and vertical).

The energy cost of walking may be calculated if the mass Horizontal. of the animal and the distance traveled are known. Equations for different species were evaluated and summarized in Moen (1973:356). More recently, direct measurements of oxygen consumption by adult reindeer and elk calves have been made by White and Yousef (1978) and Robbins et al. (1979), respectively. Oxygen consumption of reindeer walking on the horizontal was 14% higher than estimated for several smaller mammals by Taylor et al. (1970). Energy expenditures of elk calves were also higher than that predicted with equations in Taylor et al. (1970). The Taylor estimates should not be considered an interspecies mean, however, since too few animals and species were represented. Robbins also reported that the energy expenditure was a direct linear function of the speed of travel, as was oxygen consumption by adult female reindeer (White and Yousef 1978).

Vertical. The vertical component of the topography adds greatly to the cost of locomotion. Measurements have not been made on species which live on rugged topography, such as mountain goats, and those which live on more level terrain, such as pronghorn. The reindeer and elk experiments mentioned above also included measurements on different gradients. The energy cost of climbing was determined for reindeer by measuring oxygen consumption on the incline and subtracting the consumption measured on the horizontal. The energy cost of descent was determined in the same way, resulting in an estimate of locomotion downhill that was less than that on the horizontal. A similar relationship was observed for the elk calves.

The costs of locomotion at different speeds and on different gradients is of ecological interest because of potential energy costs associated with activities on different topographies. Patterns of energy conservation are discussed for white-tailed deer in Moen (1976). The extent of such differences may be estimated by calculating the costs of movements of different speeds and distances by species on different topographies, and by quantifying the behavior patterns of the species and calculating the daily costs of all activities. While the cost of movement may be more costly on rugged topography, it may be compensated for by adjustments in the time spent in different activities.

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- White, R. G. and M. K. Yousef. 1978. Energy expenditure in reindeer walking on roads and on tundra. Can. J. Zool. 56(2):215-223.

REFERENCES, UNIT 4.3

THE ENERGY COST OF WALKING

SERIALS

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS AUTHORS	YEAR
ECOLA	571	192	198	odvi	energy conservatio, winter moen,an	1976
JWMAA	424	715	738	odvi	seas chan, heart rate, act moen,an	1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 23-22 365 370 ceel energy cost of locomotion gates,c; hudson,r 1978 CBPAB 61A-1 43 48 ceel oxy util, horiz, vert loco cohen,y; robbins/ 1978 JWMAA 43--2 445 453 ceel energy expendi, elk calves robbins,ct; cohe/ 1979 JWMAA 43--2 564 567 ceel postur, activ, metab, cold gates,c; hudson,r 1979 QJEPA 62--4 333 340 ceel ener cost locom level, gra brockway,jm; ges/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 57-11 2153 2159 ala1 morph param affec loco, sno telfer, es; kelsa/ 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 56--2 215 223 rata energy expend, walk, tundr white, rg; yousef, 1978 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS---- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

BJNUA 18--1 47 54 dosh ener metab, walk lev, grad clapperton, j1 1964 PNUSA 20-- xxxi xxxii dosh ener exp, walk, lev, gradi clapperton, j1 1961

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CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS	5			AUTHORS	YEAR
JRMGA	276	437	443	rumi	phys	siol,	ener	expen,	pastu	osuji,po	1974

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS-			AUTHORS	YEAR
CBPAB	34A-4	841	846	many	ener	getic	cost,	locomotion	tucker,va	1970

CHAPTER 7, WORKSHEET 4.3a

Estimates of the energy cost of walking based on weight and gradient

Estimates of the energy cost of walking may be made based on data for domestic sheep, where the costs are calculated as a function of weight, distance, and height. Equations given in Moen (1973:356), modified with different symbols here, are:

ECWL = (0.59)(LWKG)(DSTK)

ECVA = (6.45)(LWKG)(HGTK)

where ECWL = energy cost of walking on the level, LWKG = live weight in kg, DSTK = distance in kilometers.

> ECVA = energy cost of vertical ascent and, HGTK = height ascended in kilometers.

Suppose an animal weighed 60 kg and walked a distance of 4 horizontal km at 4 km per hour on a 10% gradient as indicated in the drawing below.



The equations are:

ECWL = (0.59)(60)(4.0) = 141.60ECVA = (6.45)(60)(0.4) = 154.8,

and the total cost is 296.4 kcal.

How does this relate to MBLM? Multiply 296.4 by 24 to get the cost on a daily basis. The answer is 7113.6 kcal per day. Divide by base-line metabolism: $7113.6/(70)(60^{0.75}) = 4.71 = MBLM$. This is a rather high cost, and could not be sustained for long. If the rate of speed were cut in half (2 hours rather than 1 hour of walking time), then 148.2 kcal per hour 3556.8 and MBIM = 2.36. A further reduction in speed of travel at 1 km per hour would reduce the cost to 74.1 kcal per hour. Base-line metabolism is

Chapter 7 - Page 58a

62.9 kcal per hour, so the cost of walking 1 km per hour and ascending vertically a total of 10% of the horizontal distance plus the cost of base-line metabolism (or basal if you wish) is, in this case 74.1 + 62.9 = 137 kcal, 137/70 = 1.96 = MBLM.

Note how the example on the previous page allows for differences in the rate of speed. Variations in vertical ascent were not evaluated. New variables may be used and different combinations evaluated. The calculation of MBLM is always a good idea because of the large amount of evidence for upper limits to ecological metabolism as discussed in UNIT 6.1 of this CHAPTER.

LITERATURE CITED

Moen, A. N. 1973. Wildlife Ecology. W. H. Freeman and Co. San Francisco. 458 pp.

CHAPTER 7, WORKSHEET 4.3b

Estimates of energy expenditure of elk calves at different speeds on different upslopes

Measurements of the enegy expenditure of elk calves at different speeds on three diffferent slopes were made by Robbins et al. (1979). The energy expenditures per kg per hour on three different slopes were graphed in Figure 1, but no equations wer presented for these results. A single equation may be derived for locomotion upslope by first determining linear regression equations for each of the slopes (11.7, 4.2, 0°). The a and b values in these equations are then expressed with equations. The a values are best fit with exponential regression equations, and the b values with linear regression equations. The final, overall equations are:

 $ECWU = 3\ 28599\ e^{0.01660}\ (SLPD) + [0.97296 + 0.05722\ (SLPD)]$ SPKH

 $ECWD = 3.18453 e^{0.01741} (SLPD) + [0.96953 + 0.02381 (SLPD)] SPKH$

where ECWU = energy cost of walking upslope in kcal per kg per hour, SLPD = slope in degrees, SPKH = speed in km per hour and,

ECWD = energy cost of walking downslope in kcal per kg per hour.

Calculate several combinations of speed and slope and plot below. Different slopes may be convienently represented as a family of curves.



LITERATURE CITED

Robbins, C. T., Y. Cohen, and B. B. Davitt. 1979. Energy expenditure by elk calves. J. Wildl. Manage. 43(2):445-453.

CHAPTER 7, WORKSHEET 4.3c

The energy cost of walking for reindeer (rata)

Measurements of oxygen and consumption by reindeer waling on roads or on tundra have been made by White and Yousef (1978). Their results are expressed as equations for oxygen consumption Table 1, p. 217. Using the energy equivalent of oxygen consumed in relation to the respiratory quotient (see CHAPTER 7, UNIT 2.2, WORKSHEET 2.2a, p. 32a), a respiratory quotient of 0.85 may be used if a specific REQO is not available, calculate the energy cost for each of the several experimental conditions. Body weights are also given in Table 1, so MBLM may be calculated.

Complete the calculations and compare these costs of walking calculated in WORKSHEETS 4.3a and 4.3b. Keep in mind that these direct measurements are for all of the metabolic processes of reindeer while walking.

LITERATURE CITED

White, R. G. and M. K. Yousef. 1978. Energy expenditure in reindeer walking on roads and on tundra. Can. J. Zool. 56(2):215-223.

UNIT 4.4: THE ENERGY COST OF FORAGING

The costs of foraging are quite variable due to the differences in the styles of foraging and in the terrain inhabited by different wild ruminants. Pronghorn, for example, live in a rather sparsely vegetated habitat and cover large land areas as their home range. Forage density is much less than one finds on white-tailed deer range in the northeastern states where deciduous forests are interspersed with farms. Sheep and goats live in rugged areas with rather sparse forage densities too, and foraging for these species may be more costly than for pronghorn or deer.

Measurements of the energy costs of foraging have not been made for free-ranging ruminants. The use of oxygen consumption masks, as for the measurements on reindeer and elk discussed in the two previous units, prohibits foraging, of course. Tracheal cannulae have apparently not been tried on wild ruminants.

A single value, for the cost of foraging, based on data for grazing domestic sheep and expressed as a multiple of base-line metabolism, is given in Moen (1973:356). Such approximations are often necessary because of the lack of measured data on free-ranging wild ruminants due to the technical difficulties involved in the measurements. A WORKSHEET provides opportunities to estimate the energy cost of foraging.

LITERATURE CITED

Moen, A. N. 1973. Wildlife ecology. W. H. Freeman and Co., San Francisco. 458 pp.

REFERENCES, UNIT 4.4

THE ENERGY COST OF FORAGING

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARECOLA 57--1 192198 odvi energy conservatio, winter moen, an1976JWMAA 34--2 431439 odvi wint feed patterns, penned ozoga, jj; verme, 11970JWMAA 42--4 715738 odvi seas chan, heart rate, act moen, an1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 43--2 564 567 ceel postur, activ, metab, cold gates,c; hudson,r 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--2 391 392 rata energ metab, bar gro carib mcewan,eh 1970 CJZOA 48--5 905 913 rata seas chan, energ, nit inta mcewan,eh; whiteh 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

anam

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 23-21 359 363 ovca energ costs feedin, roc mt chappel,rw; hudso 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
CODEN	VO-NU	BEPA	ENPA	ANIM	KEY	WOR	DS			AUTHORS-		YEAR
AFBUD		38	39	doca	ener	ev (cost.	ingestin	feed	adam.i:	voung.ba/	1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARAJAEA 15--6 969 973dosh energ cost, feed act, graz graham,nc1964AJAEA 17--3 355 362dosh energy expend, resp, feedi young,ba1966PAANA 7---- 327 334dosh ener req, grazi, calorimet young,ga; corbet1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JRMGA 27--6 437 443 rumi physiol, ener expen, pastu osuji,po 1974

CHAPTER 7, WORKSHEET 4.4a

Estimating the energy cost of foraging

The energy cost of foraging may be estimated with a simple mathematical relationship given in Moen (1973:356). The equation given is $(0.54)(W_{kg})$ the rate per hour. Multiplying by 24 to convert to a 24-hour period and using 4-letter symbols, the equation is:

ECFD = (12.96)(LWKG)(FTFD)

where ECFD = energy cost of foraging per day, LWKG = live weight in kg, and FTFD = fraction of time foraging per day.

The result does not include maintenance costs; ECFD is for the cost of foraging alone and not the other concomitant productive processes.

Note that the fraction of time spent foraging per day is part of the calculation. These fractions were discussed in PART II, and need to be considered again here.

Convert the estimates to MBLMs again. Note that foraging costs less than walking at the rate of 1 km per hour. Foraging is intermittent standing and walking.

LITERATURE CITED

Moen, A. N. 1973. Wildlife Ecology. W. H. Freeman and Co. San Francisco. 458 pp.

UNIT 4.5: THE ENERGY COST OF RUNNING

Running is a highly variable cost due to the different velocities possible, but a small contributor to the daily energy expenditure of wild ruminants because only a small fraction of each day is spent running. The cost of running may be determined from estimates, or calculated with the heart rates or respiration rates and converted to estimates of metabolism, and expressed as multiples of base-line metabolism. Since muscle artifacts often interfere with the telemetered ECG signal, heart rates of running animals are difficult to obtain. It may be that multiples from 6 to 20 times base-line metabolism have to be chosen as estimates simply because measured values are unavailable.

REFERENCES, UNIT 4.5

THE ENERGY COST OF RUNNING

SERIALS

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS AUTHORS	YEAR
ECOLA	571	1 92	198	odvi	energy conservatio, winter moen,an	1976
JWMAA	424	715	738	odvi	seas chan, heart rate, act moen,an	1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR ATRLA 23-22 365 370 ceel energy cost of locomotion gates,c; hudson,r 1978 CBPAB 61A-1 43 48 ceel oxy util, horiz, vert loco cohen,y; robbins/ 1978 JWMAA 43--2 445 453 ceel energy expendi, elk calves robbins,ct; cohe/ 1979 JWMAA 43--2 564 567 ceel postur, activ, metab, cold gates,c: hudson,r 1979 QJEPA 62--4 333 340 ceel ener cost locom level, gra brockway,jm; ges/ 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 57-11 2153 2159 ala1 morph param affe loco, sno telfer,es; kelsa/ 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS---- YEAR CJZOA 56--2 215 223 rata energy expend, walk, tundr white, rg; yousef, 1978 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJPHA 219-4 1104 1107 mamm energy cost run, body size taylor, cr; schmi/ 1970 CBPAB 34A-4 841 846 many energetic cost, locomotion tucker, va 1970 SCIEA 178-- 1096 1097 runni up, down hills: size taylor,cr; caldw/ 1972

CHAPTER 7 - WORKSHEET 4.5a

Estimates of the energy costs of running

The cost of running is quite variable, depending on the speed. The cost of locomotion of elk calves at 12 km per hour on 0° slope (Robbins et al. 1979) has an MBLM of 13 to 14. The range of multiples--6 to 20--given on page 63 indicates wide possibilities for errors. Actual errors on a daily cost basis are small, however, since running can take up only a very small fraction of each day. Thus the suggested formula is:

 $ECRD = (RMBL)(70)(IFWK^{0.75})(FTRD)$

where ECRD = energy cost of running per day,

RMBL = running multiple of base-line metabolism,

IFWK = ingesta-free weight in kg, and

FTRD = fraction of time spent running each day.

A running time of 10 minutes per day is less than 1% of the daily time budget, so larger variations in MBLM are offset by small possibilities for error in FTRD.

Complete calculations of ECRD for different combinations of RMBL and FTRD. Use data in PART II on time budgets to establish realistic limits to FTRD and evaluate the possible errors of estimation. Once the costs of all activities are added up, errors in ECRD will be found to be small.

LITERATURE CITED

Robbins C. T., Y. Cohen, and B. B. Davitt. 1979. Energy expenditure by elk calves. J. Wildl. Manage. 43(2):445-453.

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TOPIC 5. VITAL SIGN: METABOLISM RELATIONSHIPS

Vital signs of free-ranging animals are difficult enough to measure, but measurement of the metabolism of wild, free-ranging ruminants is essentially impossible. Estimates of the ecological costs of living are absolutely essential for calculating energy relationships within populations and systems, however, so methods for making such estimates must be found.

The use of equations to convert data on vital signs to estimates of metabolism is often criticized due to a lack of accuracy. It may, however, be the only practical way to get estimates for long time periods. The patterns of metabolism estimates based on vital signs may give one considerable insight into variations from one season to another through the annual cycle. Conversion of heart rates to estimates of the energy cost of activity follows the sequence of calculations illustrated in the figure below, based on Moen (1978). Respiration rates are added to the sequence to illustrate how heart rates and respiration rates may be used either alone or together. Definitions of the symbols are given on the next page.



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AGDA = age in daysBHRM = bedded heart rate per minute BRRM = bedded respiration rate per minute CLWK = calculated live weight in kg ECBD = energy cost of bedding per day ECFD = energy cost of foraging per day ECRD = energy cost of running per day ECSD = energy cost of standing per day ECWD = energy cost of walking per day ELMD = ecological metabolism per day FHRM = foraging heart rate per minute FRRM = foraging respiration rate per minute FTBD = fraction of time bedded per day FTFD = fraction of time foraging per day FTRD = fraction of time running per day FTSD = fraction of time standing per day FTWD = fraction of time walking per day HRMC = heart rate to metabolism conversion factor IFMW = ingesta-free metabolic weight IFWK = ingesta-free weight in kg JDAY = Julian day PMRB = predicted metabolic rate bedded PMRF = predicted metabolic rate foraging PMRR = predicted metabolic rate running PMRS = predicted metabolic rate standing PMRW = predicted metabolic rate walking RHRM = running heart rate per minute RRRM = running respiration rate per minute SHRM = standing heart rate per minute SRRM = standing respiration rate per minute WHRM = walking heart rate per minute WRRM = walking respiration rate per minute

The energy costs of activities can be predicted with a sequence of equations that relate heart rate, metabolism, and behavior. Heart rates observed during specific activities (See CHAPTER 6, UNIT 2.1) are calculated first Then heart rate is converted to metabolism (See UNIT 5.2), and the results multiplied by the metabolic weight. This results in a 24-hour estimate of energy metabolism for a specific activity. Since an animal does not spend 24 hours a day in any one activity, the 24-hour cost is multiplied by the fraction of time spent in each activity per day (See CHAPTER 4). The daily costs of each activity are then added together to determine ecological metabolism per day (ELMD).

Two inputs--AGDA and JDAY--are necessary for the calculations. Both are used to calculate live weight (CLWK), which is converted to ingesta-free weight (IFWK) and ingesta-free metabolic weight (IFMW) = $IFWK^{0.75}$). The Julian day (JDAY) is then used to calculate the heart rates and respiration rates in each of the activities, followed by the heart rate-to-metabolism conversion factor, HRMC, discussed in the WORKSHEET that follows, and the respiration rate-to-metabolism conversions factor RRMC (See WORKSHEET that follows).

Metabolic body weight, heart beats per minute, and the heart rate-tometabolism conversion factor are then combined to estimate the metabolism of the animal for a 24-hour period of time in each of the five activities listed. These 24-hour estimates are then multiplied by the fractions of time in each of the five activities (determined from JDAY) to determine the energy cost of bedding per day (ECBD), energy cost of standing per day (ECSD), energy cost of walking per day (ECWD), energy cost of foraging per day (ECFD), and the energy cost of running per day (ECRD) throughout the year. The sum of these costs is the ecological metabolism per day (ELMD). Maintenance costs are included, however, since heart beats during activities also support the biological functions that provide for tissue maintenance.

LITERATURE CITED

Moen, A. N. 1978. Seasonal changes in heart rates, activity, metabolism, and forage intake of white-tailed deer. J. Wildl. Manage. 42(4):715-738.

REFERENCES, TOPIC 5

VITAL SIGN: METABOLISM RELATIONSHIPS

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR

edbo acpr nyny 427 many comprtv nutritn wild anims crawford,ma,ed 1968 aubo hutc loen 332 rumi energy metabo of ruminants blaxter,kl 1967 aubo wile nyny 184 rumi metabolism in the ruminant annison,ef; lewis 1959 edbo jdve zusw 259 doca energ met farm anims; symp schurch,a,ed; wen 1978

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UNIT 5.1: BODY TEMPERATURES

Body temperatures are indicators of the metabolic rate in relation to that expected under current conditions. Low body temperatures indicate depressed metabolism and hypothermia. High body temperatures indicate either elevated metabolism or a breakdown of heat loss mechanisms. Elevated metabolism may be due to infections, or to prolonged activity. Body temperatures are useful indicators of general body condition, but not of the absolute levels of metabolism. Temperatures are the effect rather than the cause.

Measurements of body temperatures are most often made in the rectum, representing a deep body temperature. Variations in temperatures do occur over different parts of the body, and the locations of measurements should be noted, especially when sites other than the rectum are involved.

REFERENCES, UNIT 5.1

BODY TEMPERATURES

SERIALS

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
APAVD	1 97 6	185	194	odvi	succinylcholine in deer	kitchen, h	1976
JWMAA	404	626	629	odvi	predic metab rat, heart ra	holter,jb; urban/	1976
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
JWMAA	191	154	155	odhe	normal temperat, colum btd	cowan,im; wood,a	1 9 55
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
				ceel			
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHOR S	YEAR
JWMAA	393	634	636	alal	physiol effec, m-99 etorph	roussel,ye; paten	1975
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
CJZOA	435	683	687	rata	body temp, barren ground c	mcewan,eh; wood,/	1965

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR 1975 JOMAA 56--3 697 698 anam normal body temp, mule de thorne, et JWMAA 35--4 747 751 anam telemetry syst, body tempe lonsdale,em; bra/ 1971 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWIDA 7---2 105 108 ovca comp phys vals, capt & wld franzmann,aw; 1971 JWMAA 35--3 488 494 ovca variation rectal temperatu franzmann, aw; heb 1971 JWMAA 36--3 924 ovca physiologc vals, env varia franzmann, aw 1972 932 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovđa CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BJNUA 21--3 769 785 dosh cont measure heart rate in webster, ajf 1967 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 22--1 3 24 caca energy metabolism, roe dee weiner, j 1977 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JPHYA 176-- 136 144 deep bod temp 12-month per bligh, j; ingram, / 1965 JWMAA 34--4 921 925 ungu radio-telem, temp, unrestr mcginnis,sm; fin/ 1970

UNIT 5.2: HEART RATE-TO-METABOLISM CONVERSIONS

Relationships between heart rates and metabolic rates have been determined under controlled conditions for several species. The general pattern of increased heart rate: increased metabolism is clear, but the application of a general equation to individuals is not good due to wide variability between individuals. Some, for example, have larger hearts and greater stroke volumes than others, so their heart rates are less per unit metabolism. Some have higher overall rates of metabolism, and some have different ratios of heart rate: metabolism due to differences in vascular resistances and the efficiencies of gas exchange.

The most complete set of simultaneous measurements of heart rate and metabolic rate has been made by Holter et al. (1976) on white-tailed deer. Seasonal effects were statistically significant. No statistically significant effects on metabolic rate of sex or nutrition level, or of sex x season, nutrition level x season, or sex x nutrition level interactions were found. They presented four linear regression equations--for summer, fall, winter, and spring--for calculating metabolic rate from heart rate.

Seasonal differences in the heart rate:metabolic rate relationships for an animal must involve gradual shifts through the annual cycle. The use of four separate equations in data analyses would result in a discontinuity at the beginning of each season. Accordingly, the four separate equations were combined into a single equation by expressing the a values and the b values as single equations for a and b rather than as seasonal constants. This is discussed in Moen (1978:723). The resulting equation is presented in WORKSHEET 5.2a.

Heart rate:energy expenditure relationships are discussed by Robbins et al. (1979) for elk calves, but an equation is not given for their data in Figure 5 (p. 451). They point out the large amount of variability in the relationship, but this variability may be due as much or even more to transient effects on heart rate of their animals while outside as to a fundamental physiological difference in the heart rate:metabolic rate relationships.

Careful interpretation of heart rates is necessary. Heart rate transients due to known stimuli may be pronounced (Moen et al. 1978 and Moen and Chevalier 1977), resulting in wide fluctuations in heart rates. The effects of such transients were eliminated in the measurements described in Moen (1978), resulting in distinct seasonal patterns to both heart rate and metabolism that would have been much more difficult to recognize if random samples of heart rates without behavioral observations had been made.

LITERATURE CITED

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- Moen, A. N. and S. Chevalier. 1977. Analyses of telemetered ECG signals from white-tailed deer. Pages 118-125 In Proc. of Biotelemetry Conf. Univ. of Wyoming, Laramie.
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- Robbins, C. T., Y. Cohen, and B. B. Davitt. 1979. Energy expenditure by elk calves. J. Wildl. Manage. 43(2):445-453.

REFERENCES, UNIT 5.2

HEART RATE-TO-METABOLISM CONVERSIONS

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
APAVD	1976	185	194	odvi	succinylcholine in deer	kitchen,h	1976
JWMAA JWMAA	404 424	626 715	629 738	odvi odvi	predic metab rat, heart ra seas chan, heart rate, act	holter,jb; urban/ moen,an	1976 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR alal CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 56--2 215 223 rata energy expend, walk, tundr white, rg; yousef, 1978 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

BJNUA 21--3 769 785 dosh cont meas heart rat indi e webster,ajf 1967

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WOF	RDS			- AUTHORS	YEAR
ATRLA	221	3	24	caca	ener	gy	metabol	ism,	roe de	e weiner,j	1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARAJCNA 22--- 696700 many heart-rate tel, energ, man bradfield,rb; hu/ 1969ATRLA 16--1 121metabo levels, homeotherms poczopko,p1971CJBIA 38-1113011309pulse rate, meta rate, man booyens,j' hervey 19601960JAPYA 14--6927936oxy inta, heart, work, man wyndham,ch: stry/ 19591959JAPYA 26--3297302energy exp, work, hear rat datta,sr; ramanat 1969

CHAPTER 7, WORKSHEET 5.2a

Heart rate to metabolism conversions for white-tailed deer

The heart rate to metabolism conversion equation in Moen (1978:723), modified to include four-letter symbols is given below.

 $MRMW = \{-0.92 \sin[(JDAY)(0.9863) + 74] - 0.005 + 2.3\} HRPM +$

 $\{31.9 \sin[(JDAY)(0.9863) + 74] + 0.087 + 0.55\}$

where MRMW = metabolic rate per unit metabolic weight = metabolic rate in $kcal/W_{kg}^{0.75}$, JDAY = Julian day, and HRPM = heart rate per minute.

The 0.9863 is the day-to-degree conversion factor (360/365 = 0.9863).

A nomogram or a table of MRMW values would be useful for later reference and calculations. Verify the equation and complete a nomogram on the grid on the next page where JDAY is on the x-axis, MRMW is on the y-axis, and HRPM a family of curves (30, 40...100) for example, or set up a table as indicated below.

HRPM =	30	40	50	60	70	80	9 0	100	110	120	>
	1										
	8										
JDAY =	15										
	22										
	29										
	•										
	•										
	•								•		
	365										

MRMW =

Ť Ť Ť Ť | | | | T T T ┍╴┾╼╊╼╊╼╊╼╊╼╊╼╊╼╊╼╊╼╊╼┠╍┠╍┠╍┠╾┠╍┠╾┠╺┠╼┠╾╊╼╊╼╊╸╊╸╊╸╊╾╊╾╊╾╊╾╊╾╊╾╊╾╊╾╊╾┠╾╊╾┠╾╋╾╄╾╄╾╄╾ | T T T ╎┯┯┯┯┯┯┯┯┯┯┯┯┯┯┯┯┯┯┯ Ť T Ť T T T ┱┲╌╖┰╼┯┯╼┯┯╼┯┯╼┯┯╼┯┯╼┯ T Ť T T **┰┰┰┰┰┰┰┰┰┰┰┰┰┰┰┰┰┰** İ Ť T T İ Τ Ť T T T T T T T T T T T T T T T T Ť T T T T T T Ť T **┰╌┰╌┰╌┰╌┲╌┰╌┰╌┰╌┰╌┰╌┰╌┰╌┰╌┰╌┰╌ ┰╶┰╶┰╴┰╶┰╶┰╌┰╸┰╸┰╸┰╸┰╸┰╸┰╸┰╸┰╸┰╸┰╸** T T T T Ť T T T T T Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ Ţ T T T Τ T T T T T T T T T Τ T T T T T T T T T T Ì T T T İ

LITERATURE CITED

Moen, A. N. 1978. Seasonal changes in heart rates, activity, metabolism, and forage intake of white-tailed deer. J. Wildl. Manage. 42(4):715-738.

CHAPTER 7, WORKSHEET 5.2b

Heart rate-to-metabolism conversions for elk calves

The relationships between energy expenditures and heart rates of five elk calves were plotted by Robbins et al. (1979:451; Fig. 5). Four of the five were quite similar. Using a grid overlay, estimate the x-y pairs for these four and derive a linear regression equation for these data. Remember that such an equation can be easily derived with hand calculations described in CHAPTER 2, UNIT 1.2, WORKSHEET 1.2b, p. 10b).

Compare the results above with those calculated in the previous WORKSHEET. Keep in mind that these are for a different species, different ages and for only a 2-month experimental period (summer).

Comparisons like those suggested above help one understand both biological and mathematical concepts, and understanding is fundamental to syntheses of ecological pictures.

LITERATURE CITED

Robbins, C. T., Y. Cohen, and B. B. Davitt. 1979. Energy expenditure by elk calves. J. Wildl. Manage. 43(2):445-453.

Chapter 7 - Page 74bb

UNIT 5.3: RESPIRATION RATE-TO-METABOLISM CONVERSIONS

Relationships between external respiration rates and metabolism are somewhat predictable since oxygen, supplied by external respiration, is necessary for metabolism. As activity levels increase, respiration rates increase, though not necessarily in direct proportion since volumes of air respired also change. Respiration and heart rates combined should be a better predictor of metabolism than either one alone.

Respiration rates have been measured by White and Yousef (1978) on reindeer and Robbins et al. (1979) on elk calves as oxygen consumption was being measured, permitting evaluations of the relationships between both respiration rate and volume and energy expenditures. results are presented graphically by Robbins et al (1979:450; Fig. 3 and 4), with equations. Variations in respiratory frequency during standing and the effects of panting are clearly illustrated in Figure 4. Respiratory frequency alone, including effects due to posture, locomotion, and panting could hardly be expected to be a good predictor of energy expenditure. Recognition of respiratory characteristics and frequencies will help when interpreting respiratory rate data.

LITERATURE CITED

Robbins, C. T., Y. Cohen, and B. B. Davitt. 1979. Energy expenditure by elk calves. J. Wildl. Manage. 43(2):445-453.

White, R. G. and M. K. Yousef. 1978. Energy expenditure in reindeer walking on roads and on tundra. Can. J. Zool. 56(2):215-223.

REFERENCES, TOPIC 5.3

RESPIRATION RATE-TO-METABOLISM CONVERSIONS

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS A	AUTHORS	YEAR
APAVD	1976-	185	194	odvi	succinylcholine in deer k	citchen,h	1976
JWMAA	404	626	629	odvi	predic metab rat, heart ra h	nolter,jb; urban/	1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ANREA 189-1 91 108 ceel carotid, orbital retia, pr carlton,c; mckean 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR alal CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR rata CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEARATRLA 22--1 324caca energy metabolism, roe dee weiner, j1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BJNUA 21--3 769 785 dosh cont meas heart rat indi e webster,ajf 1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 22--1 3 24 caca energy metabolism, roe dee weiner,j 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARAJCNA 22--- 696700 many heart-rate tel, energ, man bradfield,rb; hu/ 1969ATRLA 16--1 121metabo levels, homeotherms poczopko,p1971CJBIA 38-1113011309pulse rate, meta rate, man booyens,j; hervey 1960JAPYA 26--3297302energy exp, work, hear rat datta,sr: ramanat 1969

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TOPIC 6. ECOLOGICAL METABOLISM

Ecological metabolism is the term applied to the energy cost of living of free-ranging animals throughout the year, including all of the maintenance, activity, and productive functions as the animal goes through the annual cycle (Moen 1973).

Daily (24-hour) blocks of time are minimum for the expression of ecological metabolism because of diurnal rhythms in activity. Hourly rates of metabolism change during each 24-hour period because of activity periods. Changes in diurnal rhythms over the annual cycle are related to the light regime and to current weather conditions. The light regime changes in predictable ways through the annual cycle, and the pattern of daily ecological metabolism reflects these changes.

Ecological metabolism is very difficult to measure because the measurements must be made as the animals behave naturally over 24-hour periods throughout the annual cycle. Since direct measurements of metabolic expenditures or heat production of free-ranging animals are impossible, indirect measurements must be used. Gas exchange methods are difficult to use on wild ruminants, and they interfere with the animals' activities. Tracheal cannulae interfere less than face masks, but still are difficult to use on free-ranging animals. Calculations of metabolism made from vital signs, especially heart rates and respiration rates, do reflect metabolic patterns and provide insight into the overall role of metabolism in ecological considerations.

Absolute values of ecological metabolism per day (ELMD) may be estimated by vital sign to metabolism conversions, or by summations of maintenance, activity, and production costs as discussed in the previous units in the chapter. The ratio of ELMD to BLMD, called the multiple of base-line metabolism (MBLM) (See CHAPTER 7 - Pages 2-3) is a good way to express ecological metabolism on a relative basis. The ratio of ecological metabolism to base-line metabolism varies throughout the year because of seasonal rhythms in activity, growth, and reproduction. Transient changes in these functions contribute to a variable ratio, but long-term trends are approximated by a sine wave.

The use of the MBLM approach in calculating ELMD involves the use of live weights. Seasonal rhythms in weights (see CHAPTER 1, UNIT 1.4) are combined with seasonal rhythms in MBLM to estimate ELMD. This approach is applicable to different species of wild ruminants in northern areas of the United States and in Canada since a metabolic depression in the winter, summer metabolic highs, and an overall pattern similar to that of the white-tailed deer seems to exist in these ruminants.

Chapter 7 - Page 79

One important use for the calculation of ecological metabolism, a dynamic variable so dependent on the activity and production levels of each individual, is in determining nutrient requirements in relation to age. Also forage ingested and fat reserves, both used as sources of energy by the animal to meet the cost of ELMD, are both important as their rates of depletion determine the forage and fat resources left, both of which are of critical importance to survival.

This unit is a place for the listing of metabolism references that do not fall clearly in previous units, or for references that should be included for use again in deriving equations for ecological metabolism for different species. WORKSHEETS that follow provide opportunities for estimating ELMD from available data, and first approximations, at least, are to be derived for different species.

LITERATURE CITED

- Moen, A. N. 1973. Wildlife ecology. W. H. Freeman and Co., San Francisco. 458 pp.
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REFERENCES, TOPIC 6

ECOLOGICAL METABOLISM

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR

edbo acpr nyny 427 many comprtv nitritn, wild anim crawford,ma,ed; 1968 aubo hutc loen 332 rumi energ metabol of ruminants blaxter,kl 1967 aubo wile nyny 184 rumi metabolism in the ruminant annison,ef; lewis,1959 edbo jdve zusw 259 doca energ met farm anims; symp schurch,a,ed; wen 1970

UNIT 6.1: SEASONAL RHYTHMS IN ECOLOGICAL METABOLISM

Each individual must meet its own requirements for the basic life functions--respiration, circulation, daily activities, maintenance of body tissues, production of new tissues to replace those metabolized--that combine to establish the maintenance requirement of each individual. Productive members of a population must also synthesize new tissue, for both growth and reproduction. The timing of these productive functions results in marked changes in the rates of metabolism over the annual cycle. Highest metabolic rates occur in the summer when individuals are growing and females are lactating. In the winter, metabolism is depressed. Thus, ecological metabolism varies because maintenance, activity, and production costs vary due to seasonal patterns in growth, reproductive functions, activity patterns, weather and thermal exchange, and other biological functions. The timing of the productivity costs of ecological metabolism are dependent on the seasonal patterns of changes in range conditions.

The amount of direct information on seasonal metabolic rhythms of different species of free-ranging animals is very limited. There are clues in the literature, however. Deer are less able to withstand shock in the winter when they are in their period of metabolic depression. They also exhibit seasonal differences in their reactions to an immobilizing agent. Research at the Wildlife Ecology Laboratory has shown that there is seasonal response of white-tailed deer to succinylcholine immobilization that is apparently related to seasonal metabolic rhythms. Effective immobilization dosage rates varied from a low of 0.058 mg/kg in February to a high of 0.09 mg/kg in August (See Jacobsen et al. 1976). Stelfox and Robertson (1976) describe the seasonal reaction of bighorn sheep to succinylcholine chloride: "Lower dosage rates were required in late winter and spring compared to the fall." This suggests that bighorn sheep have a metabolic rhythm, too, and are energy conservers in the winter like white-tailed deer.

Seasonal rhythms in ecological metabolism of white-tailed deer have been described in detail in Moen (1978). It is interesting to compare the definitive pattern of high metabolism in the summer and low in the winter with supporting evidence published in earlier literature. Cyclic weight changes (Bandy et al. 1970), seasonal variations in hormone levels and reproductive functions (West and Nordon 1976), blood characteristics (Franzmann et al. 1976), thyroid activity (Hoffman and Robinson 1966), serum thyroxine levels (Seal et al. 1972), cementum annulus formation (Sauer 1973) and basal and fasting metabolic rates (Silver et al. 1969 and Holter et al. 1976) have all provided documented evidence for seasonal rhythms in metabolism. The WORKSHEETS that follow in this UNIT and UNIT 6.2 are very important for the verification of calculations of ecological metabolism.

LITERATURE CITED

- Bandy, P. J., I. McT. Cowan, and A. J. Wood. 1970. Comparative growth in four races of black-tailed deer (<u>Odocoileus hemionus</u>). Part I. Growth in body weight. Can. J. Zool. <u>48(6)</u>:1401-1410.
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- Silver, H., N. F. Colovos, J. B. Holter, and H. H. Hayes. 1969. Fasting metabolism of white-tailed deer. J. Wildl. Manage. 33(3):490-498.
- Stelfox, J. G. and J. R. Robertson. 1976. Immobilizing bighorn sheep with succinylcholine chloride and phencyclidine hydrochloride. J. Wildl. Manage. 40(1):174-176.
- West, N. O. and H. C. Nordan. 1976. Hormonal regulation of reproduction and the antler cycle in the male Columbian black-tailed deer (Odocoileus hemionus columbianus). Part I. Seasonal changes in the histology of the reproductive organs, serum testosterone, sperm production, and the antler cycle. Can. J. Zool. 54(10):1617-1636.

REFERENCES, UNIT 6.1

SEASONAL RHYTHMS IN ECOLOGICAL METABOLISM

SERIALS

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
CJPPA	566	945	949	odvi	thyroxine lev, male, femal	bubenik,ga; buben	1978
ECOLA	571	192	198	odvi	energy conservatio, winter	moen,an	1976
JOMAA JOMAA	411 472	23 266	29 280	odvi odvi	respon bucks, artifi light endocrine glands, sea chan	french,ce; mcewe hoffman,ra robin/	1960 1966
JWMAA JWMAA JWMAA JWMAA JWMAA JWMAA	342 344 364 403 424 424	431 863 1041 447 715 791	439 869 1052 453 738 798	odvi odvi odvi odvi odvi odvi	wint feed patterns, penned dig, met ener, wintr, does nutrit effec. thyroid acti seas var succinylcho immob seas chan, heart rate, act thymus, nut status indicat	ozoga,jj; verme,l ullrey,de; youat/ seal,us; verme/ jacobsen,nk; arm/ moen,an ozoga,jj; verme,l	1970 1970 1972 1976 1978 1978
CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
PNDAA	301	35	35	odhe	pituitary photoper, antlr	nicolls,ke	1976
PSEBA	93 - -1	161	162	odhe	cyclic var, thymus, mule d	browman,lg; sears	1956
778118	115-3	214	226	adha	nituitor and photoporiod	steelle he	1071
22ANA	11)-5	314	320	oane	picultal gind photoperiod	nicolls, ke	19/1
CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS	AUTHOR S	YEAR
CODEN ACATA	VO-NU 634	BEPA 580	528 ENPA 590	ANIM ceel	KEY WORDS caca, investigatn, thyroid	AUTHOR S pantic, v; stosic,	YEAR 1966
CODEN ACATA CBPAB	VO-NU 634 60a-3	BEPA 580 251	ENPA 590 256	ANIM ceel ceel	KEY WORDS caca, investigatn, thyroid doca, dosh enrg, nitro met	AUTHOR S pantic, v; stosic, simpson, am; webs/	1971 YEAR 1966 1978
CODEN ACATA CBPAB CODEN	VO-NU 634 60a-3 VO-NU	BEPA 580 251 BEPA	ENPA 590 256 ENPA	ANIM ceel ceel ANIM	KEY WORDS caca, investigatn, thyroid doca, dosh enrg, nitro met KEY WORDS	AUTHOR S pantic,v; stosic, simpson,am; webs/ AUTHOR S	1971 YEAR 1966 1978 YEAR
CODEN ACATA CBPAB CODEN NCANA	VO-NU 634 60a-3 VO-NU 101-1	BEPA 580 251 BEPA 227	ENPA 590 256 ENPA 262	ANIM ceel ceel ANIM alal	KEY WORDS caca, investigatn, thyroid doca, dosh enrg, nitro met KEY WORDS energ req, rumen fermentat	AUTHOR S pantic,v; stosic, simpson,am; webs/ AUTHOR S gasaway,wc; coady	1971 YEAR 1966 1978 YEAR 1974
CODEN ACATA CBPAB CODEN NCANA CODEN	VO-NU 634 60a-3 VO-NU 101-1 VO-NU	BEPA 580 251 BEPA 227 BEPA	ENPA 590 256 ENPA 262 ENPA	ANIM ceel ceel ANIM alal ANIM	KEY WORDS caca, investigatn, thyroid doca, dosh enrg, nitro met KEY WORDS energ req, rumen fermentat KEY WORDS	AUTHOR S pantic,v; stosic, simpson,am; webs/ AUTHOR S gasaway,wc; coady AUTHOR S	1971 YEAR 1966 1978 YEAR 1974 YEAR
CODEN ACATA CBPAB CODEN NCANA CODEN APSCA	VO-NU 634 60a-3 VO-NU 101-1 VO-NU 105-3	 BEPA 580 251 BEPA 227 BEPA 268 	ENPA 590 256 ENPA 262 ENPA 273	ANIM ceel ceel ANIM alal ANIM rata	KEY WORDS caca, investigatn, thyroid doca, dosh enrg, nitro met KEY WORDS energ req, rumen fermentat KEY WORDS spitzbergen, winter dorman	AUTHOR S pantic,v; stosic, simpson,am; webs/ AUTHOR S gasaway,wc; coady AUTHOR S ringberg,t	1971 YEAR 1966 1978 YEAR 1974 YEAR 1979
CODEN ACATA CBPAB CODEN NCANA CODEN APSCA BJNUA	VO-NU 634 60a-3 VO-NU 101-1 VO-NU 105-3 292	 BEPA 580 251 BEPA 227 BEPA 268 245 	ENPA 590 256 ENPA 262 ENPA 273 259	ANIM ceel ceel ANIM alal ANIM rata rata	KEY WORDS caca, investigatn, thyroid doca, dosh enrg, nitro met KEY WORDS energ req, rumen fermentat KEY WORDS spitzbergen, winter dorman seasonal, glucose metaboli	AUTHOR S pantic,v; stosic, simpson,am; webs/ AUTHOR S gasaway,wc; coady AUTHOR S ringberg,t luick.jr; person/	1971 YEAR 1966 1978 YEAR 1974 YEAR 1979 1973

rata continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--2 391 rata energ metab, bar gro carib mcewan, eh 1970 392 CJZOA 48--5 905 rata seas energy, nitrog intake mcewan, 1h whiteh 1970 913 CJZOA 56--2 215 223 rata energy expend, walk, tundr white, rg; yousef, 1978 JANSA 33--1 260 260. rata thyroxine secre rate, calv luick, jr; white,/ 1971 PASCC 19--- 71 1968 72 rata thyroxine secretion rate yousef, mk CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 37--4 563 573 anam energy metab of yg prongho wesley,de; knox,/ 1973 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CNJNA 59--3 611 617 bibi doca, seas enrg exp, thrmr christopherson, r/ 1979 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 56-11 2388 2393 ovca winter bioenergetics, rcky chappel,rw; hudso 1978 JWMAA 40--1 174 176 ovca immobiliz, succinyl, pheny stelfox, jg rober 1976 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 22--1 3 caca energy metabolism, roe dee weiner, j 24 1977 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR NCANA 101-1 227 262 alal energ req, rumen fermentat gasaway,wc; coady 1974 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR PZOOA 15--2 201 205 many lipid metab, free fatty ac felinski,l; kacz/ 1971

CHAPTER 7, WORKSHEET 6.1a

Estimating ecological metabolism by summation

The estimation of ecological metabolism by summation involves the calculation of the costs of each of the biological processes occurring on a given day and adding up these costs to get a total for the day. the major cost items have been discussed in the units in this chapter. There are too many variables involved that it is impossible to summarize the costs for each of these items for a species. Weights vary between sex and age groups and throughout the year, reproductive rates vary, activity patterns vary, etc. The schematic below will be useful as a check list when identifying cost items for summations.

Maintenance

Activity

Bedding Standing Walking level uphill downhill Foraging Running

Production

Growth

body growth hair growth

Reproduction gestation lactation

Establish a biochronology for the species of your choice, select the topography and behavioral regimes, estalish a reproductive rate, and identify every other maintenance, activity, and productive function that is part of that animal's life, and then complete the energy cost calculations discussed this CHAPTER 7. I suggest that you start with AGDA = 1, on the average JDAY for births. Follow that animal through its life, accounting for cost items characteristic of its life-style and events at 7 or 14 day intervals throughout the year. Such a calculation will call to your attention a great deal of natural history, behavior, and physiology, and will serve to acquaint you very well with the species. The suggestion given on the previous page is a major assignment. Programmed computing will be very helpful. Programmed desk-top calculators are efficient enough to help with the calculations while yet being simple enough to operate easily and with enough manual steps to keep you oriented.

One last reminder. As the energy costs are added up for each day selected, be sure to convert the absolute energy costs to MBLM. This will allow you to compare between sex, age, season, and species. The results should be somewhat in line with the material discussed in UNIT 6.2.

CHAPTER 7, WORKSHEET 6.1b

Seasonal rhythms in ELMD; the MBLM approach

The calculation of energy costs at selected intervals of days to be summed through the year is a major assignment. The calculation of seasonal rhythms in ELMD using the MBLM approach described in Moen (1978) can be accomplished with a single, relatively short equation. Basically, energy expenditures of white-tailed deer are sinusoidal throughout the year, reaching a low in the water and a high in the summer or early fall, depending on the reproductive rate. Lactation for two fawns shifts the maximum cost to mid-summer, for one fawn to late summer, and for no fawns to early fall.

The equation is:

 $MBLM = 2.56894 e^{0.19602} (NUFA) \left(\left\{ [1.0285 \sin [(JDAY + 170.0536 e^{0.15488} (NUFA)] 0.9863 + 0.0281] \right\} \\ [0.19 + 0.05 (NUFA)] \right\} + [0.81 - 0.05 (NUFA)] \right)$

where MBLM = multiple of base-line metabolism, NUFA = number of fawns, and JDAY = Julian day.

ELMD is determined by multiplying MBLM by (70)(MEWK). MEWK is determined from the weight equations described in CHAPTER 1. Calculate MBLM and multiply by (70)(MEWK) through the annual cycle and plot the results on the grid on the back of this page.

Compare the MBLMs with those discussed on previous WORKSHEETS in this chapter, and compare with those discussed in UNIT 6.2.

LITERATURE CITED

Moen, A. N. 1978. Seasonal changes in heart rates, activity, metabolism, and forage intake of white-tailed deer. J. Wildl. Manage. 42(4):715-738.

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JDAY =

MBLM
UNIT 6.2: INTERSPECIES COMPARISONS

The use of base-line metabolism and multiples of base-line metabolism to express ecological metabolism is very conveniently applied to other species with different weights. Calculations have been made for caribou (Moen 1980) indicating similarities in the pattern of MBLM over the annual cycle. Detailed calculations by Gasaway and Coady (1974), summarized in Moen (NAS In press) indicate that moose have a lower metabolism in the winter (about 1.7 times the basal metabolic rate; BMR) and 3.0 to 4.0 times BMR in the summer. Both the metabolic patterns and the multiples of BMR (where BMR = 70 W^{0.75} and is equivalent to base-line metabolism) of moose given by Gasaway and Coady (1974) are very similar to those determined for white-tailed deer by Moen (1978).

Interspecies comparisons using the multiple of base-line metabolism approach have been made for domestic animals; the following summary is from Moen (NAS In press). How does the use of MBLM apply to domestic cattle and sheep? First, the pattern of daily energy requirements of a 454 kg beef cow during her 12-month reproductive cycle is shown by Ensminger (1976:1083). Converting the graphed results to MBLM by dividing values on the (megacalories) Y-axis by base-line metabolism, $[70(454)^{3/4})]$, results in values of MBLM of 1.8, 2.3, 2.5, and 3.5 for maintenance, grazing, the addition of fetal development, and peak lactation, respectively. Both the general pattern and the values of MBLM are very similar to those discussed earlier for white-tailed deer and moose. The timing is different, as cows may be bred to calve in late winter and early spring, earlier than the time of parturition for wild ruminants.

The figure in Ensminger depicting metabolizable energy of a cow over a 12-month reproductive cycle is illustrative, and not designed for detailed expression of metabolizable energy and forage requirements of cows of different ages, weights, and breeds. The nutrient requirements of beef cattle and sheep have been summarized by subcommittees of the National Research Council (Nutrient Requirements of Beef Cattle, fifth revised edition, 1976 and Nutrient Requirements of Sheep, fifth revised edition, 1975). These publications include tables of metabolizable energy values in the forage consumed by animals of different weights, `growth rates, and different stages of reproduction. The tables may be converted to MBLM by dividing the metabolizable energy in the daily feed allotment by base-line metabolism.

The values of MBLM for lactating beef cattle range from 3.1 to 3.7 (midpoint 3.4) for cows from 350 to 650 kg rated as "superior" producers, and from 2.5 to 2.8 (midpoint 2.65) for those rated "average." The calculated MBLM for cows in the middle third of gestation is 1.9, and in the last third, 2.35 to 2.15 (midpoint 2.25) for cows weighing 350 to 650 kg.

Metabolizable energy values for sheep, expressed as MBLM calculated from Table 1, page 42 in <u>Nutrient Requirements of Sheep</u> (NRC 1975), include a maintenance cost after lactation has ceased of MBLM = 1.45, the lowest of the year, to an MBLM during lactation of 3.5 for sheep suckling singles and 4.0 for sheep suckling twins. An interesting and significant relationship emerges for these calculations of MBLM for cattle and sheep in relation to MBLM calculations for white-tailed deer. Maximum values of MBLM are 3.7, 4.0, and 3.9 for cattle, sheep, and deer, respectively. MBLM values for an average producing cow and sheep and deer nursing singles are 2.35, 3.5, and 3.2 respectively. Minimum MBLMs are 1.45 and 1.6 for sheep and deer, respectively, and 1.9 and 1.7 for cows and moose, respectively. Differences in ecological metabolism of free-ranging ruminants appear to be slight when compared on a relative basis using MBLM. On an absolute basis, differences between species in the amount of metabolism per day are caused by differences in their weights, especially of cattle and moose compared to sheep and deer.

Similarities in MBLM for the different four species suggest that MBLM can be estimated for free-ranging ruminants as well. A maximum of 4.0 and minimum of 1.5 likely include the expected values of MBLM of free-ranging ruminants over the annual cycle. References that contain weight and energy metabolism data on domestic cattle and sheep may be listed on the next page. WORKSHEETS that illustrate interspecies comparisions and evolutions within the 1.5 to 4.0 MBLM should also be added at the end of this UNIT.

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REFERENCES UNIT 6.2

INTERSPECIES COMPARISONS

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AFBUD 38 39 doca energy cost, ingestin feed adam,i; young,ba/ 1979 BJNUA 20--1 103 111 doca fasting metabolism, cattle blaxter,kl; wainm 1966

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJAEA 15--6 969 973 dosh energ cost. feed act, graz graham,nm 1964 AJAEA 17--3 355 dosh energy expend, resp, feedi young, ba 362 1966 AJAEA 19--5 821 dosh metab rate, bred wol prod graham, nm 824 1968 BJNUA 18--1 47 54 dosh ener metab, walk lev, grad clapperton, j1 1964 PAANA 7---- 327 dosh ener req, grazi, calorimet young,ga; corbet 1968 334 PAANA 7---- 335 341 dosh co2, index ener expen, 1mb white, rg; leng, ra 1968 PNUSA 20-- xxxi xxxii dosh ener exp, walk levl, grdi clapperton,j1 1961

CODEN	VO-NU	BEPA	ENPA ANIM	KEY WORDS AUTHORS	YEAR
CBPAB	34A-4	841	846	energetic cost, locomotion tucker,va	197 0
JRMGA	274	437	443	physiol, ener expen, pastu osuji,po	1974
JTBIA	492	3 45	362	model, est metab, rest, ac wunder,ba	1975
SCIEA	178	10 9 6	1097	runni up, down hills: size taylor,cr; caldw/	1972
WLMOA	14	1	78	pharma-physiol prin, restr hartoorn,am	1965

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CHAPTER 7, WORKSHEET 6.2a

Interspecies comparisons of MBLM and ELMD

Ecological metabolism per day, being a function of weight, varies widely betwen species, ELMD of white-tailed deer is much less than that of moose, and of domestic sheep, much less than that of cattle.

Similarities in MBLMs of different species are striking, however. MBLM is weight independent, calculated by dividing ELMD by base-line metabolism. Interspecies comparisons can be easily made when daily quantities of energy metabolism are given. Sometimes the metabolizable energy in the feed is given: this is the case for domestic cattle and sheep in the NRC publications. Consider the metabolizable energy in the daily ration to be the daily metabolizable energy and divide by BLMD.

The point to be made in this WORKSHEET is that MBLM is the expression to use for interspecies comparisons. It has been stressed throughout this CHAPTER, and will prove to be very useful in future calculations.

Mentally review MBLM, think of patterns through time, and sketch your thoughts in the space below.

CLOSING COMMENTS - CHAPTER SEVEN

CHAPTER 7 is a very important chapter in this book of seven parts. Calculations of the energy cost of living are as important to the resource manager as calculations of the dollar costs of goods are to the economist. Energy is the currency of the ecosystem, and an understanding of energy transactions helps one realize the potentials for and the limits to ecological processes in the natural world.

> Aaron N. Moen March 11, 1981

GLOSSARY OF SYMBOLS USED - CHAPTER SEVEN

AGDA = Age in days BAMD = Basal metabolism per day BIWK = Birth weight in kilograms BHRM = Bedded heart rate per minute BLM- = Base-line metabolism BLMD = Base-line metabolism per day BMR- = Basal metabolic rate BRRM = Bedded respiration rate per minute CLWK = Calculated live weight in kilograms DHGD = Duration of hair growth per day DIGE = Days into gestation DILA = Days into lacation DSTK = Distance in kilometers ECBD = Energy cost of bedding per day ECFD = Energy cost of foraging per day ECGD = Energy cost of gestation per day ECHD = Energy cost of hair growth per day ECRD = Energy cost of running per day ECSD = Energy cost of standing per day ECVA = Energy cost of vertical ascent ECWD = Energy cost of walking per day ECWL = Energy cost of walking upslope ELMD = Ecological metabolism per day FACO = Fat content FAMD = Fasting metabolism per day FATP = Fat percentFHRM = Foraging heart rate per minute FRRM = Foraging respiration rate per minute FTBD = Fraction of time spent bedded per day FTFD = Fraction of time spent foraging per day FTRD = Fraction of time spent running per day FTSD = Fraction of time spent standing per day FTWD = Fraction of time spent walking per day GENM = Gross energy in the milk HAWG = Hair weight in grams HGTK = Height ascended in kilometers HIGR DIGE = Higher number of days into gestation HRMC = Heart rate to metabolism conversion factor HRPM = Heart rate per minute IFMW = Ingesta-free metabolic weight IFWK = Ingesta-free weight in kilograms

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JDAY = Julian day KCAL = Kilocalories KCLO = Kilocalories per liter of 0_2 KJOU = Kilojoules LOWR DIGE = Lower number of days into gestation LWKG = Live weight in kilograms MBLM = Multiple of base-line metabolism MEWK = Metabolic weight in kilograms MPGD = Milk production in grams per day MRMW = Metabolic rate per metabolic weight NECM = Net energy coefficient for milk NUFA = Number of fawns PEGE = Physiological efficiency of gestation PEHG = Physiological efficiency of hair growth PMRB = Predicted metabolic rate bedded PMRF = Predicted metabolic rate foraging PMRR = Predicted metabolic rate running PMRS = Predicted metabolic rate standing PMRW = Predicted metabolic rate walking PNDM = Percent nutrients derived from milk PRCO = Protein content PRPC = Primary phase correction REHP = Resting heat production REQO = Respiratory quotient RHRM = Running heart rate per minute RMBL = Running multiple of base-line metabolism RRRM = Running respiration rate per minute SHRM = Standing heart rate per minute SLPD = Slope in degrees SMBL = Standing multiple of base-line SPKH = Speed in kilometers per hour SRRM = Standing respiration rate per minute TCCK = Total caloric content in kilocalories WHRM = Walking heart rate per minute WRRM = Walking respiration rate per minute

GLOSSARY OF CODE NAMES - CHAPTER SEVEN

ACATA Acta Anatomica AFBUD University of Alberta Agriculture and Forestry Bulletin AJAEA Australian Journal of Agricultural Research AJCNA American Journal of Clinical Nutrition AJPHA American Journal of Physiology AMNAA American Midland Naturalist AMNTA American Naturalist AMZOA American Zoologist ANREA Anatomical Record ANYAA Annals of the New York Academy of Sciences APAVD American Association Zoo Veterinarian Annual Proceedings APSCA Acta Physiologica Scandinavica ATRLA Acta Theriologica BISNA Bioscience BJNUA British Journal of Nutrition BPURD Biological Papers of the University of Alaska Special Report CAFGA California Fish and Game CBPAB Comparative Biochemistry and Physiology A Comparative Physiology CIWPA Carnegie Institution of Washington Publication CJBIA Canadian Journal of Biochemistry CJZOA Canadian Journal of Zoology CNJNA Canadian Journal of Animal Science COVEA Cornell Veterinarian ECMOA Ecological Monographs ECOLA Ecology FEPRA Federation Proceedings IJBMA International Journal of Biometeorology JANSA Journal of Animal Science JAPYA Journal of Applied Physiology JASIA Journal of Agricultural Science JCCPA Journal of Cellular and Comparative Physiology JDSCA Journal of Dairy Science JOMAA Journal of Mammalogy JPHYA Journal of Physiology JRMGA Journal of Range Management JRPFA Journal of Reproduction and Fertility JTBIA Journal of Theoretical Biology JWIDA Journal of Wildlife Diseases JWMAA Journal of Wildlife Management NATUA Nature NAWTA North American Wildlife and Natural Resources Conference, Transactions of the, NCANA Naturaliste Canadien, Le

NUMEB Nutrition and Metabolism OJSCA Ohio Journal of Science PAANA Proceedings of the Australian Society of Animal Production PAARA Pennsylvania State University College of Agriculture Agricultural Experiment Station Progress Report PASCC Proceedings of the Alaskan Scientific Conference PECTD Polish Ecological Studies PHREA Physiological Reviews PHZOA Physiological Zoology PNDAA Proceedings of the North Dakota Academy of Science PNSUA Proceedings of the Nutrition Society PNUSA Proceedings of the Nutrition Society PSEBA Proceedings of the Society for Experimental Biology and Medicine PZOOA Przeglad Zoologiczny QJEPA Quarterly Journal of Experimental Physiology and Cognate Medical Sciences QSFRA Quebec Service de la Faune Rapport **RSPYA Respiration Physiology** SCIEA Science SSEBA Symposia of the Society for Experimental Biology SZSLA Symposia of the Zoological Society of London VJSCA Virginia Journal of Science WLMOA Wildlife Monographs ZOBIA Zhurnal Obshchei Biologii ZTTFA Zeitschrift fuer Tierphysiologie Tierer naehrung und Futtermittelkunde ZZAHA Zeitschrift fuer Zellforschung und Mikroskopisch Anatomie

LIST OF PUBLISHERS - CHAPTER SEVEN

acpr	Academic Press	New York	nyny
agrc	Agricultural Res. Council	London, England	loen
ccth	Charles C. Thomas	Springfield, IL	spil
coms	Comstock	Ithaca, NY	itny
fase	Federation of American Society for Experimental Biology	Bethesada, MD	bemd
hutc	Hutchinson	London, England	loen
jdve	Joris, Druck, and Verlag	Zurick, Switzerland	zusw
mopc	Morrison Publishing Co.	Ithaca, NY	itny
prha	Prentice-Hall, Inc.	Princeton, NJ	prnj
repu	Reinhold Publishing	New York	nyny
saco	Saunders Publishing Co.	Philadelphia, PA	phpa
whfr	W. H. Freeman Co.	San Francisco, CA	sfca
wile	Wiley	New York	nyny

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5	005	036	064	095	125	156	186	217	248	278	309	339	5
6	006	037	065	096	126	157	187	218	249	279	310	340	6
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15	015	046	074	105	135	166	196	227	258	288	319	349	15
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THE BIOLOGY AND MANAGEMENT OF WILD RUMINANTS

CHAPTER EIGHT

PROTEIN METABOLISM OF WILD RUMINANTS

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Published by

CornerBrook Press Box 106 Lansing, N.Y. 14882

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Library of Congress Catalog Card Number 80-70984

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CHAPTER 8. PROTEIN METABOLISM

Protein metabolism involves the breakdown of the protein in food ingested into amino acids to be synthesized into new proteins that become part of different animal tissues. Muscle tissue is the main protein depot in the body as it is primarily protein and water.

The pathways and terms used to describe protein metabolism from ingestion to net use are listed below. The word nitrogen is used rather than protein because nitrogenous compounds are used by ruminants to synthesize protein-containing tissue.

Total		fecal		apparent
nitrogen	-	nitrogen	=	digestible
intake				nitrogen

apparent		metabolic		true
digestible	-	fecal	=	digestible
nitrogen		nitrogen		nitrogen

true		urinary		net nitrogen	for
digestible	-	nitrogen	=	maintenance	and
nitrogen				production	ı

net nitrogen for maintenance and -> production



The fundamental unit or building block of a protein molecule is the amino acid. Amino acids contain amino groups (-NH₂) and carboxyl groups (-COOH) which link together to form peptides. Many amino acids linked together form a polypeptide, and very long polypeptides are proteins.

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The breakdown of or hydrolosis of natural proteins has yielded 26 different amino acids (McCauley 1971:190). Many of these are present in animal tissue, and can be produced by the process of transamination, or the interconversion from one to another. Some cannot be produced from other amino acids, so they must be present in the diet. These are called essential amino acids.

Amino acids not synthesized into protein are deaminated in the kidneys and liver and ammonia is produced (Pantelouris 1967). Ruminants convert ammonia to urea, which may then be used by rumen microflora in their own protein metabolism.

The ruminant animal has an advantage over the monogastric animal when meeting its protein needs because the intestinal microflora synthesize proteins which are, in turn, available to the ruminant host. Thus the ruminant does not need to have all of the essential amino acids supplied directly in the diet because the microflora synthesize these for the host. The total protein in the diet must be adequate in order to provide the necessary nourishment for the microflora (Crampton and Harris 1969:171). Ruminants also have a metabolic adaptation for the conservation of protein resources. The recycling of urea is a nitrogen conservation adaptation when protein intake goes down. This is characteristic of winter range conditions when many species of wild ruminants are on browse or highly lignified diets.

This chapter deals with the applied aspects rather than the biochemical aspects of protein metabolism. The TOPICS that follow include material on the factors affecting protein metabolism (TOPIC 1), measurements of protein metabolism (TOPIC 2), and estimates of protein costs for production (TOPIC 3). The equations presented will be used when evaluating range and diet characterics in PART IV.

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PROTEIN METABOLISM

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aubo	c1pr	oxen	272		meta	al	зЪ	of	ĒĮ	pro	t,	ma	amm	a1	ь	ody	bach,	sj				1952
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edbo	rupr	nbnj	119		ser	ol	510	ogy	7,	bi	ocl	hen	n, "	pr	ote	ein	cole,	wh,e	ed			1958
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TOPIC 1. FACTORS AFFECTING PROTEIN METABOLISM

Protein metabolism of wild ruminants is affected by both biological and environmental factors. Like other mammals ruminants convert the ammonia released from the amino acids that are not used in protein synthesis into ammonia and then urea. Urea is a soluble compound that is excreted in the urine. Ruminants, however, have the ability to recycle urea, making it available to rumen microflora for synthesis into new protein tissue.

The amount of urea recycling is dependent on environmental conditions. More recycling occurs when the crude protein intake is less, and for wild ruminants, crude protein levels are a function of range condition.

Protein metabolism is also affected by the overall body condition of the animal, which is also affected by range conditions. Nutritional stress after fat reserves are depleted results in the breakdown of protein tissue, which means the animal is in a very critical situation. The interdependence of animal and range is illustrated again by these examples. Animal-range relationships of free-ranging ruminants are very dynamic, with feed-back mechanisms that eliminate direct proportions and simple equations when evaluating them mathematically.

UNIT 1.1: BIOLOGICAL FACTORS AFFECTING PROTEIN METABOLISM

Two factors affecting protein metabolism, the true biological value or metabolizable protein and urea recycling, will be discussed in this UNIT.

The true biological value of protein is a representation of the amount of protein that is actually used for metabolism, or true metabolizable protein. This is partly a function of the way the protein or nitrogenous compounds are packaged in the forage, and partly a function of the nitrogen balance of the animal.

The "packaging" of protein or nitrogenous compounds in the forage affects the digestibility of the forage. Those forages that contain nitrogen compounds that are easily broken down into amino acids are high digestible forages, and those that have cell structures which bind the nitrogen compounds or very complex proteins are low digestible forages.

The nitrogen balance of an animal affects the amount of true metabolizable protein by affecting the fate of nitrogen compounds in the forage. An animal on a high protein and low energy diet will use some of the nitrogen compounds as a source of energy. The caloric value per gram of proteins (5.7) is less than that of fats (9.5). There, is an additional inefficiency because animals do not completely oxidize proteins, but excrete portions of protein molecules in the form of ammonia and urea (Florey 1966:270). Thus the gross energy in a gram of protein oxidized in a bomb calorimeter is 5.7 kcal, and in a gram of protein metabolized in the animal, 4.8 kcal.

The nitrogen balance also affects the metabolic disposition of protein compounds. Urea recycling occurs when urea in the blood enters the gastrointestinal tract where it is broken down into ammonia and carbon dixoide. Some of the ammonia is then synthesized into protein by intestinal microflora, and the remainder is excreted in the feces.

Urea recycling represents a means of conserving nitrogen when ruminants are consuming poor quality forage (Robbins et al. 1974). When urea recycling is maximum due to very low crude protein in the forage, the only additional source of nitrogen is tissue nitrogen. Then, body condition may deteriorate rapidly. Short-term tissue nitrogen mobilization meets immediate and specific nitrogen requirements, but cannot continue for long periods of time, of course.

There are other factors affecting protein metabolism too, including requirements for maintenance and production; these are discussed in TOP-ICS 2 and 3. There are not many references available on protein or nitrogen metabolism of wild ruminants, so protein costs for various biological functions must be estimated from data on domestic ruminants.

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CHAPTER 8, WORKSHEET 1.1a

Urea recycling and the biological value of protein metabolized by white-tailed deer (odvi)

The amount of urea recycled by white-tailed deer is a function of the crude protein in the diet. The equation given by Robbins et al. (1974) is:

 $Log_e Y = 5.3197 - 0.5007 Log_e X$

where X = dietary crude protein (%) and Y = urea recycled (% entry rate).

Rewriting this equation and changing X and Y to DCPP and UREP, respectively, the new equation is:

$$UREP = e^{5.3197} - 0.5007 \ln DCPP$$

where UREP = urea recycled in percent and DCPP = dietary crude protein.

Complete the calculations and plot the values below. At what % dietary crude protein is 100% recycling reached?



The biological value of the protein increased as the crude protein decreased. The equation in Robbins et al. (1974) is:

 $Log_e Y = 4.9825 - 0.3096 Log_e X$

where X = DCPP as defined before and Y = biological value (%) = BVAP

Rewriting the equation:

 $BVAP = e^{4.9825} - 0.3096 \ln DCPP$

The relationship may be plotted below.



The decreasing biological value of the protein with increasing dietary protein may result from less efficient use of dietary protein due to more excretion and to less efficient urea recycling and use (Robbins et al. 1974).

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Robbins, C. T., R. L. Prior, A. N. Moen, and W. J. Visek. 1974. Nitrogen metabolism of white-tailed deer. J. Anim. Sci. 38(1):186-191.

UNIT 1.2: ENVIRONMENTAL FACTORS

It is difficult to separate biological factors from environmental factors when evaluating protein metabolism. Crude protein in the diet is an environmental factor that causes changes in the efficiency of use by the biological organism. The stimulus is from the environment, the response is biological and internal.

Another environmental stimulus that affects protein metabolism, catabolism in this case, is extreme cold that places the animal in a critical thermal environment (Moen 1968). An animal in good condition mobilizes fat if a thermogenic metabolic response is necessary. If the fat reserve is depleted, protein tissue is the only internal substrate for thermogenic responses.

Weight losses accelerate when protein tissue must be oxidized as a source of energy. The main reason for this is that the energy content of a gram of fat is about 9.5 kcal, and of a gram of protein, 5.7 kcal. Further, a gram of protein is not metabolized completely by animals; only 4.7 kcal of heat energy are released as part of the heat energy remains in waste products (ammonia and urea) that are excreted and lost to the animal (Florey 1966:270). Thus heat production per unit of protein tissue weight is only about half as much as heat production per unit of fat tissue. Accelerated weight losses and quick death must occur when depressed nutrition and cold stress are combined. The WORKSHEET that follows provides some insight into the relative effects of energy metabolism on fat and protein substrate.

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CHAPTER 8, WORKSHEET 1.2a

The effects of energy metabolism from fat and protein substrates on weight losses

About 9.5 kcal of heat energy are released when a gram of fat is metabolized. About 5.7 kcal are released when a gram of protein is oxidized in a bomb calorimeter, and 4.8 kcal when a gram of protein is metabolized. Demonstrate the effect of a change in substrate from fat to protein when one-half of the animals metabolic requirements are met by metabolized fat and protein tissue.

Begin with an animal weighing 100 kg with a 10 kg fat reserve and metabolism of MBLM = 1.5 so that ELMD = $(1.5)(70)(100^{0.75})$. The resulting ELMD is 3320 kcal. If one-half is met by daily intake and the other half by fat reserves, the first day weight loss is 0.17 kg. Making the calculations for one week at a time, weight loss after one week is 1.22 kg. The new weight is 100 -1.22 = 98.78. ELMD is then, $(1.5)(70)(98.78^{0.75}) = 2193$, and one-half of that is 1097 kcal of energy to be met by fat. At 9500 kcal per kg, the weight loss is 0.12 kg per day or 0.81 per week. The next week's weight is 97.94 and the calculations are cycled through again. Continue until the fat is gone, plotting the weight loss per week and the live weight in kg (LWKG) at weekly intervals below.



Continue the calculations after the fat is gone and protein must be the substrate metaboized. Continue plotting the results on the grid on the next page.

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LWKG

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UNIT 1.3: INTERSPECIES COMPARISONS

General reviews of the literature indicate that protein metabolism processers are similar in both wild and domestic ruminants. Confined domestic animals are often given specific treatments to enhance growth, but such treatments do not alter basic metabolic processes. Urea and other non-protein nitrogen sources, for example, may be used as feed additives, even though domestic animals recycle urea naturally. Wild ruminants often do not not have access to foods high in protein or nitrogen, especially in the winter; urea recycling becomes a very important protein conservation adaptation then.

Endogenous urinary nitrogen has been measured in both domestic and wild ruminants. An equation for estimating endogenous urinary nitrogen in grams per day (EUNG) excreted, based on data in Crampton and Harris (1969:174) is:

$$EUNG = 0.002 \times 70 (MEWK)$$

where EUNG = endogenous urinary nitrogen in gms per kg metabolic weight per day, and

MEWK = metabolic weight in kg = LWKG^{0.75}.

The constant 0.002 represents the average ratio of nitrogen in gms to kcal basal metabolism.

EUNG calculated with this equation is larger than that determined for white-tailed deer by Robbins et al. (1974). The equation for deer is given in UNIT 2.1.

The use of first approximations of metabolic costs for wild ruminants based on data for domestic ruminants is preferable to saying "I don't know" when faced with the need to determine total daily costs of living. This is necessary when estimating protein costs, and also when estimating mineral, water, and vitamin requirements in CHAPTER 9.

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TOPIC 2. MEASUREMENTS OF PROTEIN METABOLISM

Protein metabolism proceeds at orderly rates in relation to the biological functions involved. The basis for the requirement of protein traces back to the losses of nitrogenous end-products from the body (Crampton and Harris 1969:166). If an animal is to be in protein balance, intake must equal output. When an animal is in a positive protein or nitrogen balance, more is being ingested in and deposited in body tissue than is lost. When an animal is in a negative protein or nitrogen balance, more is lost from the body than is ingested.

Protein is necessary for the maintenance of basic life processes, including the synthesis of enzymes, replacement of catabolized body tissue, replacement of tissue abraded from internal surfaces of the gastrointestinal tract, and from the skin. Measurements of endogenous urinary nitrogen have been made on few wild species. Domestic cattle and sheep have been studied rather extensively, and the results are useful for estimating the protein costs for wild ruminants.

Protein balances are rather delicate. Protein is not stored in large quantities like fat is, nor is the protein stored passively like fat. Body composition measurements, for example, show rather constant protein fractions, while fat and water fractions are inversely proportional as weight increases (see CHAPTER 2).

The basic biological functions of domestic ruminants and wild ruminants should be similar, so the results of measurements of protein metabolism on domestic ruminants may be approximations for wild ruminants by expressing relationships per unit body weight, metabolic body weight, or to some some other baseline.

Two kinds of measurements are discussed in the two units that follow. UNIT 2.1 includes discussions of direct measurements under controlled chamber or pen conditions, and UNIT 2.2 of indirect measurements, made by evaluating the apparent effects of different levels of protein intake or productivity.

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UNIT 2.1: DIRECT MEASUREMENTS AND CALCULATIONS

Protein requirements may be estimated directly by measuring the amount of and evaluating the protein fraction of selected metabolic products. Such measurements involve captive animals and feces and urine collections. The amounts of feces and urine and the metabolic nitrogen fraction of each of these waste products provides an indication of how much protein has been catabolized. Measurements of the amounts and protein fractions of accumulated body tissues provide a basis for estimating the protein required for growth. Considerations that must be made when evaluating these protein or nitrogen fractions are discussed next.

ENDOGENOUS URINARY NITROGEN

Nitrogenous compounds are one of the by-products of protein metabolism These compounds, such as ammonia and urea, must not be allowed to accumulate in the body as they can become very toxic. Urine is formed as a result of filtration, reabsorption, and secretion processes in the excretary system that result in the collection of nitrogen compounds that cannot be used (ammonia) or that are not recycled (urea).

The amount of urea recycled, a function of the crude protein in the diet, is variable and part of an adaptive strategy for nitrogen and protein conservation that has developed in free-ranging ruminants.

Endogenous urinary nitrogen, abbreviated EUNT, is derived from the catabolism of body tissue, and the quantity of EUNT is related to the metabolic body weight of the animal. An equation for calculating endogenous urinary nitrogen excreted by white-tailed deer, based on data in Robbins et al. (1974:189) is:

$EUNG = 0.115 \times (MEWK)$

where EUNG = endogenous urinary nitrogen in gms per kg per day, and MEWK = metabolic body weight in kg = $LWKG^{0.75}$.

The calculated protein requirement in gms per day to meet this loss of EUNG is:

PRUN = (0.115)(MEWK)(6.25)

where PRUN = protein required for endogenous urinary nitrogen.

The equation for EUNG given above, based on measurements on white-tailed deer fawns that were about 6 months old, may not be the best estimation for other species of wild ruminants, and ruminants of different ages. Other estimations may be made with the equation given on page 11 of this chapter, and with values published by the Agricultural Research Council (1965). Slight differences in the estimates from these two sources are discussed in Moen (1973:335). Estimates with the equation in Crampton and Harris (see page 11) are less than estimates based on the ARC values at the lower weights and greater at the higher weights for both cattle and sheep.

FECAL NITROGEN

Nitrogen in the feces comes from four sources. One, ingested but undigested protein in the diet ends up in the feces. Such protein compounds pass through the gastrointestinal tract intact. Two, nitrogen residues originating in digestive enzymes; three, protein compounds and nitrogen residues from the metabolic activities of the intestinal microflora, and four, protein compounds and nitrogen residues from the metabolic activities of the cells lining the gastrointestinal tract all end up in the feces. These different origins are not discerned when simply analyzing the feces for protein or nitrogen. Further, the third source includes protein or nitrogen from the metabolic functions of the intestinal microflora. The second and fourth sources are of metabolic origin, and represent protein metabolized by the body and used for maintenance and productive functions.

Metabolic fecal nitrogen results from the abrasion of epithelial cells from the gastrointestinal tract as food passes through and from proteins used in the digestion of food. The quantity of metabolic fecal nitrogen is a function of both food intake and the quality of the ingested food.

The usefulness of ingested protein to the animal is expressed as the biological value of the protein. The true biological value is defined by Crampton and Harris (1969:89) as the "percentage of true absorbed nitrogen that is used for maintenance and production, or for maintenance only." It is related to the content of essential amino acids (those amino acids which must be ingested because they cannot be synthesized in the body).

Suppose the protein contained a perfect assortment of amino acids. Then, all of the protein could be used by the body and none would be deaminized and its nitrogen component excreted in the urine. The urine would contain only nitrogen of metabolic origin. Suppose that the protein in a food did not contain a perfect assortment of amino acids. Then, only a fraction of the protein could be used, with the rest deaminized and the nitrogen excreted. This fraction times the protein digestibilility results in a net protein coefficient (NPRC), which is a measure of the amount of protein that can be used for metabolic purposes. Calculations of net protein coefficients are illustrated below.

Protein characteristics of the food	Amino acid <u>"index"</u>		<u>,</u>	NPRC	
Perfect amino acid distribution,		·			0.05
high digestibility Good amino acid	1.00	x	0.95	=	0.95
distribution, medium digestibility	0.80	x	0.70	=	0.56
Poor amino acid distribution, low digestibility	0.50	x	0.50	=	0.25

These illustrations show how the amino acid distribution and digestibility interact (Oser 1951) to result in a net protein coefficient (NRPC). As NPRC decreases, food intake must increase if nitrogen requirements are to be met. The additional intake, however, raises MFNG requirements. Thus deteriorating protein quality of forage causes an increase in protein requirements, placing the animal in a negative feedback situation that results in low rates of ingestion and a negative protein balance.

GROWTH

Animals that are in a positive nitrogen balance are depositing more protein in body tissue than is being excreted. Protein is very important for body growth in young animals, it is deposited in hair as coat changes occur twice each year, and it is an important cost during reproduction as protein is an important component of fetal tissue and milk.

The protein costs associated with these productive processes may be estimated by determining the protein compositions and the quantities of the different tissues produced. Results of these "before and after" measurements may be expressed on a per day basis. Growth and chemical composition data were given in CHAPTERS 1 and 2. These data are needed when completing the WORKSHEETS in TOPIC 3.

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- Oser, B. L. 1951. Methods for integrating essential amino acid content in the nutritional evaluation of protein. J. Amer. Dietetic Assoc. 26:396.

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DIRECT MEASUREMENTS AND CALCULATIONS

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JANSA 38--1 186 191 odvi nitrogen metabolism, deer robbins,ct; prio/ 1974 JWMAA 39--3 582 589 odvi protein requirement, fawns smith,sh: holter/ 1976 JWMAA 43--4 872 879 odvi protein require, yearlings holter,jb; hayes/ 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 155-7 1085 1085 odhe urine-collect device, male richmond,m; pill/ 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BJNUA 24--3 843 855 ceel dosh, digest, nitrogen met maloiy,gmo; kay,/ 1970 CBPAB 60A-3 251 256 ceel nitrogen metabolism, red d simpson,am; webs/ 1978

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UNIT 2.2: FEEDING TRIALS

Feeding trials which give indications of the levels of dietary protein necessary for different growth and reproductive responses may be completed for captive animals in order to get some estimate of the protein fraction of the forage required for different levels of productivity. These are "black box" type experiments; inputs are compared to outputs without evaluating the mechanisms within the animal that result in the partitioning of the nutrients. Further, interactions between different nutrients are difficult if not impossible to consider and control.

Comparisons may be made between the results of feeding trials and the summation of different protein costs determined by measurements or calculations of the metabolic characteristics discussed in UNIT 2.1. The two independent approaches should have similar results, of course. Such comparisons are useful because they serve as checks on each other; "ecological accounting" is an important part of the many sequential calculations used to link animal to range in as many ways as possible.

REFERENCES, UNIT 2.2

FEEDING TRIALS

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 31--4 679 685 odvi protein requirement, fawns ullrey,de; youat/ 1967 JWMAA 43--4 872 879 odvi protein require, yearlings holter,jb; hayes/ 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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Chapter 8 - Page 21

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TOPIC 3. ESTIMATES OF PROTEIN COSTS FOR MAINTENANCE AND PRODUCTION

Estimates of protein costs for production are made whenever new tissue is produced which contains protein. New body tissue resulting from growth, fetal tissue, hair, and milk are all protein-containing tissue, and the protein in these tissues must come from either the diet or from the catabolism of body protein. Since body protein stores are not extensive, the diet is the main source of protein for production.

A general rule to follow when estimating the protein costs for production is that the cost is at least equal to the amount of protein in the new tissue. Then, there is an additional cost associated with the assimilation of that tissue, an "overhead" that results from the less than 100% efficiency of metabolic processes.

The UNITS in this TOPIC include descriptions of the protein-requiring productive processes characteristic of wild ruminants. The equations given may be used for first approximations when evaluating the total protein requirements of individuals at different times during their annual cycles.

UNIT 3.1 ENDOGENOUS URINARY NITROGEN

Some of the nitrogen excreted in the urine is derived from the catabolism of body tissues, and has traditionally been called endogenous urinary nitrogen. Such nitrogen is of metabolic origin; it has been assimilated into body tissue from dietary nitrogen, and then released again when the body tissue is catabolized. This quantity of endogenous urinary nitrogen has been considered a minimum daily requirement. A formula for estimating endogenous urinary nitrogen (EUNG) based on data in NRC (1976) and Moen (1973:335) is:

EUNG = (a)(MEWK)

where EUNG = endogenous urinary nitrogen in gms and MEWK = metabolic weight in kg.

The protein required to replace the endogenous urinary nitrogen excreted, PRUN, in gms per day, is:

PRUN = (a)(MEWK)(6.25)

The value of (a) in the formulas above is slightly different in different sources. In Crampton and Harris (1969:174) it is 0.140. In the NRC (1976:6) publication, it is 0.120. In Robbins et al. (1974:189) it is 0.115. The equations for calculating endogenous urinary nitrogen are, then:

> General: EUNG = $0.140 \text{ LWKG}^{0.75}$ Beef cattle: EUNG = $0.120 \text{ LWKG}^{0.75}$ White-tailed deer: EUNG = $0.115 \text{ LWKG}^{0.75}$

> > Chapter 8 - Page 23

The differences in these three equations are in the value of a; the general equation results in a larger estimate than the beef cattle and white-tailed deer equations. Comparisons of absolute values are made in a WORKSHEET.

LITERATURE CITED

- Crampton, E. W. and L. E. Harris. 1969. Applied animal nutrition. W. H. Freeman and Co., San Francisco. 753 pp.
- Moen, A. N. 1973. Wildlife ecology. W. H. Freeman and Co., San Francisco. 458 pp.
- National Research Council. 1976. Nutrient requirements of beef cattle. Number 4. National Academy of Sciences, Washington D. C. 56 pp.
- Robbins, C. T., R. L. Prior, A. N. Moen, and W. J. Visek. 1974. Nitrogen metabolism of white-tailed deer. J. Anim. Science 38(1):186-191.

REFERENCES, UNIT 3.1

ENDOGENOUS URINARY NITROGEN

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CHAPTER 8, WORKSHEET 3.1a

Estimates of endogenous urinary nitrogen

Endogenous urinary nitrogen of white-tailed deer is dependent upon the weight of the deer and can be calculated from the ingesta-free weight with an equation from Robbins et al. (1974):

 $EUNG = 0.115 (LWKG^{0.75})$

where EUNG = endogenous urinary nitrogen in grams and LWKG = live weight in kg.

The NRC (1976:6) equation for beef cattle is:

 $EUNG = 0.120 \ (LWKG^{0.75})$

The general equation based on Crampton and Harris (1969:174) is:

 $EUNG = 0.140 (LWKG^{0.75})$

Plot the results using these equations below. Note that LWKG of deer is seldom over 100 kg. Beef cattle reach 500 kg or more at maturity.



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Crampton, E. W. and L. E. Harris. 1969. Applied animal nutrition. W. H. Freeman and Co., San Francisco. 753 pp.

National Research Council. 1976. Nutritent requirements of beef cattle. Number 4. National Academy of Sciences, Washington, D. C. 56 pp.

Robbins, C. T., R. L. Prior, A. N. Moen, and W. J. Visek. 1974. Nitrogen metabolism of white-tailed deer. J. Anim. Sci. 38(1):186-191.

Chapter 8 - Page 26a

CHAPTER 8, WORKSHEET 3.1b

Protein required to meet EUNG losses

The protein equivalent of endogenous urinary nitrogen losses is determined by multiplying EUNG by 6.25, according to the formula on page 23. Select the weight range used, determine the amounts of protein required for endogenous urinary nitrogen losses using the 3 equations in the previous WORKSHEET, label the x axis (LWKG) and y axis (PRUN), and plot the protein required for endogenous urinary nitrogen in the grid below.



Chapter 8 - Page 26b

UNIT 3.2: METABOLIC FECAL NITROGEN

The nitrogen in the feces of dietary origin that has been assimilated into body tissue and then broken down again to appear in the feces is called metabolic fecal nitrogen. This category includes nitrogen from cells that have been absorbed from the linings of the gastrointestinal tract nitrogen from the protein compounds and nitrogen residues originating in secreted enzymes, and residual protein and nitrogen in intestinal microflora that ends up in the feces.

Metabolic fecal nitrogen in grams per day (MFNG) is related to food intake by the following formula modified from Moen (1973:366).

MFNG = k DWFK

where k = 5 and DWFK = dry weight forage in kg. DWFK necessary for maintenance is related to body weight, and has been calculated in previous UNITS in relation to energy metabolism. The calculated DWFK may be used as a starting point for calculating the protein requirements to meet quantities of metabolic fecal nitrogen excreted. If the calculated amount of food necessary to meet the energy requirements is low in protein, protein requirements may not be met by the predicted DWFK. This depends in part on the net protein coefficients of the forage ingested.

Metabolic fecal nitrogen for beef cattle given in the NRC publication (1976:6) is simply 4 g N/kg feed dry matter. Expressing this as an equation:

MFNG = (4)(DMIK)

where MFNG = metabolic fecal nitrogen in grams per day, DMIK = dry matter intake in kg per day.

The value of "k" is variable in a formula in Moen (1973:336), based on the relative amounts of milk and forage in the diet. The values are:

Note that the equations above relate metabolic fecal nitrogen to dry matter intake. As more forage is ingested, more nitrogen is released internally from the sources mentioned above to process the forage. MFNG is less when milk is ingested (c = 2.5) because fluid milk is more easily digested.

Actual calculations of MFNG are dependent on DMIK. Yet DMIK is dependent on energy and protein requirements, including MFNG. This feed-back mechanism suggests a need for an iteration procedure in final determinations of MFNG. Such a procedure is illustrated in a WORKSHEET.

LITERATURE CITED

- Moen, A. N. 1973. Wildlife ecology. W. H. Freeman and Co., San Francisco. 458 pp.
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REFERENCES, UNIT 3.2

METABOLIC FECAL NITROGEN

SERIALS

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Chapter 8 - Page 28

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CHAPTER 8, WORKSHEET 3.2a

Estimates of metabolic fecal nitrogen

Estimate the metabolic fecal nitrogen in grams with the following equation:

MFNG = (5)(DMIK)

Suppose 9 gms of nitrogen were required for all body processes except Suppse that the crude protein content of the forage was 12%. Then, MFNG. one kg of forage would contain 120 gms of protein. This is equal to 19.2 gms of nitrogen $(120/6.25 \text{ or } 120 \times 0.16 = 19.2)$. Suppose the net protein coefficient of this forage is 0.70. The amount of forage needed to supply 9 gms of nitrogen is (9.0)(1/0.70)/19.2 = 0.67 kg = DMIK. MFNG = (5)(0.67) =Thus the total nitrogen required, including MFNG, is now 9.0 + 3.353.35. Substituting 12.35 for 9.0, DMIK is now calculated by 12.35. (12.35)(1/0.70)/19.2 = 0.92. MFNG = (5)(0.92) = 4.60, which is added to 9.0, and the total nitrogen requirement, including MFNG, is now 9.0 + 4.60Substituting 13.60 for 9.0, DMIK is now calculated by = 13.60.(13.60)(1/0.70)/19.2 = 1.01. MFNG = (5)(1.01) = 5.05, and 9.0 + 5.05 =14.05. Substituting 14.05 for 9.0, DMIK = (14.05)(1/0.70)/19.2 = 1.05.

Note how DMIK began at 0.67, and successive calculations of the MFNG resulted in successive approximations of DMIK = 0.92, 1.01, and 1.05. Note that DMIK is getting larger due to the addition of MFNG requirements, but that increments are getting smaller with each cycle. The last calculation resulted in an increment of less than 4%; the next calculation [(5)(1.05)...] results in an increment of less than 1%.

The protein required to meet this metabolic fecal nitrogen requirement may be calculated with the following equation, assuming that the protein contains 16% nitrogen:

PMFN = 6.25(MFNG)

where PMFN = protein required for metabolic fecal nitrogen.

Review this sequence so you understand it thoroughly. The iterations being completed result in the estimation of total nitrogen requirements, including the effects of metabolic fecal nitrogen on dry-matter intake, and the feed-back effects of dry matter intake on metablic fecal nitrogen.

Repeat these calculations with a forage that is low in crude protein say less than 5%. Depending on the value of NPRC, there is a level of crude protein at which ingestion rates must increase in order to keep up with the MFNG costs. Above that level, ingestion is counter-productive; metabolic processing costs more than is returned from the forage.

Urea recycling and changes in the biological value of the protein (hence NPRC) makes the biological process more complex than this mathematical process. The iterations described do help one understand a bit more about the balance between nutrient requirements and nutrient densities in the forage.

Chapter 8 - Page 30a

UNIT 3.3: PROTEIN REQUIRED FOR BODY GROWTH

The daily protein required for gain is a function of the amount of total gain per day and the composition, or fraction of the gain that is protein. Equations for body composition were given in CHAPTER 2, and the protein fraction equation is used here to calculate this requirement. The daily change in weight multiplied by the protein fraction of animals at this weight is the daily protein deposited, which is the minimum protein required for body growth. The efficiency of protein deposition affects the actual amount of protein required.

The protein composition of steers weighing 100 kg is 18 percent, and of those weighing 500 kg, 9%. Corresponding values for heifers are 18% and 7%, respectively. Note that the percent composition is higher for smaller and younger cattle that are still growing, and lower for the larger and older cattle that are more physically mature. The protein composition of white-tailed deer was much more constant for a wide range of ages and weights measured by Robbins et al. (1974). In fact, the protein content of males and females combined was 20% of the body weight at 6 and 60 kg body weight. Wild ruminants are expected to have a more constant protein fraction since they are neither bred nor raised for rapid gains which under confined conditions may result in a higher fat and lower protein content in the body.

LITERATURE CITED

Robbins, C. T., A. N. Moen, and J. T. Reid. 1974. Body composition of white-tailed deer. J. Anim. Sci. 38(4):871-876.

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PROTEIN REQUIREMENTS FOR BODY GROWTH

SERIALS

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Chapter 8 - Page 31

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CHAPTER 8, WORKSHEET 3.3a

Estimates of the protein requirements of white-tailed deer for body growth

The protein fraction of body growth is obviously the minimum protein required for body growth, with increases related to the efficiency of protein deposition during growth. The protein fractions of white-tailed deer of different weights from June through October were determined by Robbins et al. (1974). The equation for calculating the protein component of the body weight in kg (males and females combined), rearranged from the published paper. is:

 $PCBK = e^{1.0091} \ln IFWK - 1.6267$

where PCBK = protein component of the body in kg, and IFWK = ingesta-free weight in kg.

The equation for the protein component of males and females combined, expressed as a fraction of ingesta-free weight, is:

 $PRFR = e^{1.0091} \ln IFWK - 1.6267/IFWK$

If IFWK = 6.0 and PCBK = 1.2, then PRFR = 1.2/6.0 = 0.20. If IFWK = 34.0 and PCBK = 6.9, then PRFR = 6.9/34.0 = 0.20. If IFWK = 68.0 and PCBK = 13.9, then PRFR = 13.9/68.0 = 0.20.

Thus the protein fraction is a constant 0.20 (or 20%) of the ingesta-free weight for a wide range of weights. The equation for the protein required for body growth is, then:

PRBG = (0.20)(DAGA)(1/PEFG)

where PRBG = protein required for body growth, DAGA = daily gain, and PEFG = protein efficiency of body growth.

The nitrogen component in kg (NTCK) is simply PRCK/6.25. Since the protein fraction is 0.20, the nitrogen fraction is 0.032, or 3.2% of the body. This is higher than the 2.5% estimate given in Moen (1973:336), based on data on domestic animals.

Protein and nitrogen components may be plotted on the grid on the next page for the range of ingesta-free weights selected. The axes are unlabeled since the 20% figure for white-tailed deer may be used as a first approximation for other wild ruminants of different weights. In other words, body composition of any wild ruminant may be estimated to include 20% protein, and this may be a better value than those based on domestic ruminants when none are available for particular wild species.

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Literature Cited

Robbins, C. T., A. N. Moen, and J. T. Reid. 1974. Body composition of white-tailed deer. J. Anim. Sci. 38(4):871-876.

UNIT 3.4: PROTEIN REQUIRED FOR HAIR GROWTH

Wild ruminants have two coats per year as a heavy winter coat is replaced by a light summer coat. Some species show color changes that are associated with the coat changes too. Physical and chemical characteristics of the hair coats and molting were discussed in CHAPTER 2, and the data given there on protein content of hair of different species should be used to estimate the cost of hair growth.

The overall nitrogen loss through hair and surface cells is calculated for cattle as: grams $N = 0.02 W^{0.75}$ (NRC 1976:6). This expresses the grams of nitrogen required for hair and surface cells each day as a directy proportion of metabolic body weight. Rewritten with four-letter symbols:

NRHG =
$$0.02 \text{ IFWK}^{0.75}$$

where NRHG = nitrogen required for hair growth and IFWK = ingesta-free weight in kg.

The protein required for hair growth (PRHG) is 6.25 times NRHG; therefore:

$$PRHG = 0.125 \ TFWK^{0.75}$$

Both NRHG and PRHG are in grams.

These equations result in average costs over the entire year. Estimates of the cost of hair growth per day for wild ruminants must consider changes in the growth rate of hair from season to season. The winter coats of North American wild ruminants begin growing in August or September, and growth is completed by November or December. The summer coat begins growing in April and May, and growth is completed by June. After growth is completed, the protein is irreversibly present in the hair, and represents a cost that is met twice each year.

The protein required for hair growth may be calculated with rates of hair growth and composition of the hair, just as the cost of body growth was calculated. Daily changes in the amounts of hair times the protein fraction of the hair are necessary inputs when calculating the daily protein cost of hair growth, just as in calculations of the costs of body growth. These options are presented in WORKSHEETS.

LITERATURE CITED

National Research Council. 1976. Number 4. Nutrient requirements of beef cattle. 5th Rev. Ed. National Academy of Sciences, Washington, D. C. 56 pp.

REFERENCES, UNIT 3.4

PROTEIN REQUIRED FOR HAIR GROWTH

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JANSA 38--4 871 876 odvi body composition of white- robbins, ct; moen/ 1974 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR odhe CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ceel CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR alal CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR rata CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JANSA 45--4 826 831 doca hair indic calor-prot stat haaland,g1; mats/ 1977

Chapter 8 - Page 36

Chapter 8, Worksheet 3.4a

Estimates of the protein requirements for hair growth

The protein fraction of the hair is obviously the minimum protein required for hair growth, with increases related to the efficiency of protein deposition during hair growth. The protein fraction of hair of white-tailed deer was given by Robbins et al. (1974) and discussed in CHAPTER 2, UNIT 1.3, Pages 11-14. Knowing daily changes in hair weights, the protein composition of hair, and the physiological efficiency of hair growth, the protein cost of hair growth may be calculated with the formula:

PCHG = (DLTA HAWG)(PRFH)(1/PEHG)

where PCHG = protein cost of hair growth in gms per day, DLTA = delta, or daily change, HAWG = hair weight in gms, PRFH = protein fraction of the hair, and PEHG = physiological efficiency of hair growth.

Refer back to CHAPTER 2 for data on hair growth, and then calculate PCHG. Compare the results over the annual cycle with the protein costs estimated with the formula for cattle (see page 31). An unlabeled grid is provided for plotting the results for the species of your choice.



LITERATURE CITED

Robbins, C. T., A. N. Moen, and J. T. Reid. 1974. Body composition of white-tailed deer. J. Anim. Sci. 38(4):871-876.

UNIT 3.5: PROTEIN REQUIRED FOR GESTATION

The minimum protein required for gestation is equal to the amounts deposited in fetal and associated reproductive tissues, with increases in proportion to the rate of physiological efficiency of protein deposition for this purpose. Protein requirements increase rapidly in the last two-thirds of the gestation period.

The total nitrogen content of the gravid uterus of white-tailed deer has been determined by Robbins and Moen (1975). The published equation (page 688) is modified to:

NCUG =
$$e^{0.2096 + 0.0275 \text{ DIGE}}$$

where NCUG = nitrogen content of the uterus in gms, and DIGE = days into gestation.

Nitrogen retention may be calculated by subtracting NCUG for successive days. An equation has already been determined for this by Robbins and Moen (1975) for white-tailed deer, and another equation given that may be adapted to any species. The equation for estimating nitrogen retention for gestation per day (NRGD) in white-tailed deer, modified from Robbins and Moen (1975:688), is:

$$NRGD = e^{0.0275 DIGE - 3.3856}$$

where NRGD = nitrogen required for gestation (gms per day), and DIGE = days into gestation.

The equation for any species, modified from Robbins and Moen (1975:688), is:

 $NRGD = [e^{0.0512} (DIGE/LEGP)100 - 1.7274] [BIWK]$

where NRGD = nitrogen required for gestation (gms per day), DIGE = days into gestation, LEGP = length of the gestation period, and BIWK = birth weight in kg.

The above equation is particularly useful since it expresses a proportional nitrogen cost in relation to length of the gestation period and fetal weight at birth. Cattle, sheep, and deer were quite similar, and there is no reason to believe that the wild ruminants would be very different in this basic biological process.

WORKSHEETS provide opportunities to calculate the protein costs of gestation, expressed both as protein and nitrogen.

LITERATURE CITED

Robbins, C. T. and A. N. Moen. 1975. Uterine composition and growth in pregnant white-tailed deer. J. Wildl. Manage. 39(4):684-691.

REFERENCES, UNIT 3.5

PROTEIN REQUIRED FOR GESTATION

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 39--4 684 691 odvi uterin comp, growth, pregn robbins,ct; moen, 1975 NAWTA 31--- 129 139 odvi effects of dietary protein murphy,da; coates 1966

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR odhe

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CHAPTER 8, WORKSHEET 3.5a

Estimates of the protein required for gestation

Estimates of the daily protein required for gestation may be made with the equations in Robbins and Moen (1975). Since the protein required is 6.25 x nitrogen, the equations below include this conversion factor. The equations are:

odvi PRGE = $(e^{0.0275 \text{ DIGE} - 3.3856})$ (6.25)

any species: PRGE = [e^{0.0512} (DIGE/LEGP)100 - 1.7274] [BIWK] (6.25)

where PRGE = protein required in for gestation in grams per day, DIGE = days into gestation, LEGP = length of the gestation period, and BIWK = birth weight in kg.

The two reproductive system variables needed (LEGP and BIWK) to calculate PRGD were given in CHAPTERS 1 and 2.

Calculate PRGD for intervals of DIGE and plot in the grid below. The results are minimum values; physiological efficiencies less than 1.0 will increase the actual cost per day of nitrogen retention.



DIGE =

LITERATURE CITED

Robbins, C. T. and A. N. Moen. 1975. Uterine composition and growth in pregnant white-tailed deer. J. Wildl. Manage. 39(4):684-691.

Chapter 8 - Page 40a

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UNIT 3.6: PROTEIN REQUIRED FOR MILK PRODUCTION

The protein required for milk production is a function of the amount of milk produced, the protein content of the milk, and the physiological efficiency of lactogenesis. Milk is relatively high in protein, a biological necessity for the rapid growth of the neonate.

Milk production was discussed in CHAPTER 7, UNIT 3.4, with calculations of milk production in gms per day given in WORKSHEET 3.4a (Chapter 7 - Page 48a). The chemical composition of milk was discussed in CHAPTER 2, UNIT 2.2. After data on these characteristics have been determined, the protein cost of lactation per day may be estimated as at least the amount of protein in the milk, with increases in the protein required due to the physiological efficiency of lactogenesis.

The WORKSHEET at the end of this UNIT provides opportunities to synthesize concepts, data, and equations from previous CHAPTERS while estimating protein required for lactation.

REFERENCES, UNIT 3.6

PROTEIN REQUIRED FOR MILK PRODUCTION

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 25--1 66 70 odvi deer milk, substitute milk silver,h 1961 JWMAA 29--1 79 84 odvi comp milk,bld nursng doe,f youatt,wg; verme/ 1965 NAWTA 31--- 129 139 odvi effects of dietary protein murphy,da; coates 1966

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CAFGA 37--2 217 odhe composit, deer milk, calif hagen, hl 1951 218 JOMAA 36--3 473 474 odhe mule deer milk browman, lg; sears 1955 JOMAA 58--3 420 423 odhe changes nutri compos, milk mueller, cc; sadle 1977 JWMAA 20--2 212 214 odhe postnat grow, comp of milk kitts,wd; cowan,/ 1956 JWMAA 44--2 472 478 odhe milk yield, black-tailed d sadleir, rmfs 1980

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BIJOA 153-3 647 655 ceel whey proteins, red de milk mcdougall,ei; ste 1976 JRPFA 37--1 67 84 ceel comp, yield of milk, red d arman,p; kay,rnb/ 1974 ZTTFA 31--5 227 238 ceel comp, milk, red deer, pt 1 brueggemann,j; d/ 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--2 213 215 alal gros comp milk, fat, minrl cook, hw; rausch,/ 1970 JWIDA 12--2 202 207 alal milk, hair, element relati franzmann, aw; f1/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 45--6 1101 1106 rata milk, gros comp, fat, prot hatcher,vb; mcew/ 1967 JDSCA 57-11 1325 1333 rata milk comp chang, grazing r luick,jr; white,/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CAFGA 37--2 217 218 anam odhe, compos milk, compars hagen, hl 1951

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 36--2 305 308 bibi the lipids in bison bison wilbur,cg; gorski 1955

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 43--5 885 888 ovca milk, gross comp, fat cons chen, ech; blood,/ 1965

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--4 629 633 ovda milk, stage lact, composit cook, hw; perarso/ 1970 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 34--6 569 571 obmo gross composition of milk tener, js 1956 CJZOA 48--6 1345 1347 obmo gros comp, ftty acid, minrl baker, be; cook, h/ 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 47--1 5 8 oram gross compos, fat acid con lauer, bh; blood,/ 1969 CJZOA 47--2 185 187 oram miner const, milk, arctic luer, bh; baker, b 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR CBCPA 42b-2 323 328 many electrophesis milk caseins lauer, bh; baker, b 1972 CJZOA 49--4 551 554 many carbohydra content, casein baker, be; lauer, b 1971 CJZOA 55--1 231 236 many amin acid comp casein milk lauer, bh; baker, b 1977 IZYBA 4---- 333 342 many composit milk wild animals ben shaul, dm 1962

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JRPFA 37--1 67 84 cerv composi milk, cerv species arman,p; kay,rnb/ 1974

CHAPTER 8, WORKSHEET 3.6a

Estimating the protein required for milk production

The protein required for milk production may be calculated by multiplying the amount of milk produced by the protein fraction of the milk by the reciprocal of the physiological efficiency of protein synthesis for milk production. In symbol form:

PRMP = (QMPK)(PRFM)(1/PEMP)(1000)

where PRMP = protein required for milk production in gms per day, QMPK = quantity of milk produced in kg per day, PRFM = protein fraction of milk, PEMP = physiological efficiency of milk production, and 1000 = the conversion factor from kg to gms.

These parameters have all been discussed in previous chapters. For QMPK, refer to CHAPTER 7; for PRFM, refer to CHAPTER 2; and for PEMP, use 0.5 to 1.0. PRFM may change during the lactation period. Then, PRFM = f(DILA), where DILA is days into lactation, and an equation replaces a single value for PRFM.

Calculate PRMP and plot the results on the grid provided. The x-axis is PRFM or DILA, the y-axis PRMP, and PEMP effects a family of curves.

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UNIT 3.7: SUMMATIONS OF PROTEIN REQUIRED

Equations for several different protein costs have been presented in this CHAPTER. If the concepts underlying each of these are understood and the costs calculated, then the total cost can be determined by summation. Addition of the separate costs is simple, with one main precaution.

The main precaution is that the maintenance costs must be either treated separately or as part of particular production costs, but not both. Some of the endogenous urinary nitrogen, for example, may come from metabolic transactions involved with gestation. If EUNG includes nitrogen from fetal metabolism, then the protein costs of fetal growth should include only the net costs. This is an important concept, and one that must be applied in each calculation from each published source. Procedures for expressing costs are not standardized, of course, so careful reading of each paper is necessary to determine just what is included in the results.

Summations may be made for protein costs throughout the year by developing the biochronology of each species and making the appropriate calculations through time. When these are plotted over the annual cycle, a pattern emerges that may be represented by a single equation as for energy metabolism (See Moen 1978 and CHAPTER 7). I must admit that at the time of this writing I have not made such comprehensive calculations for protein costs.

One WORKSHEET with reminders of the cost items to include in the summation of protein costs is included at the end of this unit.

REFERENCES, UNIT 3.7

SUMMATIONS OF PROTEIN REQUIRED

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR JWMAA 31--4 679 685 odvi protein requirement, fawns ullrey,de; youat/ 1967 JWMAA 43--4 872 879 odvi protein require, yearlings holter,jb; hayes/ 1979 PAABA 600-- 1 50 odvi nutr req growth, antl dev french,ce; mcewa/ 1955 CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR NAWTA 22--- 119 132 od-- nutrient requirements deer mcewan,lc; frenc/ 1957 NAWTA 22--- 179 188 od-- feed req for growth, maint cowan,i mct; woo/ 1957

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR cee1 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR alal CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 48--5 905 913 rata seas chan, energ, nitr int mcewan, ch; wood, a 1970 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovđa CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram

CHAPTER 8, WORKSHEET 3.7a

Estimates of total protein costs by summation

Total protein costs may be estimated by summing the costs for each of the maintenance and production processes. The costs to include are, in the order presented in this CHAPTER:

PRUN	(WS	3.1b)
+ PMFN	(WS	3.2a)
+ PRBG	(WS	3.3a)
+ PRHG	(WS	3.4a)
+ PRGE	(WS	3.5a)
+ PRMP	(WS	3.6a)
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TOTAL = = PRGD

Calculations of the appropriate costs in relation to the biochronology at 7-day intervals will provide estimates of costs throughout the annual cycle. The patterns that emerge for different sex and age groups should be related to range characteristics and forage conposition discussed in PART IV. An unlabeled grid is provided for plotting results of the summations.



CLOSING COMMENTS

The basic format of CHAPTER 8 on protein metabolism is similar to that of CHAPTER 7 on energy metabolism. Estimates of protein costs have been described which provide first approximations at least. Some equations are based on well-designed experiments on wild ruminants. It is important to realize that biological costs to the animal go on whether we realize it or not. Therefore, first approximations are better than nothing, and students should not hesitate to include such estimates when necessary to maintain the integrity of the biological concepts. Concepts are fundamental; numbers and equations may be changed.

> Aaron N. Moen March 11, 1981

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GLOSSARY OF SYMBOLS USED - CHAPTER EIGHT

BIWK = Birth weight in kilograms BVAP = Biological value in percentage DAGA = Daily gainDCCP = Dietary crude protein DIGE = Days into gestation DILA = Days into lactation DMIK = Dry matter intake in kilograms per day DLTA = DeltaDWFK = Dry weight of forage in kilograms ELMD = Ecological metabolism per day EUNG = Endogenous urinary nitrogen in grams per day EUNT = Endogenous urinary nitrogen HAWG = Hair weight in grams IFWK = Ingesta-free weight in kilograms LEGP = Length of gestation period LWKG = Live weight in kilograms MBLM = Multiple of base-line metabolism MEWK = Metabolic weight in kilograms MFNG = Metabolic fecal nitrogen NCUG = Nitrogen content of the uterus in grams NRGD = Nitrogen required for gestation per day NRHG = Nitrogen required for hair growth per day NRPC = Net protein coefficient NTCK = Nitrogen component in kilograms PCBK = Protein component of the body in kilograms PCHG = Protein cost of hair growth in grams per day PEFG = Protein efficiency for body growth PEHG = Physiological efficiency of hair growth PEMP = Physiological efficiency of milk production PNDM = Percent nutrients derived from milk PRBG = Protein for body growth PRCK = Protein component in kilograms PRFH = Protein fraction of hair PRFR = Protein fraction PRFM = Protein fraction of milk PRGD = Protein required in grams per day PRGE = Protein required for gestation in grams per day PRHG = Protein required for hair growth in grams per day PRMP = Protein fraction for milk production in grams per day PRUN = Protein required for endogenous urinary nitrogen QMPK = Quantity of milk produced in kilograms per day UREP = Urea recycled in percent WLPW = Weight loss per week

GLOSSARY OF CODE NAMES - CHAPTER EIGHT

CODEN

Australian Journal of Biological Sciences AJBSA AJPHA American Journal of Physiology AZOFA Annales Zoologici Fennici BIJOA Biochemical Journal BJNUA British Journal of Nutrition CAFGA California Fish and Game CBCPA Comparative Biochemistry and Physiology CJZOA Canadian Journal of Zoology IZYBA International Zoo Year Book JANSA Journal of Animal Science JASIA Journal of Agricultural Science JAVMA Journal of the American Veterinary Medical Association JDSCA Journal of Dairy Science JÔMAA Journal of Mammalogy JONUA Journal of Nutrition JRPFA Journal of Reproduction and Fertility JWIDA Journal of Wildlife Diseases JWMAA Journal of Wildlife Management MAAIA Journal of the Scientific Agricultural Society of Finland NAWTA North American Wildlife and Natural Resources Conference, Transactions of the, NUMEB Nutrition and Metabolism PAABA Pennsylvania Agricultural Experiment Station Bulletin PAANA Proceedings of the Australian Society of Animal Production PAARA Pennsylvania State University College of Agriculture Agricultural Experiment Station Progress Report

ZTTFA Zeitschrift fuer Zellforschung und Mikroskopisch Anatomie

LIST OF PUBLISHERS - CHAPTER EIGHT

acpr	Academic Press	New York	nyny
base	Base1	New York	nyny
blsp	Blackwell Scientific Publications	Oxford, England	oxen
butt	Butterworth	London, England	loen
ccth	Charles C. Thomas	Springfield, IL	spil
cfmo	Carl Fr. Mortensen	Copenhagen Denmark	code
clpr	Clarendon Press	Oxford, England	oxen
dcch	D. C. Church, Oregon State	Corvallis, OR	coor
else	Elsevier	Springfield, VA	spva
hein	Heinemann	London, England	loen
meth	Methuen & Co., Ltd.	London, England	loen
nhpc	North-Holland Publishing	New York	nyny
rupr	Rutgers University Press	New Brunswick, NJ	nbnj

LIST OF WORKSHEETS - CHAPTER EIGHT

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1.la	Urea recycling and the biological value of protein metabolized by white-tailed deer (odvi) 8a
1.2a	The effects of energy metabolism from fat and protein substrates on weight losses
3.la	Estimates of endogenous urinary nitrogen
3.2a	Estimates of metabolic fecal nitrogen
3.3a	Estimates of the protein requirements of white-tailed deer
JUJU	for body growth
3.4a	Estimates of the protein required for hair growth
3.5a	Estimates of the protein required for gestation
3.6a	Estimating the protein required for milk production

JULIAN DAY: MONTH AND DAY EQUIVALENTS

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Day		Jan	Feb	Mar	Apr	May	Jun	Ju1	Aug	Sep	Oct	Nov	Dec	Day
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5		005	036	064	095	125	156	186	217	248	278	309	339	5
6		006	037	065	096	126	157	187	218	249	279	310	340	6
7		007	038	066	0 9 7	127	158	188	219	250	280	311	341	7
8		008	039	067	09 8	128	159	189	220	251	281	312	342	8
9		009	040	068	099	129	160	190	221	252	282	313	343	9
10		010	041	0 6 9	100	130	161	191	222	25 3	283	314	344	10
11		011	042	070	101	131	162	1 92	223	254	284	315	345	11
12		012	043	071	102	132	163	193	224	255	285	316	346	12
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16		016	047	075	106	136	167	197	228	259	289	320	350	16
17		017	048	076	107	137	168	198	229	260	290	32 1	351	17
18		018	049	077	108	138	169	199	230	261	291	322	352	18
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23		023	054	082	113	143	174	204	235	266	296	327	357	23
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25		025	056	084	115	145	176	206	237	268	29 8	329	3 59	2 5
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THE BIOLOGY AND MANAGEMENT OF WILD RUMINANTS

CHAPTER NINE

MINERAL, WATER, AND VITAMIN METABOLISM OF WILD RUMINANTS

Ъy

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Published by

CornerBrook Press Box 106 Lansing, N.Y. 14882

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Library of Congress Catalog Card Number 80-70984

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CHAPTER 9. MINERAL WATER, AND VITAMIN METABOLISM

Minerals, water, and vitamins are essential components of life processes. The role of minerals in the growth and annual cycle of wild ruminants is obvious. Skeletal growth is extremely rapid, and antler growth in those species which grow a new set of antlers each year is truly remarkable. Water is the major component in body tissue. Vitamins have little identifiable mass in the body, but they have major roles in physiological processes.

The importance of mineral, water and vitamin balances is not reflected by the amount of literature available on these subjects. An understanding of basic life processes is such an important foundation for management, yet the literature on these subjects is very sparse. The explanation for this may likely include the tendency for management to be approached from the management end, and the difficulty in studying the interrelated roles of minerals and vitamins in free-ranging animals. Neat experiments are not easily done with minerals and vitamins. Much of the research in such areas is done with small laboratory animals, for several good reasons.

This chapter includes a few basic ideas on the three TOPICS--mineral, water, and vitamin metabolism--and lists of published literature. The requirements for each of these three categories of nutrients could be used to calculate carrying capacity in the same way as energy and protein were used, except that neither the requirements nor the chemical components of forage have been determined precisely enough to make such calculations possible. Think of the nutrient requirement: nutrient supply concept underlying the calculation of carrying capacity, even if precise calculations cannot be made.

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TOPIC 1. MINERAL METABOLISM

Some specific minerals, or inorganic elements, are required for life processes. Other minerals appear to play functional roles in life processes, but their roles are not yet understood. Still others are present in animal tissue, but they may be inert and therefore not essential.

Lists of minerals required, those apparently functional, and minerals generally present in body tissue but without known physiological significance are given below (from Hays and Swenson 1970:663).

Those required, in alphabetical order, are:

potassium
sodium
sulfur
zinc

Those which are apparently functional are:

fluorine molybdenum selenium

Those present but without known physiological significance are:

aluminum	bromine	silicon
arsenic	cadmium	strontium
barium	chromium	

The functions of some of the minerals are well-defined. Calcium and phosphorous, for example, are important in the formation of bones and teeth. Iron is important in hemoglobin synthesis. Sodium and chlorine are important in the maintenance of body fluids. The roles of each of the minerals are discussed in Hays and Swenson (1970: Chapter 33); brief discussions are included here in UNIT 1.1.

Mineral deficiencies and excesses both result in abnormalities in physiological functions. Some deformities are caused by mineral deficiencies in the soil, resulting in inadequate intake by resident animals. Iodine is an example of such a mineral. Mineral excesses occur when too much of a specific mineral is present in the soil, absorbed by the plants, and eaten by primary consumers. Selenium is an example of such a mineral. References describing the effects of mineral deficiencies and excesses will be found in the reference lists in CHAPTER 10.

Wild ruminants usually ingest minerals as part of their daily forage diet. Wild ruminants also ingest soil, and rather deliberately so at times. Some local areas contain soils that are higher than average in mineral content, and these localities, known as mineral licks, become well-used by the resident population. Such mineral licks are not present everywhere, but they are definitely well-used when present. References on mineral licks are listed in UNIT 1.3. The references listed below include books on mineral metabolism, and a list of OTHER PUBLICATIONS that include references not listed on the CODEN lists of serials. They are included here to supplement the limited literature on this TOPIC.

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TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR

edbo	acpr	nyny	879		miner met, vol 1; princip,	comar,cl,ed; bron	1960
edbo	acpr	nyny	1272		miner met, vol 2; elements	comar,cl,ed	1962
edbo	acpr	nyny			mine met, vol 3; calc phys	comar,cl,ed	1969
edbo	cupr	caen	629		symp on hormones & environ	<pre>benson,gk,ed; phi</pre>	1 97 0
edbo	coup	itny	1463 d	oan	duke's physiolog of domest	swenson,mj,ed	1970
edbo	umip	comi	457		trac substs, env, proc 8th	hemphill,dd,ed	1974
edbo	uppr	bama	775 a:	nim	trace elem metab, proc 2nd	hoekstra,wg,ed; /	1974
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UNIT 1.1: MINERAL REQUIREMENTS

The mineral requirements of beef cattle and domestic sheep are discussed in two National Academy of Sciences publications: Nutrient of Fifth Revised Edition. 1975. and Nutrient Requirements Sheep, Beef Cattle, Requirements of Fifth Revised Edition, 1976. These publications include discussions of the requirements and how they are met by feed management, and signs of deficiencies, sometimes with photos of cattle or sheep exhibiting the symptoms.

Mineral deficiencies are much more likely to occur in confined domestic ruminants than in free-ranging wild ones because domestic animals are dependent on the food supplied rather than being able to choose from the variety naturally available. Wild ruminants on severely over-used ranges may tend toward the effects of confinement; options for choosing are gone. Seasonal variations in availability occur also; sodium dynamics in the diets of moose at Isle Royal may limit the moose population (Botkin et al. 1973) with requirements being met by ingestion for only two months of the year.

The mineral requirements of beef cattle and domestic sheep are listed below, based on sheep and beef cattle data in the NRC (1975 and 1976, respectively) publications. Requirements are given as percentage of diet dry matter or mg per kg of dry diet. These values may serve some useful purpose as base-line information for comparison with data on chemical composition of forages given in CHAPTER 11.

MACROMINERAL REQUIREMENTS (percentage of diet dry matter)

Mineral	Sheep Requirements	Growing & Finishing Steers & Heifers	Dry Pregnant Cows	Breeding Bulls and Lactating <u>Cows</u>
Sodium	0 04 - 0.10	0,06	0.06	0.06
Calcium	0.21 - 0.52	0.18 - 1.04	0.18	0.18 - 0.44
Phosphorus	0.16 - 0.37	0.18 - 0.70	0.18	0.18 - 0.39
Magnesium	0.04 - 0.08	0.04 - 0.10	0.04 - 0.10	0.18
Potassium	0.50	0.60 - 0.80	0.60 - 0.80	0.60 - 0.80
Sulfur	0.14 - 0.26	0.10	0.10	0.10

Beef Cattle Requirements

MICROMINERAL REQUIREMENTS (in mg per kg of dry diet)

		Beef Cattl	e Requirements	
Mineral	Sheep Requirements	Growing & Finishing Steers & Heifers	Dry Pregnant Cows	Breeding Bulls and Lactating <u>Cows</u>
Iodine	0.10 - 0.80		0.05 - 0.10	0.05 - 0.10
Iron	30 - 50	10.0	10.0	10.0
Copper	5	4.0	4.0	4.0
Molybdenum	> 0.5			
Cobalt	0.10	0.05 - 0.10	0.05 - 0.10	0.05 - 0.10
Manganese	20 - 40	1.00 - 10.0	20.0	1.00 - 10.0
Zinc	35 - 50	20.0 - 30 0	20.0 - 30.0	20.0 - 30.0
Selenium	0.10	0.10	0.05 - 0.10	0.05 - 0.10

Mineral excesses may occur as well as mineral deficiencies. Possible toxic levels of microminerals are listed below, based on data presented in the NRC (1975 and 1976) publications.

POSSIBLE TOXIC LEVELS OF MICROMINERALS (mg/kg diet)

Mineral	Sheep	Beef Cattle
Taddaa	0.1	100
lodine	87	100
Iron		400
Copper	8 - 25	115
Molybdenum	5 - 20	
Cobalt	100 - 200	10 - 15
Manganese		150
Zinc	1000	90p
Selenium	> 2	5
Fluorine	60 - 200	

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR NAWTA 22--- 119 132 od-- nutrient requirements deer mcewan,lc; frenc/ 1957 NAWTA 22--- 179 188 od-- feed req for growth, maint cowan,imct; wood/ 1957

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR FEPRA 26-51 416 416 odvi uptake, distrib, P32, male whelan, jb; cowan/ 1967 JONUA 108-9 1439 1448 odvi seleni, vit e, bioch, surv brady, ps; brady, / 1978 JWMAA 36--3 996 odvi salt vs brows-season baits mattfeld gf; wil/ 1972 998 JWMAA 37--2 187 194 odvi calcium requi, weaned fawn ullrey, de; youat/ 1973 JWMAA 39--3 590 595 odvi phospho requi weaned fawns ullrey, de; youat/ 1975 JWMAA 40--4 610 odvi sodium deficiencies, adapt weeks, hp, jr; kirk 1976 625 NFGJA 10--2 225 227 odvi adequacy, cobalt nutri, ny smith, se; gardne/ 1963 PAABA 600-- 1 50 odvi nutr req growth, antlr dev french,ce; mcewe/ 1955 PAARA 209-- 1 odvi eff feed restric, antl dev long, ta; cowan, r/ 1959 11 PSEBA 129-3 733 737 odvi calcium strontium age antl cowan, rl; hartso/ 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AZATA 75--- 1 39 ofhr experimental feeding of de nichol,aa 1938 SWNAA 22--1 149 150 odhe bone-chewing behav, desert krausman,pr; biss 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBPAB 60A-3 251 256 ceel energ, nitr metab cold env simpson, am; webs/ 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR JONUA 107-- 1182 1189 alal copper deficiency, alaska flynn,a; franzma/ 1977 JWIDA 12--2 202 207 alal moose milk, hair elem leve franzmann,aw; fly 1976 JWMAA 23--3 356 358 alal minera anal, liver, kidney beeler,da; benso/ 1959 JWMAA 39--2 374 378 alal mineral elem, alaska, hair franzmann,aw; fl/ 1975 PNASA 70-10 2745 2748 alal sodium dynam, north ecosys botkin,db; jorda/ 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARALLKA 57... 2330rata constitution of hair anghi,c1970JOMAA 38--2275277rata disappearance shed antlers mccabe,ra1957

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

anam

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JRACB 37--1 473 481 oram season, trace elemen, livr turkstra, j; hart/ 1977

UNIT 1.2: CALCULATIONS AND PREDICTIONS OF MINERAL REQUIREMENTS

The inclusion of selected minerals in the ratios suggested in the tables of requirements in UNIT 1.1 is a relatively simple matter when formulating feeds for domestic cattle and sheep. Feed of known composition is not provided wild ruminants, however. Estimates of the amount of forage needed to satisfy energy needs may be calculated with the data and procedures found in CHAPTERS 7-8 and 11-12. Calculations of the mineral compositions of these quantities may be made if the chemical data are available.

The daily costs of energy and protein were calculated by evaluating net gains from day to day (see CHAPTERS 7-8), and a similar approach may be taken for minerals, especially the more prominent ones deposited in such conspicuous tissues as antlers. If antler growth rates and chemical compositions are known (see CHAPTER 2), then "before and after" calculations may be made for estimating daily costs of minerals (calcuim and phosphorus, for example) for antler growth, and the results compared to calculated intakes based on forage ingested estimations and forage composition data. This approach provides some means of comparing first approximations of mineral requirements by wild ruminants with empirical data on domestic ruminants.

It is likely that negative mineral balances will be calculated for certain periods of the year, such as the time of rapid antler growth. This calls attention to possible mobilization mechanisms which very likely occur in wild ruminants.

The number of these calculations that can be made is limited by the availability of data on growth rates of the animals or selected tissues and chemical compostion of both animal and forage ingested. Students particularly interested in these processes should set up WORKSHEETS that result in first approximations of mineral requirements.

REFERENCES, UNIT 1.2

CALCULATIONS AND PREDICTIONS OF MINERAL REQUIREMENTS

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odvi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ceel CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR TLPBA 14--1 105 134 alal diet optimizatn, genl herb belovsky,ge 1978 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 55--4 648 655 rata min status, nutrit, season hyvarinen, h; hel/ 1977 MAAIA 51--- 381 419 rata renal respons, prot, sodiu valtonen,m 1979 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS---- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram
CHAPTER 9, WORKSHEET 1.2a

Estimation of mineral intake based on predicted dry-weight forage intake

Mineral intake may be estimated if the mineral components and their fractions of the diet are known, and the amount of forage consumed is known or can be predicted. Calculate the predicted forage intake (See CHAPTER 12) based on energy (CHAPTER 7) and protein (CHAPTER 8) metabolism. Then, determine the intake per day of specific minerals in the forage ingested. The formula is:

SMID = (DWFK)(SMFR)

where SMID = specific minerals ingested per day, DWFK = dry-weight forage ingested in kg per day, and SMFR = specific mineral fractions of the forage.

These calculations are possible only if the mineral composition of the forage species is known (See CHAPTER 11). The mineral component is often given simply as "ash," with no differentiation beween the minerals involved.

Compare the results of these calculations with the requirements given in the tables in UNIT 1.1, and evaluate the literature further to see how these results compare with published values for specific mineral requirements.

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UNIT 1.3: MINERAL LICKS

Mineral licks, also called salt licks, are moist areas or pools with an apparently higher concentration of minerals than usual. These areas are found by animals who then use them regularly.

The mineral lick at Cornell's Arnot Forest that has been used heavily by white-tailed deer for many years is about 10 meters in diameter. It has been walked in, stirred up, ingested, and eroded to the point where the level of the moist soil is up to 20 cm lower than the surrounding forest soils. Trails lead to this lick like spokes in a wheel. The easist way to find it, once in the vicinity, is by following the trails. Another much smaller lick (about 3 m diameter) has apparently been used for a much shorter time.

Deer use of these licks seems to fluctuate seasonally. Heaviest use is in the summer, which may be a response to high rates of mineral metabolism for both antler growth and lactation. Fall and winter use appears to be much less than in the summer, although the deer certainly do visit these licks then.

Maximum desire for salt by elk occurred in late May and early June (Dalke et al. 1965). The use of salt blocks as a management tool was tested, with no effects of salt distribution on the movements between seasonal ranges. If the elk were inclined to use the salt blocks, then local factors such as the vegetation, elevation, number of animals, and the amount of disturbance by livestock affected the use of artificial salt by elk.

Observations of female and yearling caribou on a lick about 23 meters in diameter on the Yukon Arctic Coastal Plain also included trails worn in the tundra vegetation approachig from several directions. Sixty shed antlers were counted in the lick, and animals were observed pawing, chewing, and licking the soil in the lick (Calef and Lortie 1975).

The role of mineral licks in the movement and ecology of wild ruminants is not known, except that the licks do attract the animals, and they do provide minerals that are of physiological importance. The nutritional benefits of mineral licks compared to dispersed minerals in the forage are unknown, although there may be some mineral implications in productivity.

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR CJZOA 53--4 378 384 odhe ceel, activity patterns at carbyn, 1n 1975 JRMGA 32--1 67 71 odhe soil ingestion n centr col arthur, wj, III; al 1979 JWMAA 2---3 79 81 odhe ceel, salt, contrl distrib case, gw 1938

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS---- YEAR

alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 56--1 240 242 rata mineral lick, barren groun calef,gw; lortie, 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 49--5 605 610 oram natural salt licks, ecolog hebert,d; cowan,i 1971 JWMAA 42--3 591 597 oram behav in reln highwa, mont singer,fj 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 30--4 379 387 wiru natural game licks, canada cowan,im; brink,v 1949 NAWTA 18--- 247 258 wiru chem charac, natural licks stackstad,ds; mo/ 1953

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS AUTHORS Y	YEAR
BTROA	114	311	313		elemtl compo of lick, peru emmons, lh; stark, l	1979
JZ00A	158-3	293	310		chem proprties lick, eleph weir, js	1969
00 KHA	3	10	11		artificial salt licks kotov,v 1	1964
ZEJAA	143	107	118	many	comp intk slt lcks, eur sp ueckermann,e	1968

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TOPIC 2. WATER METABOLISM

Water is not only an important but the major component of the body of ruminant animals. It is not the water as a compound composed of hydrogen and oxygen that is of significance, however, but rather its properties that allow the many physical-chemical processes to occur.

The water content of fetuses is very high--90% or more--and declines with advancing gestation. Adult cattle, however, still have water contents of 40 to 70% or more, and white-tailed deer have water contents of 60% of ingesta-free adult body weight (Robbins et al. 1974).

Body composition data given in CHAPTER 2 show that water and fat fractions are inversely proportional. This is because the water content of fat is only about 10% or less. Thus fat animals, such as wild ruminants in the fall at maximum weights during the annual cycle, have a lower water fraction than lean ones.

Water molecules can rapidly penetrate most cell membranes (Houpt 1970:745). If a pressure gradient exists, then water molecules are expected to move from the higher pressure to the lower pressure. The same should occur if there are differences in osmotic concentrations; water should move from the lower concentration to the higher one. Since water molecules are often more likely to penetrate through membranes than solute molecules, concentration differences may be equalized by the movement of water.

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WATER METABOLISM

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR

edbo	nyha	nyny 255	symp on salt & water metab fishman,ap,ed	1960
edbo	else	nyny 251	wat, elctrolyt met, 2nd ed de graeff, j, ed; 1	1964
edbo	macm	nyny 570	symp, thirst, reg body wat wayner,mj.ed	1964
aubo	moco	salo 169	wat, elect met, acd-ba bal muntwyler,e	1968
edbo	base	nyny 260	n, electro, wat & ener met rechcigl,m,jr ed	1970

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- Johnson, C. E. 1940. Waterholes for wildlife. National Park Service, Region Three Quart. 2(2):9-11.
- Smith, A. D. 1954. How much water does a deer drink? Utah Fish & Game Bulletin 10(9):1,8.
- Talbot, M. W. 1926. Range watering places in the southwest. U. S. Department of Agriculture Department Bulletin No. 1358. pp. 1-43.

b.

UNIT 2.1: WATER COMPARTMENTS

Body water may be divided into two compartments: intracellular and extracellular. These terms refer to the fluid inside the cells and outside the cells, respectively. Extracellular fluid is further divided into interstitial fluid and plasma (Houpt 1970:744). The plasma is part of the vascular system, of course, and interstitial fluid is within tissues but not in cells.

LITERATURE CITED

Houpt, T. R. 1970. Water, electrolytes, and acid-base balance. pp. 743-766 In M. J. Swenson, Ed. Dukes' physiology of domestic animals. 8th Ed. Cornell University Press, Ithaca, N. Y. 1463 pp.

REFERENCES, UNIT 2.1

WATER COMPARTMENTS

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odvi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ceel

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 50--1 107 116 rata chang body water extracell cameron, rd; luick 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 34--4 908 912 anam energ flux, water kinetics wesley, de; knox,/ 1970 XIBPA 1.... 250 250 anam water kinetics in pronghor wesley, de; knox,/ 1971 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

PNUSA 16--2 108 112 functions of water in body robinson, jr 1957 SZSLA 31--- 261 296 mamm comprtv watr & energ econo macfarlane, wv; ho 1972

UNIT 2.2: REQUIREMENTS AND TURNOVER RATES

The water balance is obviously an important part of the physiology of wild ruminants since water is such a major component of animal tissue. Water consumption is very difficult to determine, however, because the sources of water include forage (often 50% or more water), dew on the vegetation (sometimes twice a day), snow (often present in large quantities), and open water. A unique experimental set-up is described by Elder (1954) who was able to measure the water consumption of individuals and groups of mule deer that drank from tubs and barrels in which water levels could be measured. Six to seven quarts were consumed in an average of three minutes of drinking time.

Wild ruminants are not as dependent on open water as domestic ones are. Many species do not seem to frequent open water at all, except perhaps in the heat of summer when water losses are high. The vegetation and snow seem to be adequate sources under most natural conditions. Nevertheless, white-tailed deer, a species that can get along well in a frozen habitat, will use open water when available. There are reports in published literature of a lack of interest in open water (see Hosley 1956), yet whitetails do use open water in otherwise frozen habitat. I have observed large amounts of deer activity in a seeepage area below an earth dam in west-central Minnesota, and in natural seepage areas at Cornell's Arnot Forest in New York State.

It is difficult to separate requirements from preferences. Water requirements are met in several different ways; preferences for meeting those water requirements vary through the year and from place to place. The water consumption of pronghorn varied inversely with the quantity and succulence of the preferred forage species (Beale and Smith 1970). Succulent spring vegetation has a high water content and excess water is excreted not only in urine but also in feces.

References on water consumption follow. The WORKSHEET is included which relates to forage intake calculations and the water content of the forage.

LITERATURE CITED

- Beale, D. M. and A. D. Smith. 1970. Forage use, water consumption, and productivity of pronghorn antelope in western Utah. J. Wildl. Manage. 34(3):570-582.
- Elder, J. B. 1957. Notes on summer water consumption by desert mule deer. J. Wildl. Manage. 18(4):540-541.
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REFERENCES, UNIT 2.2

REQUIREMENTS AND TURNOVER RATES

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 39--2 355 360 odvi milk consumpti weight gain robbins,ct; moen, 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJVRA 31--4 673 677 odhe dosh, tot body watr turnov longhurst,wm; ba/ 1970 JWMAA 18--4 540 541 odhe summer wate consumpt deser elder,jb 1951 JWMAA 33--2 389 393 odhe water turnover in mule dee knox,kl; nagy,jg/ 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ceel

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 54--6 857 862 rata tritium wat dilu, wat flux cameron,rd; whit/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 34--3 570 582 anam forag, watr consum, produc beale,dm; smith,a 1970 UTSCB 29--1 3 6 anam seasonal forage use, utah beale,dm; scotter 1968 anam continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR WGFBA 12--- 1 61 anam food hab, abundan, distrib sundstrom,c; hep/ 1973 XIBPA 1.... 250 250 anam water kinetics in pronghor wesley,de; knox,/ 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CAFGA 54--4 289 296 ovca summer water requirements blong,b; pollard, 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJAEA 19--4 655 672 doca eff wat restrc & exer, dig thornton, rf; yate 1968

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORI)S			AUTHORS-			YEAR
JANSA	356	1271	1274	dosh	eff	wat	restric	nutrnt	dig	asplund,	im:	pfand	1972

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORL)S				AUTHORS	-	YEAR
PNUSA	162	108	112		func	tior	ns of	wa	ter i	In body	robinson, jr		1957
SZSLA	31	261	296	mamm	comp	rtv	watr	&	energ	g econo	macfarlane,wv; h	0	1972
YAXAA	1955-	14	18		anim	als	and	fow	1 and	l water	sykes,jf		1955

}

CHAPTER 9, WORKSHEET 2.2a

Water consumption as part of forage ingested

The amount of water ingested with the forage may be determined if the amounts of forage ingested are determined based on energy or protein requirements as discussed in CHAPTER 12. The calculations of forage ingested are made on a dry weight basis, so the water fraction of field-weight forage must be known.

Determine the water fraction of field-weight forage, calculate dry-weight forage required to meet energy and protein requirements, and estimate the water component of the diet. Compare this to the limited data on water consumption.

TOPIC 3. VITAMIN METABOLISM

Vitamins are organic compounds that function as metabolic catalysts, usually in the form of coenzymes (Smith 1970:634). Vitamins may be divided into a fat-soluble group (A, D, E, and K) and a water-soluble group (the B complex and C).

Vitamins have essential roles in metabolic processes. Vitamin requirements are met both by ingesting foods that contain the vitamins, and by synthesis in the body. Rumen bacteria synthesize vitamins in the Bcomplex and vitamin K, so ruminant animals are not dependent on a dietary source for these vitamins. Neonates, however, do not have a functional rumen, so they are dependent on dietary vitamins in the first few weeks of life. Solar radiation, especially the ultraviolet wavelengths, is an important factor affecting the amount of vitamin D in the body. Sunlight is not the "source," but the stimulus for the formation of vitamin D at the skin surface to be absorbed by the body.

A deficiency of vitamins is called avitaminoses. An excess is called hypervitaminoses. Vitamin imbalances cause malfunctions in different body functions, especially metabolic pathways, as they function as metabolic catalysts. The vitamin metabolism of wild ruminants is largely unknown, except a inferred from studies on domestic ruminants. A few measurements of vitamin levels in the tissue of Odocoileus have been made, but that is the extent of published literature found for wild ruminants.

Vitamin requirements of beef cattle are listed below, from a table in the NRC (1976) publication.

VITAMIN REQUIREMENTS OF BEEF CATTLE (in amount per kg of dry diet; IU)

Beef Cattle Requirements

Vitamin	Sheep Requirements	Growing & Finishing Steers & Heifers	Dry Pregnant Cows	Breeding Bulls and Lactating <u>Cows</u>
Α	IU	2,200	2,800	3,900
D	IU	275	275	275
E	IU	15-60	<u> </u>	15-60

This TOPIC 3 is not divided into UNITS because of the lack of information, and there are no worksheets. The paucity of information may stimulate someone who specializes in vitamins to review the pertinent literature on wild and domestic animals, and to write a good review of the concepts and principles involved in vitamin nutrition and metabolism.

LITERATURE CITED

- National Research Council. 1976. Nutrient requirements of beef cattle. Fifth Revised Edition. National Academy of Sciences. Washington, D. C. 56 pp.
- Smith, S. E. 1970. Vitamins. Pages 634-659 In M. J. Swenson, Ed. Dukes' physiology of domestic animals. 8th Ed. Cornell Univ. Press, Ithaca, NY 1463 pp.

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VITAMIN METABOLISM

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR

edbo	nvif	nyny	118		symp on v	vitami	n meta	bolism	brow	ı,gm,ed;	1	1956
aubo	acpr	nyny	366		biosyn vi	itami	& rel	compds	goodv	vin,tw		1963
aubo	hein	loen	148		vitamins	in en	docrin	e meta	jenni	lngs,iw		1970
aubo	vare	nyny	278		hndbk of	vitam	ins &	hormon	kutsl	cy,rj		1973
edbo	nasc	wadc	72	dosh :	nutrient	requi	rem of	sheep	nat1	research	ı cou	1 97 5
edbo	nasc	wadc	56	doca	nutrient	req o	f beef	catt1	natl	research	i cou	1976

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS AUTHORS	YEAR
JAPRA	93	130	131	odvi	avitaminosis e, white-tail indianapolis zoo	1968
JOMAA JOMAA	364 413	553 410	557 411	odvi odvi	vitamin excretion by deer teeri,ad; pomera/ vitamins a and e in blood haugen,ao; hove,e	1955 1960
JONUA	108-9	1439	1448	odvi	<pre>seleni, vit e, bioch, surv brady,ps; brady,/</pre>	1978
JWMAA	401	172	173	odvi	vitam a concentr in livers youatt,wg; ullre/	1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBPAB 41B-4 745 758 odhe carote, vit a, liver, seru anderson, ae; med/ 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ceel

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARAASNA 19--- 2228rumi aspects of vit e deficienc oksanen, he1973JAVMA 151-4430436rumi vit a nutition of ruminant mitchell, ge1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AASNA 19--- 87 96 chemistry of vitamin e hjarde,w; leerbe/ 1973

CLOSING COMMENTS

This chapter has included a few basic ideas and lists of references in the areas of mineral, water, and vitamin metabolism. The amount of material in the chapter is limited by both the lack of published information on these nutrients and my own limited knowledge of the roles of these nutrients. This chapter is a good example of an opportunity for someone specializing in these nutrients to particiapte as an author in future revisions.

> Aaron N. Moen March 12, 1981

GLOSSARY OF SYMBOLS USED - CHAPTER NINE

DWFK = Dry-weight forage ingested in kilograms per day

SMFR = Specific mineral fraction of the forage
SMID = Specific minerals ingested per day

GLOSSARY OF CODE NAMES - CHAPTER NINE

CODEN

AASNA Acta Agriculturae Scandinavica Supplementum AJAEA Australian Journal of Agricultural Research AJVRA American Journal of Veterinary Research ALLKA Allattani Kozlemenyek (Hungary) AMNAA American Midland Naturalist AZATA Arizona Agricultural Experiment Station Technical Bulletin BTROA Biotropica CAFGA California Fish and Game CBPAB Comparative Biochemistry and Physiology A Comparative Physiology CJZOA Canadian Journal of Zoology CORTB Clinical Orthopaedics and Related Research FEPRA Federation Proceedings JANSA Journal of Animal Science JAPRA Journal of Small Animal Practice JAVMA Journal of the American Veterinary Medical Association JOMAA Journal of Mammalogy JONUA Journal of Nutrition JRACB Journal of Radioanalytical Chemistry JRMGA Journal of Range Management JWIDA Journal of Wildlife Diseases JWMAA Journal of Wildlife Management JZOOA Journal of Zoology MAAIA Journal of the Scientific Agricultural Society of Finalnd NAWTA North American Wildlife and Natural Resources Conference, Transactions of the, NFGJA New York Fish and Game Journal OOKHA Okhota i Okhotnich'e Khozyaistvo PAABA Pennsylvania Agricultural Experiment Station Bulletin PAARA Pennsylvania State University College of Agriculture Agricultural Experiment Station Progress Report PNASA Proceedings of the National Academy of Sciences of the United States PNUSA Proceedings of the Nutrition Society PSEBA Proceedings of the Society for Experimental Biology and Medicine SWNAA Southwestern Naturalist SZSLA Symposia of the Zoological Society of London TLPBA Theoretical Population Biology

- UTSCB Utah Science
- WGFBA Wyoming Game and Fish Commission Bulletin
- XIBPA US-IBP (International Biological Program) Analysis of Ecosystems Program Interbiome Abstracts

YAXAA USDA Yearbook of Agriculture

ZEJAA Zeitschrift fuer Jagdwissenschaft

LIST OF PUBLISHERS - CHAPTER NINE

acpr	Academic Press	New York	nyny
base	Basel	New York	nyny
coup cupr	Cornell University Press Cambridge Univ. Press	Ithaca, NY Cambridge, England	itny caen
dohr	Dowden, Hutchinson, & Ross	Stroudsburg, PA	stpa
e1se	Elsevier	New York	nyny
hein	Heinemann	London, England	10en
macm moco	MacMillan Company C. V. Mosley Company	New York St. Louis, MO	nyny salo
nasc nvif nyha	National Academy of Science National Vitamin Foundation New York Heart Association	Washington, D. C. New York New York	wadc nyny nyny
umip uppr	Univ. of Missouri Press University Park Press	Columbia, MO Baltimore, MD	comi bama
vare	Van Nostrand - Reinhold	New York	nyny

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4		004	035	063	094	124	155	185	216	247	277	308	338	4
5		005	036	064	095	125	156	186	217	248	278	309	339	5
6		006	037	065	096	126	157	187	218	249	279	310	340	6
7		007	038	066	097	127	158	188	219	250	280	311	341	7
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10		010	041	06 9	100	130	161	191	222	253	283	314	344	10
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THE BIOLOGY AND MANAGEMENT OF WILD RUMINANTS

CHAPTER TEN

FACTORS AFFECTING THE PHYSIOLOGY AND METABOLISM OF WILD RUMINANTS

bу

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CornerBrook Press Box 106 Lansing, N.Y. 14882

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Library of Congress Catalog Number 80-70984

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CHAPTER 10. FACTORS AFFECTING THE PHYSIOLOGY AND METABOLISM

OF WILD RUMINANTS

The physiology and metabolism of wild ruminants has been discussed in CHAPTER 6 - 9, with discussions of measured results and equations that permit calculations of rates and quantities. The discussions have centered about presumably normal conditions and functions. Rates of energy metabolism for example, have been discussed in CHAPTER 7. The general equation for calculating ecological metabolism per day as a function of weight, reproductive rate, and JDAY gives an expected metabolism, unencumbered by unusual circumstances. The effects of cold weather, for example, are built into the equation as part of the winter metabolic depression

There are a number of factors that affect the physiology and metabolism of wild ruminants, causing deviations from the norm. It is these deviations that cause much of the drama in ecology as the struggle for survival proceeds from day to day. Many, if not most of these factors are ordinary components of daily existence, with the effects becoming pronounced only when unusually high levels of the effector are encountered, or the effects of several factors are combined and the accumulated effect is great.

Parasites, for example, are part of the daily life of all wild ruminants. Unusually large numbers of parasites, however, may have a pronounced detrimental effect on the productivity of the host. Lesser parasite loads may have an impact when other factors, such as deteriorating range conditions during an exceptionally long winter, exist and act together to depress overall ecological efficiency.

This chapter includes extensive reference lists in three TOPICS. These three topics and the units in each are not discussed at length, however, because ecological analyses--the evaluation of relationships in the ecological context between primary consumer and range, parasite and host--have not been made. The extensive and organized reference lists may increase the efficiency of literature reviews to the point where persons interested in these factors will be more ecologically comprehensive in the approach to understanding the roles of these factors.

TOPIC 1. PARASITES, DISEASES, AND ABNORMALITIES

Parasites, diseases and abnormalities are an extensive array of factors that affect the physiology and metabolism of wild ruminants. They are also a normal part of life; parasites and diseases are expected to be present, and the responses necessary to cope with their effects serve to strengthen the species' ability to withstand future impacts. It must be assumed that a natural balance exists between parasite and host, with short-term fluctuations in favor of one or the other.

The impacts of parasites and diseases may be greatest when a ruminant species is moved into suitable but unoccupied areas. Then, the additional stresses due to the moving plus the possibility of encounters with new parasites and disease may make the transplants more susceptible. Generations may build up a resistance to the effects, resulting in increased productivity in later years.

It is conceivable that a species could be moved into a new area and experience less environmental resistance, including lesser effects of parasites and diseases, than in the previous range. Initial productivity may be higher than normal in the previous range for a time. An initially high rate of productivity may be followed by depressed productivity as factors affecting physiology and metabolism begin to take effect.

The effects of parasites and diseases tend to increase and decrease through the year and over the years. Abnormalities are more of an either-or effector; either an animal is abnormal or it isn't. The degree to which the abnormality affects physiological functions may vary, of course, resulting in a gradient-type effect on the animals productivity.

There are rich opportunities for exploring the effects of parasites, diseases, and abnormalities on ecological productivity, but the rich opportunities are also very challenging. The effects are hard to measure because productivity is hard to measure. The results of different levels of effects, providing they can be quantified, are often not available for two or more years when working with large animals such as wild ruminants because of the time needed to reach reproductive maturity. Nevertheless, the opportunities exist, and it is my hope that the organized reference lists in the units that follow will make study and work in these areas not only more efficient but also more appealing.

REFERENCES, TOPIC 1

PARASITES, DISEASES, AND ABNORMALITIES

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR faog roit 119 anim diseases free-liv wild ani mcdiarmid, a 1962 aubo 1963 ccth spil 967 anim diseases trans anim to man hull, tg, ed edbo aubo hein loen 136 ecology of parasites 1966 croll, na veterinary clinical pathol coles, eh aubo wbsc phpa 462 1980 acpr nyny 332 anim diseases free-liv wild ani mcdiarmid, a, ed edbo 1969 aubo isup amia 421 mamm infectious dise, wild mamm davis, jw; karsta/ 1970 edbo isup amia 364 mamm parasitic disea, wild mamm davis, jw; anders/ 1971 aubo lefe phpa 1521 anim veterinary pathology,4th e smith,ha; jones, 1972 edbo plpr nyny 686 many wildlife diseases page, la 1976

OTHER PUBLICATIONS

- Fyvie, A. 1969. Manual of Common Parasites, Diseases and Anomalies of Wildlife in Ontario, 2nd ed. Dept. of Lands and Forests, Ontario. 102 p.
- McDiarmid, A. 1960. Diseases of Free-Living Wild Animals. Animal Health Monograph #1, FAO Working Document. FAO, Rome. 91 p.
- Reed, D. and H. Shane. 1976. Deer Diseases in South Dakota, 1972-76. Department of Game, Fish and Parks, Pierre, S. D. 32 p. [parasites and diseases, odvi, odhe]
- Worley, D. E. and K. R. Greer. 1976. Parasites and Diseases of North American Elk (Cervus spp.) - An annotated bibliography. 52 p. Montana Dept. of Fish and Game in Cooperation with Montana Agricultural Experiment Station, Montana State Univ., Bozeman, Montana.

UNIT 1.1: PARASITES

Parasites inhabit wild ruminants, living off the ruminant host as the ruminant lives off the range. The effects may not be limited to resource use, however (anemia, for example, in the case of a blood-sucking parasite) but may also include effects on behavior as well as physiology. "Moose sickness" is such a case; moose infected with <u>Pneumostrongylus tenuis</u> exhibit aberrant behavior as a result of the effects of the parasite in the central nervous system. It is interesting to note that this parasite may be resident in white-tailed deer without the effects observed in moose. The prevalence of white-tailed deer carrying the parasite is great enough to cause problems in moose populations when their habitats overlap.
The larval stages of parasites often do more harm than the adult stages. Further, the different stages in the life cycles of different parasites result in different levels of susceptibility under different habitat conditions. Range conditions, buth vegetative and physical, may be very important factors affecting levels of parasite infections. This is wellknown in the treatment of domestic livestock; the development of clinical parasitism depends not only upon the number and activity of the parasites, but also on the age, resistance, and nutritional status of the host, climatic conditions and management practices (Siegmund 1967: 699-700). Putting it another way, range conditions, weather conditions, and the age, resistance and nutritional status of wild ruminants affect the number and the effects of parasites on their hosts.

One example of differences in the abilities of individual animals to withstand parasites and in the ecological effects of different parasites on host populations is given by researchers on domestic sheep. Georgi and Whitlock (1967) established a direct cause and effect relationship between exposure of sheep to infection by Haemonchus contortus (twisted stomach worm), a helminth parasite of ruminants usually found in the abomasum, and The number of erythrocytes is important in the onset of erythrocyte loss. determining the efficiency of the oxygen transport system, which is fundamental to efficient tissue metabolism. Georgi (1964) clearly shows that the rate of iron loss from sheep infected with H. contortus was greater than that from non-infected sheep. Evans and Whitlock (1964) concluded that, other things being equal, an animal (sheep) with a low erythrocyte volume has a smaller chance of surviving a natural challenge with this parasite than an animal having a greater erythrocyte volume. Such experimental evidence in sheep indicates that responses to parasites by the host are important determinants of metabolic efficiencies.

Basic research on the reactions of wild ruminants to pathogens and parasites should be made along with studies on apparently healthy animals in frder to evaluate the ecological importance of these interactions. Such research is especially difficult on wild ruminants because the results of such research are not necessarily important at clinical levels established for domestic animals, but could be rather important, in relation to ecological metabolism and its relation to productivity. These relationships are not only hard to evaluate; they are hard to identify. Van Volkenburg and Nicholson (1943) found 11 specific parasites in the nasal passages, skin, abomasum, and small intestines of dead deer in the Edwards Plateau, Texas. They concluded that parasites were unimportant among deer on ranges with sufficient food but made no efforts to quantify metabolic relationships between host and parasite.

The next several pages contain lists of references with information on parasites in relation to the different species of wild ruminants. The lists should prove to be convenient when reviewing this area of the biology of wild ruminants.

LITERATURE CITED

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REFERENCES, UNIT 1.1

PARASITES

SERIALS

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY W	ORDS						AUTHORS			YEAR
APRPA APRPA APRPA	13-44 141 21-34	445 1 475	481 13 479	cerv cerv cerv	helmi helmi trich	nths Inth Nostr	, fau ne	he ina emat	lmin in ode,	thi po mo	lases oland ongol	drozdz, drozdz, drodz,j	j j ; na	mzyl,g	1965 1966 1973
CJZOA	445	851	861	cerv	exper	stu	d,	pne	umos	tro	ongy1	anderso	n,rc	; mur/	1966
JOPAA JOPAA JOPAA	131 392 593	25 119 569	28 134 578	cerv cerv cerv	nemat fluke haema	od, in phys	sto ame ali	eric s m	h, c a ru ages	erv min h,	vidae nants japn	schwart price,e saito,y	z,b w ; ho	ogstra	1926 1953 1973
NAWTA	30	156	167	cerv	cereb	rosp	ina	l n	emat	odi	lasis	anderso	n,rc		1965
PPPAA	64	439	458	cerv	ecolo	ogy o	fd	lerm	acen	tor	alb:	howell,	de		1939
WDABB	84	304	310	cerv	eco1	rela	tio	n, 1	meni	ng	worm	anderso	n,rc		1972
ZOANA	110	202	208	cerv	nemat	odes	, n	am	er r	umi	nant	wehr,ee	; di	kmans/	1935
CODEN	VONU	BEPA	ENPA	ANIM	KEY W	ORDS						AUTHORS			YEAR
AZATA	75	1	39	od	exper	imen	t f	eed	ing	of	deer	nichol,	aa		1938
AJVRA AJVRA	23 231	199 21	201 23	od od	new r doca,	ecor tra	d, nsm	nema	atod apla	, (.s,	alif tick	annerea osebold	ux,r ,jw;	f doug/	1941 1962
				od	conti	nued	on	th	e ne	xt	page				

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
CAFGA CAFGA CAFGA CAFGA CAFGA CAFGA CAFGA	154 301 314 321 324 331 513 554	309 58 201 17 182 54 208 307	315 59 208 18 189 54 210 316	od od od od od od od	parasitism in deer eye worm infection, califo some worm parasites, calif the nose bot fly of deer foot worm parasite, califo deer foot worm infect, cal tick paralysis, california parelaphostrongylus, calif	<pre>vanroekel,h herman,cm herman,cm herman,cm; bischo herman,cm brunetti,oa brunetti,oa</pre>	1929 1944 1945 1946 1946 1947 1965 1969
CNVJA	33	71	78	od	cul, ser evi, lepto, ontar	abdulla,pk; kars/	1961
COVEA	403	3 15	323	od	experim transmis, lungworm	o'roke,ec; cheatu	19 50
FPARA	201	41 .	48	od	yearly cycle, helminthoses	erhardova-kotrla/	1973
CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
JFUSA	408	540	543	od	foot-wor dis, n rocky mt r	denio,rm; west,rm	1942
JOPAA JOPAA JOPAA	193 291 554	246 80 720	246 80 725	od od od	onchocerca, subcut abscess corynebacterium, louse fly dist, meningi worm, se u s	dikmans,g steinhaus,ea prestwood,ak; smi	1933 1943 1969
JWIDA JWIDA JWIDA	82 82 103	109 163 225	111 164 227	od od od	devic, aid exam brain, men devic, aid exam sm intesti trans echinoc gran, coyote	smith,jf doster,gl romano,mn; brune/	1972 1972 1974
JWMAA JWMAA JWMAA	72 301 313	220 202 595	223 203 597	od od od	parasitism, malnutrit, tex deer, cattle fever ticks endoparasites, welder refu	<pre>van volkenberg,h/ park,rl; skov,o;/ glazener,wc; know</pre>	1943 1966 1967
NAWTA NAWTA NAWTA	3 10 16	882 242 135	885 246 143	od od od	deer, fever tick erad, flo dee, domes livest par, dis effec, screw-worm, southea	shillinger,je herman,cm allen,gw	1938 1945 1951
NYCOA	83	32	32	od	deer fly, mechan dee repel	lee,j	1953
PARAA	36	199	208	od	nematod lungworms, no amer	dougherty,ec	1945
PCGFA	8	1	19	od	screw-worm problem, florid	strode,dd	1954
PMASA	18	33	33	od	ecto, endoparasites, monta	senger,cm	1958
PPASA	392	73	77	od	id nemato eggs, larv, penn	samuel,wm; beaudo	1965
WDABB WDABB	14 51	44 27	48 30	od od	microfilar, wehrdikm, ariz elaeophorosis, new mexico	hibler,cp hibler,cp; adcoc/	1965 1969
WIDIA	10	1	5	od	the foot worm, northe utah	yuill,tm; low,jb/	1961

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AESAA 62--5 1208 1210 odvi pharyngeal bot in oklahoma hair, ja; howell,/ 1969 883 odvi antigen, serodiag, theiler gadir, fa; hidalg/ 1970 AJVRA 31--5 879 AJVRA 39-12 1901 1903 odvi anthelmint activ, albendaz foreyt,wj; drawe, 1978 AMNAA 79--2 528 530 odvi rate, tick attachmen, fawn montgomery,gg 1968 AMNAA 85--2 507 513 odvi seas fluc tricholipeur, tex samuel, wm; traine 1971 AMNAA 86--2 506 508 odvi new sp. trichostrongylus maples, wp; englan 1971 APRPA 19-11 141 145 odvi 2 nematode species, poland drozdz, j 1971 CJCMA 33--4 280 286 odvi eff pneumostrongyl on kids anderson, rc; stre 1969 CJCMA 38--3 271 279 odvi path chang, fasciola infec presidente, pja; / 1974 CJCMA 39--2 155 odvi exp fasciola-hepati infect presidente, pja; / 1975 165 CJCMA 39--2 166 177 odvi fasciola infec, path featu presidente, pja; / 1975 CJZOA 34--3 167 173 odvi elaphostrongyl, cranial ca anderson, rc 1956 odvi inci, dev, tr, pneumostron anderson, rc CJZOA 41--5 755 792 1963 CJZOA 45--3 285 289 odvi penetrat, pneumostrongylus anderson, rc; stre 1967 CJZOA 46--3 373 383 odvi gastropods, pneumostrogylu lankester, mw; and 1968 CJZOA 47--6 1411 1411 odvi host recor, nematodirus sp webster,wa; macka 1969 CJZOA 52--6 785 odvi demodex odocoilei, new spp desch,ce; nutting 1974 789 CJZOA 53--1 87 odvi seas cha, aboma worms, ont baker, mr; anderso 1975 96 CJZOA 53--8 1047 1054 odvi seas change, louse populat watson, tg; anders 1975 357 odvi antibodies, theileria infe kutler,kl; robin/ 1967 CNJMA 31-12 354 CNVJA 5--11 287 296 odvi elaphostrongylosis, moose, smith, hj; archib/ 1964 COVEA 40--2 211 212 odvi new host, dicrocoelium, ny mapes, cr; baker, d 1950 COVEA 51--3 431 odvi cerebra nematodiasis, penn alibasoglu,m; kr/ 1961 441 COVEA 68--2 139 149 odvi demodex canis, redescripti nutting, wb; desch 1978 DABBB 34B-- 5002 5002 odvi lone star tick, fawn survi hoch,al 1974 DABBB 35B-6 3114 3114 odvi survey helminth parasites owen, wb, jr 1974 DABBB 36B-8 3806 3807 odvi epizoot larg amer liver fl foreyt.wj 1976 EVETB 6---2 263 269 odvi seas abund, lonestar ticks patrick, cd; hair, 1977 GRLEA 8---1 1 29 odvi the deer flies of indiana burton, jjs 1975 JAVMA 96--- 607 608 odvi tropic var cattle-fev tick knapp, jv 1940 JAVMA 163-6 556 561 odvi abomas helmin, tex, virg is prestwood, ak; ha/ 1973 JAVMA 166-8 787 odvi helmin w-t deer, cattle, s prestwood, ak; ke/ 1975 789 JAVMA 169-9 896 900 odvi intest nema, w-t deer, s e pursglove, sr; pr/ 1976 JAVMA 171-9 933 935 odvi prev, distr, setaria se us prestwood, ak; pur 1977 JAVMA 171-9 936 938 odvi fascioloides infect, se us purseglove, sr; p/ 1977 JAVMA 173-9 1242 1243 odvi otitis media caused by pso rollor, ea: nettl/ 1978

odvi continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JMENA 7---5 567 574 odvi amblyomma, s texas, theile samuel, wm; traine 1970 JMENA 8---5 495 498 odvi arthropod parasites, w-t d kellogg, fe; kist/ 1971 JMENA 9---1 104 106 odvi lipoptena mazamae dip, tex samuel,wm; traine 1972 220 JNYEA 85--4 217 odvi epizo, bovin babesias, tex hourrigan, jl 1977 JOMAA 16--1 70 71 odvi onchocerciasis, montana rush,wm 1935 JOPAA 29--2 158 158 odvi protostrongylus, lungs, ny goble,fc 1943 JOPAA 45--4 32 32 odvi odhe louse-flies, w montan senger, cm; capell 1959 JOPAA 53--4 691 691 odvi oesophagostomum, se u s payne,rl; maples/ 1967 JOPAA 54--3 441 444 odvi ostertagia, med stom worms becklund, ww: walk 1968 JOPAA 54--6 1091 1091 odvi trichostrongylus axei samuel, wm; traine 1968 JOPAA 55--4 720 725 odvi distr meninge worm, se usa prestwood, ak: smi 1969 JOPAA 56--1 123 127 odvi geog dist, gongylonem, par prestwood, ak; sm/ 1970 JOPAA 56--4 6 7 odvi ecol menigea worm, cervids anderson, rc 1970 300 JOPAA 56--4 299 odvi parasites, relat epizoo fac samuel, wm 1970 JOPAA 56--5 1038 1038 odvi new host for psoroptes sp strickland, rk; g/ 1970 JOPAA 57--3 468 493 odvi spiculosteragia nematoda doster,gl; friend 1971 odvi fail boophil, trans anapla kuttler,kl; grah/ 1971 JOPAA 57--3 657 659 JOPAA 57--6 1292 1292 odvi cestodes of w-t de, se usa prestwood,ak 1971 JOPAA 58--5 897 902 odvi parelaphostrongylus, new sp prestwood, ak 1972 JOPAA 58--5 1010 1011 odvi occuren, fascioloides, tex foreyt,wj; todd, 1972 JOPAA 59--1 208 209 odvi oxyclozanide, fascioloides forety,wj; todd,a 1973 JOPAA 59--6 1091 1098 odvi lone star tick, hemat chan barker, rw; hoch,/ 1973 JOPAA 60--4 573 573 odvi leptostrongyl, georgia, lou prestwood, ak; pur 1974 JOPAA 60--6 1059 1060 odvi trichostrongylus, ostertag pursglove, sr, jr;/ 1974 JOPAA 61--1 145 146 odvi trypanosomes, new york krinsky,wl 1975 JOPAA 62--1 26 32 odvi devel large amer liv fluke foreyt,wj; todd,a 1976 JOPAA 62--1 144 145 odvi parenteral infec, fasciola foreyt,wj; todd,a 1976 JOPAA 62--1 166 168 odvi oesophagostomum in w-t dee baker,mr; pursglo 1976 JOPAA 62--4 574 578 odvi eucyathostomum webbi, new s pursglov, sr, jr 1976 JOPAA 62--5 824 825 odvi trypanosomes from oklahoma krinsky, wl; burgd 1976 JOPAA 62--6 1000 1002 odvi tech det amblyomma infecti durham, ka; corst/ 1976 JOPAA 63--5 938 odvi mamm hosts fasciola, washi lang,bz 939 1977 JOPAA 63--6 974 978 odvi low lev infec parelaphostr prestwood, ak; net 1977 JOPAA 63--6 1050 1052 odvi fascioloides magna, w-t de foreyt,wj; samue/ 1977 JOPAA 64--1 14 16 odvi 2 sp of entamoeba, georgia kingston,n: stabl 1978 JOPAA 64--3 567 568 odvi filariasis, w-t d, missour robbins,dj; clark 1978 JPROA 8---1 33 41 odvi ophryoscolecid fauna, w-t zielyk,mw 1961 JPROA 18--2 306 308 odvi ecol factors influ eimeria samuel, wm; traine 1971 42 JWIDA 6---1 35 odvi experimental haemonchosis foreyt,w; trainer 1970 JWIDA 6---1 56 63 odvi compar, parasit, 2 populat beaudoin,r1; sam/ 1970 JWIDA 6---1 84 86 odvi neurol dis, meningeal worm prestwood, ak 1970 JWIDA 6---3 182 183 odvi pulex porcinus, s texas samuel, wm; traine 1970 JWIDA 6---4 430 436 odvi mening worm invas of brain eckroade, rj; rhe/ 1970

odvi continued on the next page

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS	AUTHORS	YEAR
JWIDA	64	437	440	odvi	hosts, cattle fever ticks	kistner, tp; hayes	1970
JWIDA	72	133	134	odvi	natural occur haemonchosis	prestwood,ak: kel	1971
JWIDA	73	142	146	odvi	pharyngeal bot fly larvae	samuel,wm;train/	1971
JWIDA	73	149	154	odvi	lungworms in w-t d, se usa	prestwood, ak sm/	1971
JWIDA	81	63	6 6	odvi	fail establ babesia infect	kuttler.kl; grah/	1972
JWIDA	82	109	111	odvi	device exam brain, helmint	smith, jf	1972
JWIDA	82	112	114	odvi	occurrence demodex, oklaho	carpenter, jw; fr/	1972
JWIDA	82	163	164	odvi	devi, exam sm intes, helmi	doster,gl	1972
JWIDA	83	233	236	odvi	elaeophoros, distri se usa	prestwood,ak; rid	1972
JWIDA	84	304	310	odvi	ecol rela mening wor, cerv	anderson, rc	1972
JWIDA	84	381	383	odvi	mening worm parelaph, okla	carpenter, jw; jo/	1972
JWIDA	92	136	143	odvi	parelaphostrongylus, maine	gilbert,ff	1973
JWIDA	92	182	193	odví	hepatozoon procyonis, texa	clark,ka; robins/	1973
JWIDA	101	11	17	odvi	strongyloidosis, captiv de	forrester, dj ta/	1974
JWIDA	102	146	148	odvi	strongyloid, fawns, florid	forrester,dj; ta/	1974
JWIDA	104	404	409	odvi	distrib muscleworm, se usa	prestwood, ak; ne/	1974
JWIDA	112	256	262	odvi	ceel, morph trypanos, mich	stuht, jn	1975
JWIDA	121	65	71	odvi	ixodes scapularis, ontario	watson, tg; anders	1976
JWIDA	123	361	366	odvi	eff 6 fasciolicides, fasci	foreyt,wj; todd,a	1976
JWIDA	123	380	385	odvi	dosh, parasit, common rang	prestwood, a; pur/	1976
JWIDA	131	2	8	odvi	culicoid vector hemorr, ky	jones, rh; rought/	1977
JWIDA	131	9	16	odvi	trans hemorrhag dis, culic	foster, nm; breck/	1977
JWIDA	141	89	96	odvi	mening worm parelaphos, nh	thurston, dr; stro	1978
JWIDA	151	51	56	odvi	sarcocystis, herbi, nat bi	pond, db; speer, ca	1979
JWIDA	151	55	56	odvi	elaeophora schneideri, tex	forent, wj; forent	1979
JWIDA	151	83	89	odvi	clin-path change, fasciolo	forent, wj; todda	1979
JWIDA	152	273	280	odvi	exp infect, elaeophora sch	titche, ar; prestw	1979
JWIDA	153	405	408	odvi	gast-intest helminths, ill	<pre>cook,tw; ridgewa/</pre>	1979
.TWMAA	52	141	158	ivho	tissue chan, lungworm infe	coble fo	1941
JWMAA	152	216	220	odvi	disease, winter mortality	cheatum el 1	951
TWMAA	312	299	303	odvi	alal pneumostrongyl minn	karns nd	1967
TWMAA	313	455	459	odvi	theileriagis in texas wtd	robinson rm· kut/	1967
JWMAA	324	963	966	odvi	pneumostrongylus, maine	behrend.df: witte	1968
JWMAA	334	888	894	odvi	tech surv helmin protoz	samuel.wm. traine	1969
JWMAA	343	546	552	odvi	mort, tissue destr. ticks	bolte in: hair i/	1970
TWMAA	364	1349	1353	odvi	dist meningeal worm canad	hindernagel is a	1972
TWMAA	372	183	186	odvi	deer ked infestatio texas	davis iw	1973
TWMAA	373	327	330	odvi	mening worm moose pop out	saunders bn	1973
TWMAA	403	579	581	odvi	protozo helmnth key deer	schulte iw klim/	1976
TWMAA	41 2	169	177	odvi	manage impl abomagal parag	eve the kellogg f	1977
	·	207	177	UUVI	manage impi abomabai parab	eve, jii, kerrogg, i	2777
MUATA	159	1	20	odvi	helminths of deer, minneso	olsen,ow; fenster	1943
NAWTA	1	473	478	odvi	lungworm situation, michig	o'roke,ec	1936
NAWTA	4	244	249	odvi	prevalenc, dis, para, mich	whitlock,sc	1939
NAWTA	28	225	232	odvi	tropical cattle fever tick	marshall,cm; sea/	1963
NAWTA	33	364	372	odvi	domes rumin, endoparasites	samuel,wm	1968

odvi continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR 49 1970 NFGJA 17--1 45 odvi nematode, pneumostrong, ny behrend, df NFGJA 20--2 158 161 odvi liver flukes, adironda, ny behrend, df; matt/ 1973 PAVEA 2---- 360 379 odvi develop pneumostrongy, cns anderson, rc 1965 PCGFA 12--- 224 odvi liver flukes, southeast de holland, jb 1958 227 odvi incid, pneumostrongyl, vir dudak,d; cornwel/ 1965 PCGFA 19--- 128 141 PCGFA 29--- 651 655 odvi nasal bots, w-t dee, se usa nettles, vf doste 1975 PHSWA 6---2 102 104 odvi helminth parasites, florid dinaburg, ag 1939 PHSWA 42--2 141 143 odvi range exten rec, helmin, ky heuer, de; philli/ 1975 odvi helminth paras, nj, oklaha pursglove, sr, jr PHSWA 44--1 107 108 1977 PHSWA 44--2 179 odvi trypanosoma cervi, se usa kingston,n; crum, 1977 184 PRLBA 206-- 395 402 odvi malaria paras, rel plasmod garnham, pcc; kutt 1980 PSDAA 48--- 47 57 odvi odhe, parasites, so dakota boddicker,ml; hug 1969 PUNMA 79-18 1 6 odvi nema, ostertagia, vir deer dikmans,g 1931 SENTD 4---3 192 odvi tick survey, sou tex count garnett, wh; ahren 1979 194 TAMSA 54--2 138 144 odvi lungworms from w-t deer, m dikmans,g 1**9**35 TISAA 60--2 203 203 odvi lone star ticks, illinois montgomery,gg; ha 1967 TRCIA 34... 57 92 odvi helminth, arthro parasites anderson, rc 1962 VTPHA 13--5 381 odvi exp parelaphostrongy infec nettles, vf; prest 1976 393 WDABB 2---4 100 107 odvi 2 surv meth, helminth infe samuel, wm; beaudo 1966 odvi tick transm, theileriasis kuttler, kl; robi/ 1967 WDABB 3---4 182 183 WDABB 4---4 142 143 odvi isolation of a babesia emerson, hr; wrigh 1968 WDABB 5---1 25 26 odvi sarcocystis in w-tail deer karstad,1; traine 1969 WDABB 5---2 108 odvi new host, amblyommainornat cook, rs; glazene/ 1969 108 WDABB 5---3 137 139 odvi comp, parasit, cent, e tex emerson, hr 1969 WDABB 5---3 351 356 odvi eimeria, penn, tex, wiscon anderson, dr; samu 1969 WDABB 5---4 398 399 odvi trypanosomiasis in w-t dee kistner, tp; hanso 1969 59 odvi path, exp fasciola hep inf presidente, pja; / 1974 WIDIA 63--- 1 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AESAA 68--1 63 67 odhe dermacentor alb hydridizat ernst, se; gladney 1975 AJVRA 38--8 1205 1209 odhe hist study, exp sarcoc hem koller,1d; kistn/ 1977 odhe life hist, food hab, calif dixon, js CAFGA 20--3 181 282 1934 CAFGA 56--1 71 71 odhe jay feeding on ectoparasit schulz, ta; budwis 1970

odhe continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS-----AUTHORS-----YEAR CJZOA 53--7 977 992 odhe behavior of nose bot flies anderson, jr 1975 187 1943 CNRDA 21--6 171 odhe 2 dipterous paras, lif his cowan, i mct odhe parasites, dis, injur, b c cowan,i mct CNRDA 24--3 71 103 1946 GRBNA 37--3 407 410 odhe occur ticks, mule de, utah pederson, jc 1977 IZSPB 1---- 1 45 odhe odvi, checklist, parasites walker,ml; becklu 1970 1940 JAVMA 96--- 542 542 odhe host, nemato eye wor, cali oberhansley, fr JMENA 10--2 225 226 odhe euschoengastia sp, oregon easton, er; krantz 1973 JMENA 13--2 169 173 odhe dada, transf bovicola, cal westrom, dr; nels/ 1976 odhe hematophag acti, hybomitra clark,gg; hibler/ 1976 JMENA 13--3 375 377 JMENA 16--5 448 449 odhe barbary sh, ectoparas, tex gray,gg; pence,db 1979 JOPAA 45--4 32 32 odhe louse flies, w montana senger, cm; capell 1959 JOPAA 45--4 32 odhe cephenemyia jellisoni, mon capelle,kj; senge 1959 32 JOPAA 53--2 392 394 odhe nematodirus odocoilei becklund, ww; walk 1967 JOPAA 56--5 1039 1040 odhe ear scab mites, psoroptes roberts, ih; mele/ 1970 JOPAA 60--6 972 975 odhe prev eye worm, 3 pop, oreg beitel, rj; knapp/ 1974 JWIDA 6---2 110 111 odhe experim infect, elaeophora hibler, cp; adcoc/ 1970 JWIDA 8---4 325 326 odhe trypanosomes, n mex, color clark,gg 1972 JWIDA 9---1 34 odhe treat, dictyocaulus, capti presidente, pja; / 1973 40 JWIDA 9---1 41 43 odhe doca suscep dictyocaul iso presidente, pja; k 1973 JWIDA 9---3 213 odhe host, filarial worms, calif weinmann, cj; and/ 1973 220 JWIDA 10--1 44 46 odhe exp infect elaeophora schn hibler, cp; gates/ 1974 JWIDA 10--4 361 369 odhe morph larv stag elaeophora hibler, cp; metzge 1974 JWIDA 11--2 214 odhe exp induc fasciola infect kistner, tp; kolle 1975 220 JWIDA 11--4 519 521 odhe trypanosomes, mule d, wyom kingston,n; mort/ 1975 JWIDA 12--1 86 odhe doca, sp diff, sarcocystis hudkins-vivion,g/ 1976 87 JWIDA 13--1 33 39 odhe trypanosoma cervi, wyoming matthews,mj; kin/ 1977 JWIDA 13--1 80 84 odhe sarcocystis hemionilatrani hudkins,g; kistne 1977 JWIDA 13--2 137 odhe mening worm indu neuro dis nettles, vf; pres/ 1977 143 JWIDA 15--1 51 53 odhe sarcocystis, herbi, nat bi pond, db; speer, c 1979 JWMAA 12--1 105 106 odhe granular tapeworm, echinoc cowan, im 1948 JWMAA 23--1 119 122 odhe cysticerci of taenia, colo olsen,ow; william 1959 NAWTA 18--- 168 188 odhe dosh, parasite interrelati longhurst, wm; dou 1953 NAWTA 22--- 160 166 odhe dosh, exp trans, nematodes baker1nf; longhu/ 1957 PHSWA 18--2 120 122 odhe porcupine nematode, colora olsen, ow; tolman, 1951 PHSWA 39--1 135 odhe prev helminth paras, monta worley, de; eustac 1972 138 PHSWA 45--1 139 odhe helmin, sympat barbary she gray,gg; simpson/ 1978 141 PHSWA 47--2 266 267 odhe gongylonema, n. w. califor botzler, rg; fanto 1980

odhe continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR PPETA 19--4 139 147 odhe ixodes sp, mule dee, skunk cooley, ra; kohls, 1943 PPETA 21--2 48 57 odhe deer lousefly calif hare, je 1945 PPETA 21--3 120 120 odhe cephenemyia, nasal bot 1945 herman, cm. TAMSA 88--4 589 591 odhe new, eimeria ivensae, mont todd,ks,jr; ogara 1969 odhe internal parasites, wyomin landram, jf; hones 1955 WGFBA 8---3 13 22 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ceel subcutaneous filariases in dykova,i; blazek, 1972 ACVTB 41--1 117 124 ACVTB 41--2 197 201 ceel microfilariae, tiss reacti dykova,i 1972 APMIA 65--4 613 620 ceel tick, encephal virus, swed svedmyr, a; zeipe/ 1965 APRPA 8--21 345 350 ceel paramphistomum cervi, dise zadura, j 1960 ceel echinococcus granulo, banf green, hu CAFNA 63--5 204 205 1949 CJZOA 47--5 795 803 ceel incidenc, abund, parasites flook, dr; stenton 1969 1969 FPARA 16--1 74 74 ceel elaphostron, cent nerv sys dykova,i JAVMA 140-7 689 690 ceel transfer, psoroptic mites hepworth, wg; thom 1962 JOPAA 59--6 1132 1133 ceel trypanosomes from elk, wyo kingston,n; morto 1973 JOPAA 61--1 17 ceel trypanosom cerv sp. n. wyo kingston, n; morto 1975 23 JPROA 21--3 448 448 ceel babesia sp. in sc red deer blewett, da; adam/ 1974 JWIDA 5---3 348 ceel reserv host, domest parasi worley, de; barre/ 1969 350 JWIDA 8---1 57 ceel doca, cross-transmis exper presidente, pja; / 1972 62 JWIDA 9---2 148 ceel neurol dis, meningeal worm carpenter, jw; jo/ 1973 153 JWIDA 10--1 63 ceel trypanosomes, horse flies davies, rb; clark, 1974 65 JWIDA 11--1 40 44 ceel preval dictyoc vivp infect bergstrom, rc 1975 JWIDA 11--1 66 ceel psoroptic mange in wapiti colwell, da; dunla 1975 67 JWIDA 13--2 149 154 ceel prev, parelaphostr, capt p woolf,a; mason,c/ 1977 JWIDA 14--2 263 268 ceel neurolog diseas, capt herd olsen,a; woolf,a 1978 JWIDA 15--1 33 36 ceel prev, parelaphostron, capt olsen, a; woolf, a 1979 JWIDA 15--1 51 53 ceel sarcocystis, herbi, nat bi pond,db; speer,c 1979 KLVBA 1962. 49 ceel dada endoparasites, denmar gui'dal, ja 1962 61 NEZTA 24--1 22 23 ceel elaphostrongylus cervi in mason, pc; kiddey/ 1976 NEZTA 24--1 23 23 ceel dicty viv, elaph cer, wapi mason, pc; mcallum 1976 NEZTA 24-11 263 ceel elaph cer, gross hist lesi sutherland, rj 1976 266 NZASA 11--4 182 183 ceel gastrointest parasitism in mason, pc 1977 ceel continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR NZJSA 11--4 686 692 ceel ophryoscolecid ciliat, n z clarke, rtj 1968 PAHUA 9---- 73 84 ceel seasonal incidence larvae sugar,1 1976 PAHUA 10... 95 ceel occur elaphostron, hungary sugar,1; kavai,a 1977 96 PARAA 73--1 1 11 ceel isola, char, babesia from adam, kmg; blewet/ 1976 PAVEA 6---- 185 213 ceel path-elaeophorosis adcock, j1; hible/ 1969 PHSWA 42--2 179 ceel mult stag tryp cerv, splee kingston, n; morto 1975 181 PHSWA 47--2 266 267 ceel gongylonema, n. w. calif botzler, rg; fanto 1980 TMPRA 26--1 60 69 ceel onchocera flexuosa, germny schulz-key,h 1975 TMPRA 26--3 348 358 ceel filariidae of, germany schulz-key,h 1975 VETRA 100-- 252 252 ceel warble fly in red deer (co dear, jp 1977 VETRA 100-- 346 346 ceel warble fly in red deer (co ray, rj 1977 WDABB 1---4 48 48 adcock, j1; hibler 1965 ceel elaeophoriasis in elk WDABB 2---1 5 6 ceel dictyocaulus, s-cent monta barrett, re worley 1966 WDABB 5---1 23 24 ceel some parasites, new mexico wilson,gi 1969 WDABB 5---1 27 30 ceel elaeophorosis, new mexico hibler, cp; adcoc/ 1969 WDABB 5---2 95 98 ceel fringed tapeworm, yellowst jacobson, rh; wor/ 1969

WGFBA 8---4 25 28 ceel eimeria of elk, new specie honess, rf 1955

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AESAA 67--- 204 214 alal tax, lif hist moos fly, wy burgerr, jf; ander 1974 AJVRA 3---9 403 408 alal parasites, northern minnes olsen, ow: fenster 1942 AJVRA 27--- 548 558 alal cerebro nema, minn, pneummo kurtz, hj; loken, / 1966 ATMPA 58... 307 314 alal intro reindee, hydatid dis sweatman,gl 1964 BEREA 17--- 249 249 alal moose-tick in saskatchewan cameron, ae; fulto 1927 CAENA 102-- 1461 1473 alal od, tabanid, distr, ontar smith, sm; davies/ 1970 CJZOA 43--4 635 639 alal neurologic signs, ontario anderson, rc 1965 CJZOA 52--2 235 245 alal nematode, trematod, manitob lankester, mw 1974 CJZOA 52--3 401 403 alal od, elaphostrongy, alberta samuel,wm; holmes 1974 CJZOA 54--3 307 alal helminths in moose, albert samuel, wm; barre/ 1976 312

alal continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CNVJA 5--11 287 296 alal elaphostrongylosis, deer, smith, hj; archib/ 1964 CNVJA 12--2 56 alal infect dictyocaulus calv gupta, rp; gibbs, h 1971 56 JAVMA 37--5 770 775 alal parasites, moose, minnesot wallace, fg 1934 JAVMA 171-9 987 988 alal cerebr nematodiasi, n hamp stackhouse, 11 1977 JEBCA 67--- 24 29 alal odhe, dermacen tick, paral wilkinson, pr 1970 JOMAA 39--1 139 145 alal paras, dis, wells gray her ritcey, rw; edward 1958 JOMAA 39--3 449 450 alal onchocerca in an alaska mo williams, rb; babe 1958 JOMAA 48--4 668 669 alal parasites, newfoundland threlfall,w 1967 JOPAA 18--4 219 231 alal new disease in moose, minn thomas, 1j; cahn, a 1932 JOPAA 18--4 303 303 alal hematodirella, moose, minn dikmans, g 1932 JWIDA 8---3 242 244 alal elaeophorosis, montana worley, de; and er/ 1972 JWIDA 15--2 281 284 alal dermacentor, ontario addison, em; john/ 1979 JWMAA 23--1 122 alal preva, hydatid dis, alaska rausch, ra 1959 123 JWMAA 31--2 299 303 alal pneumostrong, deer, minnes karns, pd 1967 JWMAA 33--2 431 433 alal pneumost, deer-free island karns, pd; jordan 1969 JWMAA 37--1 327 330 alal odvi, mening worm, ontario saunders, bp 1973 JWMAA 38--1 42 46 alal parelaphostrongylus, maine gilbert, ff 1974 NCANA 101-- 23 50 alal infect, parasi dis, n amer anderson, rc; lan 1974 PAVEA 1--- 289 322 alal neuro dis, exp pneumostron anderson, rc 1964 PAVEA 2---- 360 alal motor ataxia, paral, pneum anderson, rc 376 1964 XENOB 6.... 168 169 alal deer louse fly, se finland brander,t 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AEHLA 25--4 246 252 rata obs on some nat-foc zoonos rausch,r1 1972 AGBOB 12--1 23 28 rata warble-fly problem in alas washburn, rh; kle/ 1980 AVCSA 3---3 201 rata cereb, muscu nematod, swed roneus,o; nordkvi 1962 225 AVCSA 6---4 353 359 rata experim babesia infections nilsson,o; nordk/ 1965 AVCSA 14--4 642 644 rata onchocercosis in swedish r rehbinder, c 1973 AVCSA 16--1 14 23 rata giant cell reac, echinococ roneus,o 1975 AVCSA 17--3 359 362 rata echinococcosis, slaughtere kummeneje,k; waag 1976 AVCSA 18--1 75 85 rata keratitis in reindeer rehbinder,c 1977 AVCSA 18--1 86 90 rata dictyocaulus viviparus inf kummeneje,k 1977 ATMPA 58... 307 314 rata intro reindee, hydatid dis sweatman,gl 1964 rata continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BPURD 1---- 509 rata warble fly distrib, canada kelsall, jp 1975 517 CAFNA 90--2 189 1976 191 rata paras, woodland caribo, bc low,wa CJZOA 49--2 159 166 rata odvi, neurolog dis, ontari anderson, rc 1971 CJZOA 49--2 280 rata fascioloides magna tremato chouquette, 1pe; / 1971 281 CJZOA 54--5 680 684 rata protostrongylid nematode lankester, mw; cr/ 1976 CNJMA 21--6 199 203 rata parasites, reindee, canada choquette, 1pe; w/ 1957 CNRDA 10--- 527 531 rata skrjabinema oreamni, canad swales, we 1934 DOBTA 119-3 621 621 rata life cycle, nematode, rein mitskevich, v iu 1958 FPWTA 33--- 30 39 rata dis condit, parasi, bar gr broughton, e; chou 1969 JOPAA 9---1 38 39 rata parasites, alaska 1922 hadwen,s JOPAA 9---1 48 rata nematodirus tarandi, reind hadwen,s 48 1922 JOPAA 28--5 423 423 rata parasites, woodland caribo erickson, ab; high 1942 JOPAA 46--5 624 628 rata redescri, avitellina arcti gibbs,hc 1960 JWIDA 7---4 242 245 rata occure setaria in reindeer dieterich, ra; lui 1971 JWIDA 9---4 376 378 rata mort due to meningeal worm trainer,dd 1973 JWIDA 12--4 566 rata besnoitiosis in woodla car wobeser,g 1976 571 NOVDA 16--4 390 395 rata hist stud, parasit nodule 1964 lisitzin,p NOVDA 25--6 203 210 rata elaphostrongylosis, reind bakken,g; sparboe 1973 NOVTA 30--4 214 216 rata gastric para, winter slau rehbinder,c; szo/ 1978 NOVTA 31--6 282 283 rata parasit visceral granuloma rehbinder,c; chr/ 1979 PARAA 66--1 123 132 rata solenopotes-tarandi, ba-gr weisser, cf; kim, k 1973 ZEVMA 25B-1 81 87 rata antiparas eff, tetramisolu kurkela, p; kaante 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JOPAA 19--3 246 246 anam onchocerca, subcut tissue 1933 dikmans,g JOPAA 28--2 167 168 1942 anam a new coccidium from anam huizinga,h JOPAA 40--6 637 648 anam internal parasites, n dako goldsby, ai; evele 1954 JOPAA 44--5 514 514 anam ornithodoros (otobins) meg schad,ga 1958 JOPAA 46--5 671 671 anam thysanosoma actinioides allen, rw; samson, 1960 JOPAA 56--4 759 767 anam dosh, immun, haemonch isol allen, rw; samson/ 1970 JOPAA 56--5 917 917 anam otobius-megni, ears, n mex meleney, wp; rober 1970 JWIDA 8---4 332 334 anam capillaria-hepatica, alber barrett, mw; chalm 1972 JWIDA 10--1 60 62 anam infect podo dermati, alber chalmers,ga; barr 1974

anam continued on the next page

anam protostr-macrotis nem, wyo greiner, ec; worl/ 1974

JWIDA 10--1 70

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR PHSWA 12--1 2 4 anam distribut, pseudostertagia lucker, jt; dikman 1945 PHSWA 20--2 96 97 anam thysanosoma, pronghorn ant allen, rw; kyles, p 1953 PHSWA 27--1 69 73 anam helminth parasites, ne mex gilmore, re; allen 1960 PHSWA 42--1 61 anam dosh, marshallagia, wyomin bergstrom,rc 63 1975 PNDAA 6---- 25 1955 25 anam coll, ident parasit. nd an goldsby,ai PNDAA 17--- 70 71 anam od epizootic, n dak badlan richards, sh 1963 TRSTA 66--2 335 335 anam trypanosome infection, cap roberts, cj; gray, 1972 WDABB 3---2 71 73 anam inci, eimeria antelocaprae todd,ks jr; hamm/ 1967 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AESAA 63--3 635 639 bibi associati face fly, n amer burger, jf; anders 1970 bibi parasites, disease, canada chouquette, 1pe; / 1961 CNVJA 2---5 168 174 COVEA 69--2 198 205 bibi ostertagiosis, case rep, n wade, se; haschek/ 1979 JAVMA 119-- 386 bibi parasitism in bison 1951 387 frick.ej JOPAA 22--5 517 518 bibi arthropod, helminth parasi roudabush,rl 1936 JOPAA 39--1 58 bibi parasites, bibi, nw u s 1953 59 locker,b JOPAA 50--5 655 bibi speleognathus australis, b drummond, ro; mad/ 1964 655 JOPAA 53--4 724 bibi sheep liver fluke, wyoming bergstrom,rc 724 1967 JWIDA 11--3 412 414 bibi bovine coccidia, wyoming ryff,k1; bergstro 1975 JWIDA 15--1 51 53 bibi sarcociptis, herbi, nat bi pond, db; speer, ca 1979 NAWTA 28--- 233 239 bibi paras, dis, anthrax, canad novakowski, ns; c/ 1963 NOSCA 26--2 61 64 bibi tick paralysis, amer bison kohls,gm; kramis, 1952 NPSMD 1.... bibi bison, yellowstone natl pk meagher,mm 1973 • • • VETRA 100-- 406 406 bibi redwater in, babesia major findlay, cr; begg, 1977 WMBAA 16--- 1 52 bibi biol, mngt, wood buf na pk fuller,wa 1966 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ARMIA 25--- 231 254 ovca pathogenicit, soil amoebas culbertson, cg 1971 CEXIA 8---2 345 354 ovca lungw infec, homocytotr an hudson, rj; bandy/ 1971 CJCMA 36--1 69 73 ovca cell adher, lungworm infec hudson, rj; bandy/ 1972 ovca continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 41--6 913 918 ovca parasi, calif bigh, sou bc blood, da 1963 CJZOA 49--4 507 512 ovca helminths, rocky mt, canad uhazy, ls; holmes, 1971 CJZOA 49-11 1461 1464 ovca coccidia, rocky mt, canada uhazy, 1s; mahrt,/ 1971 CJZOA 51--8 817 824 ovca lungworms, rocky mt, canad uhazy, ls; holmes/ 1973 CJZOA 53--4 391 394 ovca lymp cell adher, protostro hudson, rj 1975 DABBB 40B-4 1640 1640 ovca pathogenesis pulm protostr spraker,tr 1979 JKESA 35--4 369 370 ovca a new species, mallophaga emerson,kc 1962 JOPAA 41--6 583 ovca parasites, new mexico 1955 587 allen,rw JOPAA 52--3 618 621 ovca oestrus ovis, wyom, montan capelle,kj 1966 JOPAA 53--1 157 165 ovca parasi, checklist, rocky m becklund, ww; seng 1967 JOPAA 58--1 29 33 ovca moufflon hybri, protostron monson, ra; post, g 1972 JOPAA 59--1 136 140 ovca redescr trichuris-schumako knight, ra; uhazy, 1973 JWIDA 6---3 169 170 ovca mycoplas, pneumo, rocky mt woolf,a; kradel,/ 1970 JWIDA 8---4 389 389 ovca transplac trans, protostro hibler, cp; lange/ 1972 JWIDA 10--1 39 41 ovca trans placen, protostr inf hibler, cp; metzg/ 1974 JWIDA 11--2 193 ovca wyominia-tetoni, washingto colwell, da; dun1/ 1975 194 JWIDA 12--1 48 ovca influ rain, lungworm infec forrester, dj; 11/ 1976 51 JWIDA 13--2 117 124 ovca pneumon, muelleri sp, capt demartini, jc; dav 1977 JWIDA 13--2 125 130 ovca helminth parasites, oregon kistner, tp; mat1/ 1977 JWIDA 13--3 248 ovca prenat infec, lungw, alber gates, cc; samuel, 1977 250 JWIDA 15--4 561 561 ovca transplacental trans, prot kistner, tp; wyse/ 1979 JWIDA 16--1 77 ovca psoroptic scabils, new mex lange, re; sandov/ 1980 82 491 JWMAA 28--3 481 ovca surv, lungworm infect, mon forrester, dj; sen 1964 JWMAA 38--4 771 774 ovca lamb prod, surv, mort, col woodard, tn; guti/ 1974 JWMAA 43--2 461 467 ovca eval drug treatment, lungw schmidt, rl; hib1/ 1979 PHSWA 10--1 8 9 ovca lungwor, protostron, yello dikmans,g 1943 PHSWA 19--1 39 39 ovca parasites, new mexico allen, rw; kennedy 1952 PHSWA 23--1 68 69 ovca lungworms, montana marquardt,wc; sen 1956 PMASA 22--- 82 92 ovca land mollusca, protostrong forrester, dj 1962 WDABB 3---2 74 76 ovca virus isola, lung, nemato forrester, dj; wad 1967

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS AUTHORS	YEAR
JPROA	212	197	199	ovda	new	spec protozoa, eimeria clark,gw; colwell	1974
JWIDA	134	427	428	ovda	ρορι	ula, bovicola jellisoni kim,kc	1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 36--4 529 532 obmo helminth parasit, nwt, can gibbs,hc; tener,j 1958 CJZOA 55--6 990 obmo ovca, new eimeria species duszynski,dw; sa/ 1977 999 CNVJA 17--5 138 obmo haemonchosis, captive calf macdonald, dw; sa/ 1976 139 ETBRA 30--- 222 224 obmo hypodermatid larvae dipter jansen, j, jr 1970 JAVMA 131-3 195 196 obmo parasitosis in a musk ox durrell, wb; bolto 1957 JWMAA 38--4 775 782 obmo parasitic infection in mus samuel, wm; gray, d 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARCNRDA 10--- 527531oram skrjabinema oreamni, canad swales,we1933JOPAA 51--6945947oram nematodirus maculosusbecklund,ww1965JWMAA 30--4786790oram parasites, w centr alberta kerr,gr; holmes,j 1966jumaa 35--194103oram parasite, pestic, so dakot boddicker,ml; hu/ 1971

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR
ACVTB 43--4 411
                416 many treatment fasciola hepatic chroustova,e
                                                                      1974
DKBSA 199-- 505
                506
                          dev caus agent, setariosis shol,va; drobisch 1972
ENAMA 21--1 1
                220
                     many melophaginae (diptera)
                                                     bequaert, j
                                                                      1942
FPARA 19--3 253
                256
                     many studies, toxoplasmosis, na catar,g
                                                                      1972
FPARA 24--1 35
                     many helmint, wiru, intro czech kotrla,b; kotrly, 1977
                40
IJLAA 43-10 944
                947 mamm species composit, coccidia bhatia, bb; chauh/ 1974
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CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JMENA 13--1 115 118 repellency of n,n-diethyl- schreck,ce; smit/ 1976 275 JMENA 13--3 225 many anim dis agent tran, flies krinsky,wl 1976 JMENA 13--3 375 377 hematophag, hybomitra spec clark,gg; hibler/ 1976 JOHLA 40--- suppl 1 mamm bibliography, lung nematod forrester, dj; fo/ 1966 JOPAA 56--4 6 7 many ecol relation, meningea wo anderson, rc 1970 many tax, host, dist sefaria, u becklund,wm; wal/ 1969 JOPAA 55--2 359 368 many helminths, big game, s.d. boddickey, ml; hu/ 1969 JOPAA 55--5 1067 1074 JOPAA 56--4 408 many furthr toxoplasma isolatns catar,g 408 1970 JOPAA 57--1 191 many prev, geog distr, cooperia burtner, rh; beckl 1971 192 JWIDA 6---4 194 204 ecol approach, zoonoti dis johnson, hn 1970 JWIDA 9---1 21 25 many horse fli, elaephor, n mex clark,gg; hibler, 1973 JWIDA 9---2 160 parasites of blackbuck ant thornton, je; gal/ 1973 162 JWIDA 11--4 486 many elaleophora-schneid, monta worley, de 1975 488 many trypanoso, game anim, wyom morton, jk; kingst 1976 JWIDA 12--2 233 236 mamm prev antibod, toxoplas, ont tizard, ir; bille/ 1976 JWIDA 12--3 322 325 JWIDA 12--4 550 cerebral coenurosis, wild toofanian, f; ivog 1976 551 JWIDA 14--2 157 162 many toxoplasmosis, norway, swe kapperud, g 1978 JWIDA 15--1 51 54 sarcocystis, herbivor, mon pond, db; speer, ca 1979 NEZTA 21--3 43 47 many host paras checklist n zea andrews, jrh 1973 OUOKA 24-11 2 7 ticks can kill! hair, ja 1968 anal, host-parasi interact ractliffe,1h; ta/ 1969 PARAA 59--3 649 661 PHSWA 1---2 63 64 many helminth parasites 1934 dikmans,g PHSWA 6---2 97 101 many helminths, n.a., wild rumi dikmans,g 1939 POASA 55--- 101 102 many prevale marshallagia, wyom bergstrom, rc 1975 SEHEA 8---3 267 hematolog manif, paras inf conrad, me 1971 . . . WGFBA 9---- 1 279 many wildlife wyoming, disease honess, rf; winter 1956 ZTMPA 24--4 457 466 many studies on stephanofilaria patnaik, b 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR ACVTB 45--1 133 140 rumi dynamics of cocc, helminth tilc,k; hanuskova 1976 JOPAA 32--1 7 16 rumi hosts, protostron sp, key dougherty,ec; gob 1946 TGMEA 30... 156 ... rumi doca, theileriidae sporozo uilenberg,g 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ASISA 29--- 536 536 wiru parasitic bronchopneumonia julini,m; peracin 1976 NEZTA 21--3 43 47 wiru host-parasite checklist, h andrews,jrh 1973 PAHUA 7---7 181 189 wiru occ of nasal throat bot f1 sugar,1 1974 ZTMPA 24--4 467 478 wiru species of haemonchus, dom sachs,r; gibbons/ 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWIDA 15--1 51 53 herb sarcocystis in free-rangin pond,db; speer,ca 1979 PAHUA 9---- 85 96 biga incidence larvae of hypode sugar,1 1976 TMPRA 29--2 223 233 ungu invest, trypanosome carrie drager,n; mehlitz 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR IVEJA 49--2 133 135 axax obser on fascioliasis, zoo rao,at; acharjyo, 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ACVTB 43--1 65 caca helminths, coccidia, ecolo dyk,v; chroust,k 1974 77 NATUA 200-2 490 490 caca host, muellerius capillari fairlie,g 1963 NATWA 65--- 395 395 caca intermediate host, coccidi entzeroth,r; sch/ 1978 PAHUA 9---- 67 71 caca dipetalonema rugosicauda n meszaros, f; sugar 1976 PARAA 55--4 739 475 caca gastro-int helminths, brit dunn,am 1965 RSZOA 81--1 13 24 caca endoparasites of roe deer andrews, jrh; hor/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR JAASA 42--- 133 133 dada surv, intrnl paras, alabam brugh,th,jr 1971 JWIDA 13--1 55 58 dada nat occur neurologic disea kistner,tp; john/ 1977 JWIDA 13--4 440 444 dada cerebrospnl parelaphostron netties vf: pres/ 1977

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CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARJZOOA 177-4 494 496 dada occurrence certain ectopar jackson,j1975PHSWA 41--2 250 250 dada new host rec, setaria yehi phillips,jh; har/ 19741974TMPRA 26--4 494 498 dada caca, filariidae, germany schulz-key,h1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BSKBA 31--- 153 154 ceni pathol studi, fascioliasis tomimura,t; hira/ 1979 JWIDA 14--1 137 141 ceni elaeophorosis, texas robinson,rm; jon/ 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 12-26 377 384 bibo helminthofauna, euro bison drozdz,j 1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR JWIDA 15--2 263 265 ovli gast-intest helminths, ira eslami,a; meydan/ 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ACVTB 43--2 123 131 ovmu caca, helminths, coccidia dyk,v; chroust,k 1974 FPARA 15--2 178 178 ovmu neostrongylus linearis chroust,k; dyk,v 1968 VPARD 1---2 145 150 ovmu caca, incid, trans, coccid dyk,v; chroust,k 1975

UNIT 1.2: DISEASES

The word disease can be loosely interpreted to include any departure from health, or more strictly interpreted as a response to a pathogenic microorganism. The Merck Veterinary Manual tends toward the first interpretation. as references are made to parasitic diseases, or malfunctions caused by parasites. The sorting of large numbers of references in this area was done on the basis of the second definition. Thus the reference lists in this UNIT 1.2 includes those departures from normal health that can be attributed to infectious diseases, or microscopic organisms that upset normal physiological functions and metabolic pathways.

Since extra energy demands result from infections, adequate quantities of good quality food help prevent or alleviate the effects of pathogens. While there are large amounts of information on the nutrient requirements of healthy domestic ruminants, there is very little on their requirements when exposed to infections and metabolic disorders, and even less information on wild ruminants. The general principle stated with reference to the effects of parasites also applies to diseases; animals on good range will very likely be affected less than those on poor range. Prevention of problems is better than having to cure them, and wild ruminant ranges in good condition, not suffering from overgrazing or overbrowsing, will likely not harbor an unusually large number of pathogens.

The next several pages of reference lists on diseases have been sorted according to the criteria explained above. Such sorting cannot be perfect, so there may be important papers in both UNITS 1.1 and 1.2 that should be consulted when reviewing the literature. The arrangement should greatly facilitate finding of the literature.

REFERENCES, UNIT 1.2

DISEASES

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBCPA 57A-4 391 398 arti haemoglobins, erthro sickl butcher, pd; hawke 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR JWIDA 6---4 376 383 cerv maligna catarrhal fev, tex clark,ka robins/ 1970 NAWTA 28--- 199 206 cerv leptospirosis, ontario dee mcgowan,je; kars/ 1963 WDABB 5---1 12 15 cerv 4 cases, systemati mycosis fletch,al; anders 1969 WIDIA 43--- 1 233 cerv diseases of, bibliography karstad,1, ed 1964 WIDIA 52--- 1 114 cerv dis, bibliogr supplement 1 karstad,1 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AGVIA 41--3 259 266 od-- mult epizoo haemorr diseas boorman, j; gibbs, 1973 722 od-- brucellosis, leptospirosis ferris, dh; hanso/ 1961 AJHEA 51--5 717 AJPAA 70--3 379 od-- pathogenesis, epizoo hemor tsai,k-s; karstad 1973 400 AJVRA 18-66 162 od-- vesicular stomatitis in de karstad,1; hanson 1957 166 AJVRA 22-86 109 116 od-- electr micros, anaplasma m ristic, m; watrach 1961 AJVRA 23-92 21 od-- doca, transm anaplas, tick osebold, jw; doug/ 1962 23 AJVRA 28--- 599 od-- anaplasma, sier nevad, cal christensen, jf m 1967 600 AJVRA 35--6 817 819 od-- propagat epiz hem dis, avi hoff,gl; spalati/ 1974 ANYAA 241-- 594 604 od-- sickling phenomena of deer taylor, wj; easley 1974 ASMAC 72--- 209 209 od-- quant, neutraliz, epi hemo stim, tb; debbie,/ 1972 ASMAC 74--- 213 od-- dosh, isol blueton vir, ep bando, bm 213 1974 AZATA 75--- 1 39 od-- experiment feeding of deer nichol, aa 1938 CAFGA 36--1 19 20 od-- papilloma, skin tumors, de herman, cm; bischo 1950 CAFGA 47--3 293 300 od-- abscess, corynebacterium p rosen, mn; holden, 1961 CAFGA 58--2 141 144 od-- rabies in a deer cappucci, dt, jr; 1 1972 CJCMA 35--3 187 191 od-- bluetongue virus, comparas thomas, fc; traine 1971 CNREA 31-10 1373 1375 od-- pulmonary fibroblastomas i koller,1d; olson, 1971 CNVJA 3---3 71 od-- cul, ser evi, lepto, ontar abdulla, pk; kars/ 1961 78 - COVEA 49--1 97 115 od-- latent anaplasma infection osebold, jw; chri/ 1959 CVTJA 24--1 42 42 od-- traumatic splenitis in a d kapur, mp; sadana, 1976 DABBB 33B-3 1165 1165 od-- cell-virus inter, char epi tsai,k-s 1972 DABBB 36B-8 3806 3806 od-- study vasc lesions catarrh clark,ka 1976 DABBB 39B-- 5250 5250 od-- characteriz, virus epiz he willis,ng 1979 od-- tuberculosis in captive de basak,dk; chatte/ 1975 IJAHA 14--2 135 136 INFIB 8---3 463 474 od-- ultrast char genome epizoo tsai,k-s; karstad 1973 IVEJA 54--5 409 410 od-- pasteurella multocida infe srinivasan,va; v/ 1977 od-- continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 91---2186 192 od-- pseudotuberculosis of deer hammersland, h; jo 1937 JAVMA 98--- 129 131 od-- caseous lymphadenitis seghetti, 1 mcken 1941 JAVMA 99--- 54 54 od-- paratyphoid, fawn, n carol gauger, hc; gordon 1941 JAVMA 112-- 337 337 od-- foot-mouth dis, wildl, cal anonymous 1948 JAVMA 130-- 513 od-- leptospir, brucello titers wedman, ee; driver 1957 514 JAVMA 132-7 289 292 od-- infect, antibody, anaplasm christensen, jf; / 1958 JAVMA 132-7 293 296 od-- serol surv, leptospi, mass reynolds, im; smit 1958 JAVMA 136-9 426 od-- latent anaplasma marg infe christensen, jf; / 1960 427 JAVMA 139-8 892 896 od-- inves, brucel, lepto, illi ferris, dh; hanso/ 1961 od-- brain tumors (ependymomas) kradel,dc; dunne, 1965 JAVMA 147-- 1096 1098 JAVMA 167-7 565 od-- surv, toxoplasma antibodie franti,ce; conno/ 1975 568 JAVMA 169-9 975 979 od-- necropsy, lab findin, free reed, de; shave, h/ 1976 JAVMA 175-9 914 915 od-- obs on diag test, paratube temple, rms; musc/ 1979 JCVPA 87--3 345 352 od-- epiz haemorr dis, tis cult lawman, mjp; gibb/ 1977 JIDEA 58--5 366 368 od-- doca, trans of warts, host koller, 1d; olson, 1972 JOPAA 29--1 80 80 od-- corynebacterium, louse fly steinhaus, ea 1943 JWIDA 6---4 479 482 od-- sero ev, arbovir, wis, tex issel,cj; hoff,g/ 1970 JWIDA 8---1 72 74 od-- obs on malignant catarrhal clark,ka; robins/ 1972 JWIDA 13--2 184 190 od-- dermatoph-congol enzoo, ny gordon, ma; salki/ 1977 JWMAA 8---3 247 250 od-- contag panophthalmitis, wy rosenfeld, i; beat 1944 JWMAA 15--2 216 220 od-- disease, winter mortal, ny cheatum, el 1951 JWMAA 24--1 44 52 od-- leptos, bruc ser reac, wis trainer, do; hanso 1960 JWMAA 28--2 377 381 od-- epizootic hemorrhagic dise trainer, do 1964 JWMAA 30--4 777 780 od-- serol stud, epiz hemorra d wilhelm, ar; train 1966 LANCA 2---- 1374 1375 od-- ruptured psoas in deer 1972 thrower, wr MMLRA 4---3 75 78 1974 od-- mortality in deer mcdiarmid, a NAWTA 3---- 886 889 od-- malignant edema in deer mckenney,fd 1938 NAWTA 10--- 242 246 od-- dee, domes livest par, dis herman, cm 1945 NAWTA 16--- 164 178 od-- epizootic, foot rot, calif rosen, mn;/ 1951 NAWTA 21--- 173 184 od-- epizootic in deer in michi fay, ld; boyce, ap/ 1956 NAWTA 23--- 133 136 od-- southeast coop diseas stud hayes, fa; greer,/ 1958 NAWTA 29--- 196 201 od-- virus, epizo hemorrhag dis ditchfield, j; de/ 1964 NAWTA 30--- 196 205 od-- pathogen, viral hemorrhagi debbie, jg; rowse/ 1965 NAWTA 32--- 381 386 od-- epizootiology, hemorrhagic shope, re 1967 1979 NEZTA 27--3 55 55 od-- diseases of deer mcallum.h NEZTA 27--8 151 151 od-- tuberculosis in deer hellstrom, j 1979 PCGFA 11--- 1 18 od-- se cooperative od dis stud greer, we; shotts/ 1957 PIAIA 74--- 78 86 od-- incid, dis antibodie, iowa haugen, ao 1968 od-- continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR PSEBA 88--4 533 535 od-- an infectious fibroma of d shope, re 1955 SAGCA 25--1 2 od-- fall deaths, acorn toxin 1977 2 kradel.dc od-- low incid leptospirosis, b twigg,gi; hughes/ 1973 VETRA 93--4 98 100 VIWIA 7---1 15 18 od-- tumors of deer quortrup,er 1946 VTPHA 14--6 643 647 od-- glioma, rhabdomyosa, brain holscher,ma; pag/ 1977 WDABB 1---3 31 32 od-- anitibod, myxovirus parasi shah,kv; schalle/ 1965 WDABB 5---1 3 7 od-- exper expos, calif encepha cook, rs; trainer, 1969 WDABB 5---3 174 181 od-- leptospiros, illinois herd andrews,r 1969 WDABB 5---3 295 296 od-- chronic tox hepatit, louis seger, cl; newsom/ 1969 WSCBA 4---2 31 32 od-- epithelial papillomas in d chaddock.tt 1939

CODEN VO-NU BEPA ENPA ANIM KEWO----- AUTHORS----- YEAR AJEPA 96--5 379 382 odvi exp infect venez equine en hoff,gl; trainer, 1972 AJHEA 63--3 201 203 odvi wildlife, monitors, diseas trainer, dd 1973 AJPAA 66--- 30A 31A odvi vascul chang, epizo hemorr tsai,ks; karstad, 1972 AJTHA 22--3 414 417 odvi isol jamestown canyon viru issel,cj 1973 AJVRA 20-78 925 926 odvi experimental brucellosis youatt,wg; fay,ld 1959 AJVRA 22-86 104 108 odvi epizoo hemorrag dis, s dak pirtle, ec; layton 1961 AJVRA 31--2 271 odvi bluetongue virus, w-t deer thomas, fc; traine 1970 278 AJVRA 33-12 2545 2549 odvi blood serum, erysipelothri sikes,d; kistner/ 1972 AJVRA 37--7 837 840 odvi viral particl, malig catar clark, ka; adams, 1 1976 AJVRA 38--3 361 364 odvi isola, hemorrhag dis virus fosberg, sa; stau/ 1977 APAVD 1978 169 169 odvi vit e, selen, white muscle ullrey, de; brady, 1979 AUVJA 51--4 174 177 odvi bluetongue-like disease of frank, jf; willis, 1975 CJCMA 35--3 224 229 odvi pathogen, exp epizo hemorr fletch, al; karsta 1971 387 CNJMA 32--1 382 odvi experim bluetongue disease vosdingh, ra; tra/ 1968 CNJMA 32--1 388 391 odvi staphylococcal arthritis sikes,d; hayes,f/ 1968 odvi histopath, exp bluetongue karstad,1; traine 1968 CNVJA 8--11 247 254 CNVJA 14--5 106 109 odvi dis resemblin catarr fever wobeser,g; majka/ 1973 CNVJA 20-11 323 325 odvi polioencephalomalacia in w wobeser,g; runge, 1979 odvi continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CNREA 31-10 1373 1375 odvi pulmonary fibroblastomas koller, ld; olson, 1971 CNSVA 16--2 15 15 odvi streptothricosis, dee, man dean,dj 1961 COVEA 33--4 323 odvi some diseases, minnesota fenstermacher, r;/ 1943 332 hadlow,wj COVEA 45--4 538 odvi degenerative myopathy 1955 547 odvi experim infect, leptospira ferris, dh; hanso/ 1960 COVEA 50--3 236 250 COVEA 52--1 94 odvi leptospirosis in the w-t d reilly, jr; muras/ 1962 98 DABBB 31B-- 6785 6785 odvi blue tongue virus, pt 1, 2 thomas,fc 1971 issel,cj DABBB 34B-6 3000 3001 odvi studies with arboviruses, 1973 JANSA 32--5 1037 1041 odvi rata, atherosclerosis in r wiggers, kd; jaco/ 1971 JAVMA 133-7 359 361 odvi surv, brucel, leptosp, sou shotts, eb; greer/ 1958 JAVMA 137-3 190 191 odvi serol stud, brucel, southe hayes, fa gerard/ 1960 JAVMA 140-1 53 odvi experim leptospiros infect reilly, jr; muras/ 1962 57 odvi cutaneous streptothricosis kistner, tp; shot/ 1970 JAVMA 157-5 633 635 JAVMA 159-5 605 odvi malignant catarrhal fever wyand, ds; helmbo/ 1971 610 JAVMA 167-7 571 odvi dermatophilus, alternaria salkin, if; gordo/ 1975 573 JAVMA 169-9 971 974 odvi myopathy, myoglobinuria in wobeser,g; bella/ 1976 JEMEA 108-6 797 802 odvi infectious cutaneo fibroma shope, re; mangol/ 1958 JEMEA 111-2 155 170 odvi virus-indu epizo hemor dis shope, re; macnam/ 1960 JIDIA 108-3 278 286 odvi experimental leptospirosis trainer, do, jr; k/ 1961 JNYEA 85--4 217 220 odvi epizo, bovin babesias, tex hourrigan, jl 1977 JOMAA 37--4 552 554 odvi lymphangioma, w-tailed dee chute, hl; chamber 1956 JOPAA 57--3 657 659 odvi fail boophil, trans anapla kuttler, kl; grah/ 1971 JWIDA 6---4 226 odvi anthrax epizootic, w-t dee kellogg, fe; pres/ 1970 228 JWIDA 6---4 295 odvi leptospiral antibo, se usa shotts, eb, jr; hay 1970 298 JWIDA 6---4 384 odvi bluetongue vir 1, preg dee thomas, fc; traine 1970 388 JWIDA 6---4 389 396 odvi salmonellosis, w-t d fawns robinson, rm; hid/ 1970 JWIDA 6---4 397 401 odvi use wildl, monitor zoonose trainer, dd 1970 odvi surv, stomatitis vir antib jenney, ew; hayes/ 1970 JWIDA 6---4 488 493 JWIDA 7---1 52 odvi aspergillos, renal oxalosi wyand, ds langhe/ 1971 56 JWIDA 9---1 56 57 odvi caseous lymph adenitis stauber,e; armst/ 1973 JWIDA 9---3 245 248 odvi infec calif arbovir, texas issel,cj; hoff,g/ 1973 JWIDA 10--1 25 odvi bluetongue, epizoo hemorrh hoff,gl; trainer, 1974 31 JWIDA 10--1 34 odvi subcutaneous mycotic infec salkin, if; stone, 1974 38 JWIDA 10--2 158 163 odvi blueton vir, enzootic, tex hoff,gl; trainer/ 1974 JWIDA 10--3 187 189 odvi id virus, 1971 hemorrag di thomas, fc; willi/ 1974 JWIDA 10--3 217 224 odvi 1971 hemorrhag dis, se u s prestwood, a; / 1974

odvi continued on the next page

	CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS	AUTHOR S	YEAR
	JWIDA	111	62	65	odvi	ossifving fibroma, ear tag	roscoe.de: veik1/	1975
	JWIDA	111	116	121	odvi	alal, arthropathy in white	wobeser.g: runge.	1975
	JWIDA	112	177	186	odvi	hemorrhagic dis, kentucky	roughton.rd	1975
	JWIDA	112	195	200	odvi	serol, venez eg enceph, tex	smart.dl traine/	1975
	JWIDA	113	398	401	odvi	dermatophilosi, twin fawns	roscoe.de: lund./	1975
	JWIDA	114	508	515	odvi	exp infect. rinderpest vir	hamdv.fm: dardir/	1975
	JWIDA	114	552	553	odvi	presumpti paratuberculosis	libke.kg: walton.	1975
	JWIDA	122	143	147	odvi	visual defcts, 6 case rpts	howard.dr: krehb/	1976
	JWIDA	123	306	309	odvi	prev renal urolithias, cap	woolf.a: kradel./	1976
	JWIDA	123	396	401	odvi	hepatic fatty cirrhos, tex	blankenship, lh; /	1976
	JWIDA	124	516	522	odvi	infec peste-des-petits rum	hamdy,fm; dardiri	1976
	JWIDA	131	9	16	odvi	trans hemorrhag dis, culic	foster, nm; breck/	1977
	JWIDA	132	184	190	odvi	dermatophilus dermatit, ny	gordon.ma; salki/	1977
	JWIDA	133	281	285	odvi	occur rumenitis in sup fed	woolf,a; kradel,d	1977
	JWIDA	133	297	299	odvi	fibro sarcoma in a w-t dee	elwell,mr; burge/	1977
	JWIDA	161	19	24	odvi	w-t deer reservior brucell	boeer.wi; crawfo/	1980
							, , , , , , , , , , , , , , , , , , , ,	
	JWMAA	111	91	94	odvi	actinomycosis, minnesota	cass, is	1947
	JWMAA	114	317	323	odvi	die-offs, edwards pla, tex	, 5	
	JWMAA	152	216	220	odvi	disease, winter mortal, ny	cheatum.el	1951
	JWMAA	192	320	321	odvi	surv. brucellosis. missour	steen.mo: brohn./	1955
	JWMAA	222	203	204	odvi	granular venereal disease	helmboldt.cf: mcd	1958
	JWMAA	233	345	348	odvi	brucellosis, leptosp, mich	vouatt.wg: fav.1/	1959
	JWMAA	261	27	31	odvi	experim stud, brucellosis	baker.mf; dills./	1962
	JWMAA	281	35	41	odvi	leptosp reac rates, illino	ferris.dh: verts.	1964
	JWMAA	354	850	852	odvi	sarcoma in an aged w-t dee	kistner.tp: haves	1971
	JWMAA	373	331	335	odvi	epizo hemorrhag dis. n dak	hoff.gl: richard/	1973
						······································	,0-,,	
	NAWTA	4	244	249	odvi	prevalenc, dis, para, mich	whitlock.sc	1939
	NAWTA	26	203	211	odvi	odhe, curr status, brucell	fay.ld	1961
						··· , · · · · · · · · · · · ·	.,	
	NEZTA	20-10	194	195	odvi	pleurisy, pericarditis in	andrews, jrh	1972
	NFGJA	101	118	123	odvi	tuberculosis in a wild dee	friend,m: kroll,/	1963
	NFGJA	191	92	97	odvi	dermatoph cutane streptoth	stone, wb	1972
•	NFGJA	192	184	184	odvi	anoth case, dermatophilosi	stone,wb	1972
	NFGJA	251	1	15	odvi	mortility, fawns, seneca a	opezio,jp	1978
	PAVEA	5	164	173	odvi	spontane bluetongue, texas	<pre>stair,el; robins/</pre>	1968
	PCGFA	12	206	208	odvi	surv brucell, leptospi, se	<pre>shotts,eb; greer/</pre>	1958
	SABOA	133	329	333	odvi	phoma peyronellae, zoo path	<pre>gordon,ma; salki/</pre>	1975
	SBJOD	12	61	61	odvi	fibroma skin tumors, kentuc	heuer, de: harley/	1 9 75
	TNWSD	33	35	44	odvi	1975, hemorrhagic dise, nj	<pre>mcconnel1,pa; lu/</pre>	1976
	VTPHA	151	133	135	odvi	thalamic ependymoma in a w	nettles,vf; vande	1978

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odvi continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR WDABB 1---3 35 35 odvi mastitis, wild w-tail deer evans, w; friend, 1965 WDABB 3---1 38 39 odvi lymphosarcoma in a w-t dee debie jg; friend, 1967 WDABB 3---3 98 101 odvi listeria monocytoge, feces mccrum, mw; evela/ 1967 WDABB 3---3 102 odvi škin tumors, new york deer friend,m 104 1967 WDABB 3---4 152 155 odvi chlamydia antibody study debbie, jg 1967 WDABB 4---1 12 12 odvi salmonella, captive fawns debbie, jg 1968 WDABB 5---2 68 72 odvi meth, leptospi surv, ontar kingscote, bf 1969 WDABB 6---4 226 228 odvi anthrax epizootic, w-t dee kellog, fe; prest/ 1970 WDABB 6---4 295 298 odvi leptospiral antibodies, us shotts, eb, jr; hay 1970 WDABB 6---4 384 odvi bluetongue vir, pregnan de thomas, fc; traine 1970 388 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJVRA 35--4 523 525 odhe exp trans malig catarr, col pierson, re; stor/ 1974 AJVRA 38--3 411 412 odhe nonleth exp inocu, hemorrh stauber, eh; farr/ 1977 CAFGA 20--3 181 odhe life hist, food hab, calif dixon, js 1934 282 CAFGA 58--2 141 144 odhe rabies in a deer, californ cappucci, dt, jr; 1 1972 CELLB 9---3 489 494 odhe endogenou type c rna virus aaronson, sa; tro/ 1976 JAVMA 169-9 937 938 odhe fetal macer, suppur metrit stauber, eh; olso/ 1976 JAVMA 170-3 334 339 odhe eval, anaplas carrier, uta renshaw, hw; vaug/ 1977 JOMAA 36--2 308 310 odhe "parrot mouth", rocky mt robinette, w1; ald 1955 JWIDA 9---4 314 319 odhe epizoo studies anaplasm, o peterson,kj; kis/ 1973 JWIDA 10--1 74 76 odhe dermato mycosis, alberta chalmers, ga; barr 1974 JWIDA 10--2 166 odhe investigation, tansy ragwo dean, re; winward, 1974 169 odhe testicular atrophy in cali demartini, jc; con 1975 JWIDA 11--1 101 106 JWIDA 12--3 427 434 odhe card agglut test, anaplasm howarth, ja; hoka/ 1976 JWIDA 12--3 459 463 odhe tularemia in a mule deer emmons, rw; ruski/ 1976 JWIDA 16--1 89 98 odhe wasting disease, encephalo williams, es; youn 1980 JWMAA 3---4 360 1939 360 odhe large fibroma, head, neck honess, rf JWMAA 35--2 205 209 odhe diarrhea capt fawns, esche kramer,tt; nagy,/ 1971 36 1942 NAVEA 23--1 34 odhe hemorrhagic septicem, utah quortrup, er SCIEA 91--- 168 168 odhe anaplasmosis, deer, calif boynton, wh; woods 1940 WDABB 4---4 143 143 odhe phenylketonuria, mule deer studier, eh; ewing 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJVRA 37--5 615 617 ceel survey of anaplasmosis, id vaughn, hw; rensh/ 1976 APAVD 1977- 166 1977 176 ceel capture myopathy in elk haigh, jc kronevi,t; holmb/ 1977 AVCSA 18--2 159 167 ceel lens lesions in the elk BVJOA 116-6 284 287 ceel louping ill, virus, scotla dunn,am 1960 ceel enzootic ataxia of red dee terlecki,s: done/ 1964 BVJOA 120-7 311 321 BVJOA 129-1 73 78 ceel enzyme change, cns, ataxic robinson, n 1973 COVEA 69--4 402 410 ceel eval anaplas card agg test magonigle, ra; eck 1979 ceel capture myopathy in elk in lewis, rj; chalme/ 1977 JAVMA 171-9 927 932 JOMAA 13--4 371 372 ceel bang's disease, yellowston rush,wm 1932 JWIDA 6---2 144 48 ceel bluetongue virus, nor amer murray, jo; traine 1970 JWIDA 9---2 129 ceel exp infect, epizoo hemorrh hoff,gl; trainer, 1973 132 ceel reprod, dis, physiol blood vaughn, hw; knigh/ 1973 JWIDA 9---4 296 301 JWIDA 11--3 330 ceel listeria-monocytog, roosev martyny, jw; botz1 1975 334 JWIDA 12--3 386 389 ceel yersiniae, roosevlt, calif martyny, jw; botzl 1976 JWIDA 14--1 74 81 ceel brucellosis in elk, part 1 thorne, et; morto/ 1978 JWIDA 14--3 280 ceel brucellosis, effects, tran thorne, et; morton 1978 291 snyder, sp; davie/ 1979 JWIDA 15--2 303 306 ceel embryonal nephroma JWIDA 15--2 307 ceel myxosarcoma in a wapiti snyder, sp; davies 1979 308 JWIDA 15--3 379 ceel eval anaplas card agg test renshaw, hw; mago/ 1979 386 JWIDA 15--4 529 ceel yersiniae, soil, infec ran botzler, rg 1979 532 JWMAA 15--3 284 287 ceel actino, osteogenic sarcomo drake, ch 1951 NEZTA 26--5 130 131 ceel salmonellosis, dee, calves mcallum, hjf; fam/ 1978 NZJSA 9---2 399 408 ceel brucel, lepto, salmo, n ze daniel, mj 1966 VETRA 88-23 616 617 ceel tuberculos, atypic mycobac hime, jm; keymer,/ 1971 VETRA 100-9 187 187 ceel bovine viru diarr antibody mcmartin, da; sno/ 1977 VETRA 102-- 484 ceel exper challenge, mycobacte orr,mb; hunter,a/ 1978 485 VETRA 104-6 120 ceel outbreak mal catarrhal fev reid, hw; buxton, / 1979 123 VETRA 105-- 574 ceel exper pneumonia, ovine chl mcmartin, da; hun/ 1979 576 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJPAA 15--4 445 454 alal encephalitis, maine, minne king, ls 1939 alal rata, arbovi, serol, finlnd brummer-korvenkon 1973 AJTHA 22--5 654 661 alal "inclusion-body-like" conf villar, ja; rehbin 1977 AVCSA 18--2 287 289 CAFNA 91--1 94 95 alal trauma induc paralys, calf chalmers, ga: barr 1977 alal continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CNJMA 22--1 9 20 alal brucellos, elk isl, albert corner, ah; connel 1958 CNJMA 22--7 244 248 alal moose "sickness", nov scot benson, da 1958 alal moose "sickness" n scot II benson,da 1958 CNJMA 22--8 282 286 COVEA 32--3 241 alal further stud, disease, III fenstermacher,r; 254 1942 alal capture myopathy in a moos haigh, jc; stewar/ 1977 JAVMA 171-9 924 926 alal rata, brucel, 1965-74, usa fox, md; kaufmann, 1977 JIDIA 136-2 312 316 JOMAA 39--1 139 alal paras, dis, wells gray her ritcey, rw; edward 1958 145 JWIDA 8---1 95 98 alal fatality, range overextens miller, fl; broug/ 1972 JWIDA 11--2 116 wobeser,g; runge, 1975 121 alal odvi, arthropathy JWMAA 17--2 217 218 alal brucellosis in a moose jellison,wl; fis/ 1953 JWMAA 24--1 44 52 alal narcosis, moose, nicotine rausch, ra; ritcey 1961 JWMAA 38--1 42 alal parelaphostrongylus tenuis gilbert, ff 1974 46 NCANA 101-- 23 50 alal infect, parasi dis, n amer anderson, rc; lank 1974 WDABB 3---4 183 184 alal johne's disease in a moose soltys, ma; andre/ 1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AEHLA 25--4 246 252 rata obser, natural-focal zoono rausch,rl 1972 AJTHA 22--3 400 403 rata doca, arboviruses in finla saikku,p 1973 APAVD 1976. 40 44 rata systemic mycosis, rein her kennedy, s; monta/ 1976 AVCSA 14--2 292 300 rata struc reind cornea, kerati winqvist,g; rehbi 1973 AVCSA 17--1 107 108 rata isolation of neisseria ovi kummeneje,k 1976 AVCSA 17--4 488 494 rata pasteurellosis in reindeer kummeneje,k 1976 AVCSA 18--1 54 64 rata keratitis in reindeer, bac rehbinder,c; glat 1977 AVCSA 18--1 65 74 rata keratitis in reindeer, myc rehbinder, c 1977 AVCSA 19--1 129 132 rata cutan malig lymphome, immu kummeneje,k; popp 1978 AVSPA 66--1 1 27 rata clin epizoot studies kerat rehbinder, c 1977 BJDEA 99--6 647 654 rata obmo, parapoxvir infection falk, es 1978 **BPURD 1---- 498** 506 rata q fever and alaska caribou hopla, ce 1975 CJMIA 24--2 129 135 rata brucellosis, inoc experime rausch, rl; huntle 1978 CJPEA 64--1 82 82 rata clin lab, epidem brucellos brazeau,m; kien,/ 1973 rata continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 36--5 819 835 rata entodiiniinae, canad arcti lubinsky,g 1958 CJZOA 36--6 937 959 rata diplodiniinae, canad arcti lubinsky,g 1958 CJZOA 48--5 1023 1027 rata brucellos, reinde, ba gr c broughton,e; cho/ 1970 rata dis condit, parasi, bar gr broughton, e; chou 1969 FPWTA 33--- 30 39 JAVMA 175-9 996 998 rata osteomyelitis, arthritis i haigh, jc; pharr,/ 1979 JWIDA 6---3 136 139 rata rangif brucell, alask cani neiland, ka 1970 JWIDA 6---4 483 487 rata isol bunyamwera group arbo hoff,gl; stalati/ 1970 140 JWIDA 8---2 138 rata cutan fibr papill, bg cari broughton,e; mil/ 1972 JWIDA 11--1 45 53 rata rangiferine brucell, alask neiland, ka 1975 JWIDA 11--4 465 rata radiog exam, mandib lesion miller, fl; cawle/ 1975 470 rata mandibular lesions, alaska doerr, jg; dieteri 1979 JWIDA 15--2 309 318 JWIDA 15--3 433 rata arterial disease, alaska dieterich, ra; lui 1979 435 JZAMD 10--1 31 34 rata clostridial myositis herron,aj; garma/ 1979 NOVTA 25--4 196 202 rata clostrid perfringens enter kummeneje,k; bakk 1973 NOVTA 26--7 456 458 rata encephal, neuritis, calves kummeneje,k 1974 VETRA 105-3 60 61 rata contagious ecthyma, reinde kummeneje,k; krog 1979 WDABB 4---4 227 236 rata brucellos, caribou, alaska neiland, ka: king/ 1968 WDABB 6---4 483 rata isolatio, arbovirus, carib hoff,gl; spalati/ 1970 487 rata keratitis, proced sim sola offosson,a; rehb/ 1978 ZEVMA 25A-1 48 56 ZEVMA 25A-2 89 109 rata keratitis, induced in mice rehbinder, c; moe/ 1978 ZEVMA 25A-2 110 128 rata keratitis, experim induced rehbinder,c; glat 1978 ZOGAA 37... 134 142 rata reticulo sarcoma, reindeer beyer, j; kronberg 19 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJVRA 33--5 1013 1016 anam bluetongue virus, pronghor hoff,gl; trainer, 1972 APAVD 1972-15 16 anam frostbite in antelope horn simmons, 1g 1972 CNVJA 16--1 3 9 anam odvi, necrobacillosis, sas wobeser,g; runge/ 1975 JAVMA 171-9 918 923 anam capture myopathy in canada chalmers, ga; barr 1977 JWIDA 8---1 67 71 anam caseous lymph adenitis, pr clark,ka; robins/ 1972 JWIDA 10--1 60 62 anam infectious pododermatitis chalmers,ga; barr 1974 JWIDA 12--3 347 348 anam rabies in a pronghorn ante wempe, jm 1976 JWIDA 12--4 488 491 anam reovirus, neo-nat diarrhea reed, de; daley, c/ 1976 JWIDA 13--3 323 326 anam anaplasmo reservio, mantan jacobson, rh; wor/ 1977

anam continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR JWMAA 30--3 488 496 anam pronghorn die-off, texas hailey,tl; thoma/ 1966 JWMAA 33--4 1032 1033 anam clostridium, hand-rear ant nagy,jg barber,/ 1969 TAEPA 3011- 1 15 anam hard yellow liver dis, rum dollahite, jw; oh/ 1971 TAEPA 3460- 1 57 anam hard yel liv fatty cirrhos bailey,em,, jr; u/ 1977 TAMSA 63--1 27 29 anam selenomonas, new, calif an chattin, je; herm/ 1944 XAGCA 154-- 1 anam bibi, doca, anaplasmosis 10 stiles,gw 1939 ZTMPA 24--2 192 197 anam game anaplasmosis: isolati lohr,kf; meyer,h 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS---- YEAR AJVRA 38--8 1183 1185 bibi demo of bacteriocin activi chengappa, mm; car 1977 CAFNA 86--2 127 132 bibi parasites, diseases, canad choquette, lpe; b/ 1972 CNJMA 22--1 9 20 bibi brucellos, elk isl, albert corner, ah; connel 1958 CNVJA 2---5 168 bibi tubercul, wood buf nat'l, choquette, 1pe; g/ 1961 174 JAVMA 100-- 19 22 bibi tuberculosis in the buffal hadwen,s 1942 JAVMA 171-9 913 917 bibi malignant catarrhal fever ruth, gr; reed, de/ 1977 JOMAA 13--4 371 372 bibi bang's disease, yellowston rush,wm 1932 JWIDA 11--3 395 397 bibi absence, anaplasma infecti peterson,kj; roby 1975 JWIDA 14--3 329 332 bibi serol surv brucell, canada choquette, lpe; b/ 1978 bibi paras, dis, anthrax, canad novakowski,ns; c/ 1963 NAWTA 28--- 233 239 WDABB 5---3 206 207 bibi septicemic pasteurellosis heddleston,kl; ga 1969 WMBAA 16--- 1 52 bibi biol, mngt, wood buf na pk fuller,wa 1966

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 157-- 1507 1511 ov selenium-responsive diseas muth,oh 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

od--

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJVRA 33--1 87 90 ovca id, char, mycoplasma-argin al-aubaidi, jm; t/ 1972 CJZOA 51--5 479 482 ovca stress, lympho stim, phyto hudson, rj 1973 JAVMA 161-6 669 672 ovca parainflu 3 virus, rocky m parks, jb; post,g/ 1972 JAVMA 173-9 1178 1180 ovca chronic frontal sinus, ost paul, sr; bunch td 1978 198 ovca neoplasia captive mammals effron, m; griner/ 1977 JNCIA 59--1 185 ovca hemorrag septice, rocky mt potts.mk JOMAA 18--1 105 106 1937 JWIDA 6---3 169 170 ovca mycoplas, pneumo, rocky mt woolf,a; kradel,/ 1970 JWIDA 9---1 7 11 ovca electrophor, chron pneumon woolf, a; nadler, / 1973 JWIDA 9---1 12 ovca mortality, captive bighorn woolf,a; kradel,d 1973 17 JWIDA 10--2 107 ovca serol survey, viral infect parks, jb; england 1974 110 JWIDA 10--3 228 231 ovca vaccina trials, bluetongue robinson, rm; hai/ 1974 JWIDA 11--1 26 ovca oram, contag ecthyma, can samuel,wm; chalm/ 1975 31 JWIDA 15--2 221 227 ovca johne's disease, colorado williams, es; spr/ 1979 JWIDA 15--4 499 503 ovca isolation, chlamydial agen pearson, hj; engla 1979 JWMAA 31--1 165 168 ovca bluetongue, desert bighorn robinson, rm; hai/ 1967 JWMAA 35--2 270 274 ovca contagiou ecthyma, rocky m blood, da 1971 NAWTA 3---3 893 897 ovca diseases, rocky mt nati pk potts,mk 1938 WDABB 2---2 34 37 ovca myxovirus parainfluenza-3 howe,dl; woods,g/ 1966 WDABB 3---2 74 76 ovca virus isola, lung, nemato forrester, dj; wad 1967 WIDIA 23--- 1 14 ovca pasteurellosis, rocky mt post,g 1962

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AVCSA 19--3 461 462 obmo contagious ecthyma (orf) i kummeneje,k; krog 1978 VTYMA 65-11 1063 1067 obmo corynebacter infect, domes beckley,jc; diete 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWIDA 13--2 135 136 oram contagious ecthyma, britis herbert,dm; samu/ 1977 JWIDA 15--2 221 227 oram johne's disease, colorado williams,es; spr/ 1979 JWMAA 35--4 752 756 oram white muscle disease herbert,dm; cowan 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR APAVD 1977-136 145 many avian tuberc, cerv tub tes kollias, gv, jr 1977 AUVJA 51--4 203 208 many epizootiology of bluetongu hourrigan, jl; kli 1975 CNJMA 22--1 9 many brucellos, elk isl, albert corner, ah; connel 1958 20 JANSA 32--5 1037 1041 many atherosclerosis in ruminan wiggers, kd; jaco/ 1971 JIDIA 124-2 217 222 pathogen epiz hemorr disea debbie, jg; abelse 1971 JIDIA 136-2 312 316 many brucellosis in usa 1965-74 fox, md kaufmann, 1977 many diseas agents trans, flies krinsky,wl 1976 JMENA 13--3 225 275 JWIDA 13--1 74 79 many viruses, capt, free, alber thorsen, j; karst/ 1977 JZAMD 7---4 5 7 many symp on mycobac infec, zoo montali, rj 1976 NZJSA 10--4 949 963 many survey of diseases, new ze daniel, jm 1967 SABOA 13--3 329 333 phoma (peyronellaea), zoop gordon,ma; salki/ 1975 TAMSA 70--2 185 many selenomonas, calif mammals herman, cm; sayama 1951 187 VETRA 94-24 559 559 many brucellosis in wildlife mcdiarmid,a; matt 1974 VETRA 96-23 503 506 many manag deer, exp stud, foot gibbs, epj; mcdia/ 1975 VETRA 99-20 398 398 many isolation rotavirus fr dee tzipori,s; caple/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CBCPA 64A-3 411 418 dosh nature of erythro sickling butcher,pd; hawkl 1979 JANSA 17-1 224 236 dosh doca, white muscle disease blinko,c; dye,wb 1958 JONUA 99-3 331 337 dosh selenium def myopathic lam whanger,po; wesw/ 1969 CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR

AJTHA 21--1 100 109 wldl buttonwillow virus, califo hardy,jl; lyness/ 1972 AMFOA 52... 24 26 biga when animals get sick rush,wm 1946 CNVJA 19--5 139 139 wldl brucellosis and wildlife hudson,m 1978 DABBB 33B-7 3378 3379 wiru epizootiological study, vi hoff,gl 1973 JNCIA 59--1 185 198 mamm nature, rate of neoplasia effron,m; griner/ 1977 continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOHYA 81--2 189 196 windbrne bluetng, portugal sellers, rf; pedg/ 1978 JWIDA 13--- 74 79 wiru viruses from captive, free thorsen, j; karst/ 1977 JWIDA 14--3 351 354 rumi rotavirus infect, neonatal eugster, ak; stro/ 1978 NAWTA 12--- 520 523 biga the hoof and mouth disease broughton, ib 1947 WIDIA 42--- 1 6 ungu epizo hemorrhag dis, alber chalmers, ga; van/ 1964

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 165-8 735 735 axax mycobacterium bovis infect sawa, tr; thoen, c/ 1974 JAVMA 165-- 998 axax mycobac bovis infec, hawai sawa.tr; thoen,c/ 1974 999 JAVMA 174-8 841 843 axax doca, dada, paratuberculos riemann,h; zaman/ 1979 RVETA 29--3 126 130 axax polyarterit nodosa-like le neumann,f; nobel/ 1972 VETRA 98-26 525 526 axax tuberculosis, whipsnade pa jones.dm; manton/ 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AVCSA 7---4 330 363 caca dis resemb paratuberculosi hillermark,k 1966 JCVPA 84--2 215 220 caca ceel, dada, foot-and-mouth forman, aj; gibbs, 1974 NOVTA 28-11 539 546 caca mycobacteriosis, caus pige joergensen, jb; c1 1976 VETRA 98--6 116 116 caca isolatio louping-ill virus reid, hw; barlow,/ 1976 VETRA 104-- 214 215 caca adenoma in british roe dee craig, wa 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR 752 AJVRA 36--6 749 ceni in vivo erythrocy sickling parshall, cj, jr; / 1975 AJVRA 36--6 753 756 ceni ophthalmic manif, induc si vainisi, sj; pars/ 1975 JCVPA 85--3 361 366 ceni stud, foot-and-mouth disea gibbs, epj; herni/ 1975 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 173-9 1243 1243 dada malig hyperthermia, etorpi pertz,c; sundberg 1978 JWIDA 15--2 299 301 dada adrenal cortical adenoma schmidt,re; flet/ 1979 VETRA 98--4 74 74 dada case of bovine tuberculosi wilson,p; harring 1976 VETRA 104-- 450 453 dada 4 cases of chondrodystroph baker, jr; ashton/ 1979 VETRA 105-- 573 dada yersiniosis in free-living chapman, di; chap/ 1979 574 CODEN VO-NU BEPA ENPA ANIM KEWO----- AUTHORS----- YEAR PZSLA 136-3 477 483 elda malig catarrh, pere david d tong, eh; senior, / 1961 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 12-30 445 452 bibo foot-and-mouth dis, poland podgurniak,z 1967 RVTSA 26--2 165 171 bibo isol mal catarrhal fever v straver, pj; bekk/ 1979 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ASISA 29--- 535 535 ruru coronary arterioscl, free- guarda, f; cornagl 1976

UNIT 1.3: ABNORMALITIES

Abnormalities occur in wild species, but the frequencies of abnormal characteristics are not known. The basic principle that must operate is that abnormal individuals are less likely to live and reproduce than normal ones. This statement is based on the assumption that abnormalities are detrimental.

Some abnormalities are physical and easily recognized, such as abnormal antlers and dental anomalies. Some are physical but internal; fetal abnormalities are an example. Some are metabolic; a whitetail fawn at Cornell's Wildlife Ecology Laboratory had a severe calcium metabolism problem, mobilizing from the bones and excreting calcium with a concomittant loss of bone structure. The fawn died in a few days. Such a case would never survive in the wild, of course.

The number of references on abnormalities is small. Some of the cases described are curious instances, while others have physiological or genetic implications. The vast majority of abnormalities in wild ruminants go unnoticed.

REFERENCES, UNIT 1.3

ABNORMALITIES

SERIALS

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS	AUTHOR S	YEAR
PCGFA	13	174	177	od	poisonous plants, s e dise	hayes,fa; jenkins	1959
CODEN	vo-nu	BEPA	ENPA	AN IM	KEY WORDS	AUTHORS	YEAR
COVEA	454	538	547	odvi	degenerative myopathy	hadlow,wj	1955
JCVPA	822	219	221	odvi	intraocular lacrimal gland	wvand.ds: lehav./	1972
JCVPA	874	557	568	odví	anophthalmia, microphthalm	fulton, ab; alber/	1977
JOMAA	484	674	676	odvi	genital hypoplasia, w-t de	marburger,rg; ro/	1967
JWIDA	71	1	2	odvi	ventral hernia in a w-t de	wobeser,g; maclen	1971
JWIDA	84	311	314	odvi	antler malform, leg injury	marburger, rg; ro/	1972
JWIDA	84	320	320	odvi	scrotal hernia in a w-t de	<pre>schlegel,mw; lee/</pre>	1972
JWIDA	94	356	3 58	odvi	multiple anomalies, fetus	wobeser,g; runge,	1973
JWIDA	112	237	240	odvi	male pseudoherma, antl doe	<pre>scanlon,pf; urbs/</pre>	1975
JWIDA	114	497	501	odvi	congenital anomalies, neon	barrett, mw; chalm	1975
JWIDA	122	143	147	odvi	6 repor visual defec, mich	howard, dr; krehb/	1976
NFGJA	191	32	46	odvi	mandibul, dental anomalies	free,sl; bergstr/	1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR BAPBA 22--1 67 72 ceel induc antler grow, pedicle jaczewski,z; krzy 1974 JEZOA 195-2 247 250 ceel induc antler growt, polled lincoln,ga; fletc 1976 JWIDA 8---4 319 319 ceel injurious antler anomaly i schlegel,mw; lee/ 1972 JWIDA 15--2 303 306 ceel embryonal nephroma, wapiti snyder,sp; davie/ 1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CAFNA 91-1 94 95 alal trauma induc paralys, calf chalmers,ga; barr 1977 JWIDA 11--2 116 121 alal odvi, arthropathy wobeser,g; runge, 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJVRA 25--- 1783 1785 rata immunity study, reind herd bolton,wd; murray 1964 JRPFA 57--1 127 130 rata abnormal testes in reindee leader-williams,n 1979 JWIDA 7---4 307 309 rata polydactylism. nw manitoba miller,fl; brough 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 171-9 918 923 anam capture, myopathy, alberta chalmers,ga; bare 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 171-9 988 989 bibi atresia ani, recto vag fist marler, rj; leipo/ 1977
CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JZOOA 180-4 518 523 dada mandib, maxill dental abno jackson, je 1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEARJZAMD 6---2 2828axax ventricular septal defect gendron, ap1975

TOPIC 2. NATURAL AND INTRODUCED TOXINS

Toxins are poisonous substances. Many toxins are produced by microorganisms, and many by plants and animals. There are natural toxins, a part of the natural world, and problem-causing only when local concentrations are great enough to upset the physiology and metaoblism of an individual.

Introduced toxins may be for specific target organisms, or they may be compounds introduced into the environment that have side effects. Examples of the former are herbicides and pesticides and of the latter, inorganic fertilizers that may reach unexpected concentrations in water.

Brief comments and fairly short lists of references are included in UNITS 2.1 and 2.2.

REFERENCES, TOPIC 2

NATURAL AND INTRODUCED TOXINS

BOOKS

TYPE PUBL CITY PGES ANIM KEY WORDS----- AUTHORS/EDITORS-- YEAR

edbo tcpc rege 380 anim (p. 131-151) magnesium, en jones, jb, jr; bloun 1972 edbo tcpc rege 380 anim (p. 153-175) magnesium, en jones, jb, jr; bloun 1972 edbo acpr nyny 270 many fate of pestic in 1g anims ivie, gw; dorough, 1977

UNIT 2.1: NATURAL TOXINS

The definition of a natural toxin is based not on the presence of an element or compound in the diet of an animal, but on the quantity of the element or compound in the diet. Thus even minerals required in the diet, for example, may become toxic and cause metabolic disorders when present in too large quantities. Selenium is such a mineral. Several different naturallymoccurring elements and compounds are described as poisonous substances in PART II, TOXICOLOGY in the Merck Veterinary Manual (1967).

Natural toxins usually do not become a problem for wild ruminants unless range conditions are poor and the animals are in stressful conditions. As in the case of parasites and diseases, prevention of a problem is better than having to cure it. Good management practices which result in productive animals on a productive range will eliminate any potential harmful effects of natural toxins almost entirely. If animals are forced to consume potentially toxic plants on overused range, the danger of plant poisoning increases. Over 50 different plants found in the United State are listed in the Merck Veterinary Manual as poisonous. The list includes such common plants as Quercus (oaks) and Prunus (cherries). It may be that wild ruminants have a higher tolerance to the effects of such species than domestic ones, or it may be simply a matter of conditioning and adaptation.

The reference lists include a number of papers that may be of interest, as well as some that are on both the poisonous plant list and the nutrition lists in CHAPTER 11.

LITERATURE CITED

Siegmund, O. H., Ed. 1967. The Merck Veterinary Manual. Merck and Co., Inc. Rahway, N. J. 1686 pp.

REFERENCES, UNIT 2.1

NATURAL TOXINS

SERIALS

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS				AUTHOR S	YEAR
AZATA	75	1	39	od	expe	erimental	feeding	of	de	nichol,aa	1938

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ECOLA 12--2 323 333 odvi mt laurel, rhododen, foods forbes,eb; bechde 1931 WDABB 3---2 42 46 odvi fluorosis in deer karstad,1 1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR FLUOA 8---4 182 191 odhe odvi, industrial fluorosis kay,ce; tourange/ 1975 JANSA 41--1 412 412 odhe eff high molybdenum intake nagy,jg; chappel/ 1975 JWIDA 12--1 39 41 odhe fluorosis in black-ta deer newman,jr; yu,m 1976 PYTCA 17--4 803 803 odhe prunasin, amelanchier alni majak,w; bose,rj/ 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR cee1 1 ł CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR alal CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR rata CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JOMAA 36--1 146 146 anam chokecherry toxic to antel ogilvie,sl 1955 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR bibi CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 157-- 1507 1511 ov selenium-responsive diseas muth, oh 1**9**70 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JONUA 99--3 331 337 dosh selenium def myopathic lam whanger, po; wesw/ 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JRMGA 28--4 252 256 anim areas molybden tox, grazin kubota, j 1975

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR ECOLA 52-5 935 939 anim/model, absorp, ret ing ele goldstein ra; elw 1971 JRMGA 29--5 356 363 rumi majr plant toxic, west u s james, lf; johnson 1976 JRMGA 30--3 237 238 rumi toxicity, larkspur extract olsen, jd 1977

UNIT 2.2: INTRODUCED TOXINS

Pesticides, herbicides, air pollutants, solid wastes, radioactive fallout... all represent toxins introduced by the activities of man that may affect the physiology and metabolism of wild ruminants. The toxicity to animals is often a secondary effect that, at least a few years ago, was given little thought Now, after seeing some of the effects on a variety of wild species, more care is given to the handling of introduced compounds that are potentially toxic. Short-term financial advantages may still be gained by rapid dumping of waste products, but these can never be justified if the long-term functions of the ecosystem are disrupted.

The effects of introduced toxins must not be measured only in terms of dead animals, but also in relation to the health of the entire ecosystem. The health of the ecosystem can hardly be known or understood if the normal physiological functions of its components are not known and understood. Thus the need for courses in physiology in wildlife curricula is clear to those of us who think about the functions of individuals and populations rather than of populations alone. Population responses to introduced toxins will ultimately be understood only by having a knowledge of the physiology and metabolism of the productive members of the population.

REFERENCES, UNIT 2.2

INTRODUCED TOXINS

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR NATUA 182-- 1294 1294 cerv fall-out radioact, antlers hawthorn, j; duckw 1958

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR HLTPA 34--6 691 696 od-- iodin-129 in man, cow, dee ballad, rv; tan, s/ 1978 HLTPA 36--4 516 od-- pluton conc, nucl plnt dee kirkham, mb; adri/ 1979 519 JWMAA 17--- 33 od-- ammate in the diet of deer haugen, ao 1953 36 PAAZA 21--6 16 17 od-- DDT in man, anim, soil ari roan,cc; laubsch/ 1969 PCGFA 5.... 1 7 od-- ammate in the diet of deer haugen, ao 1951

CODEN VO-NU BEPA ENPA ANIM OEY WORDS----- AUTHORS----- YEAR BMSCB 1---- 39 56 odvi ecol, physiol eff insectic morris, je 1972 919 odvi odhe, industr fluorosis in day,ce; touragne/ 1975 FLUOA 8.... 182 HLTPA 25--5 515 516 odvi rapid field-mon cesium-137 rabon,ew; johnson 1973 JWMAA 28--1 45 49 odvi sampl antler for stront-90 schultz, v 1964 JWMAA 29--1 33 1965 38 odvi stron-90 lev in antle, man schultz, v JWMAA 29--1 39 odvi sex, age, stront-90 accumu schultz,v; flyge, 1965 43 JWMAA 34--4 887 odvi repro, grow, tis res diel murphy, da; korsch 1970 903 1972 MISTB 6---- 166 166 odvi ecol, physiol eff insectic morris, je 737 odvi calcium, stront, age, antl cowan, rl; hartso/ 1968 PSEBA 129-3 733

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BECTA 8---1 1 9 odhe pesticide levels in deer benson, ww; smith, 1972 BECTA 13--3 316 323 odhe metabo, oral dose DDT, DDE watson,m; pharao/ 1975 FLUOA 8.... 182 191 odhe odvi, industr fluorosis in kay,ce; touragne/ 1975 HLTPA 11--- 1407 1414 odhe accum fallout cs 137, colo whicker, fw; farr/ 1965 HLTPA 21--6 864 866 odhe tissue distri radio cesium hakonson, te; whic 1971 JANSA 31-1 235 235 odhe infl pesticides rum bacter barber, ta; schwa/ 1970 odhe accumu stront-90, yearling longhurst, wm; sc/ 1978 JWMAA 32--3 621 623 NATUA 214-- 511 odhe radionuclides, liver, colo whicker, fw; walt/ 1967 513 NAWTA 36--- 153 162 odhe eff pesti on rumen bacteri barber,ta; nagy,j 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ceel

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AUMGA 65--5 284 287 rata lichen, cari, high radiati pruitt,wo 1963 BPURD 1--- 64 rata radiocesiu cycl, reind/car holleman,df; luic 1975 70 CATRB 4---1 1 12 rata osteocytic osteolysis belanger,1f 1969 CJZOA 54--6 857 rata tritium wat dilut wat flux cameron, rd; whit/ 1976 862 ESTHA 1--11 932 rata radioacti, 137 CS, alaskan blanchard, rl; kea 1967 939 HLTPA 18--2 127 rata lead-210, polonium-210, ala blanchard, rl; moo 1970 134 rata stront-90, annu vari, swed persson, br HLTPA 20--4 393 402 1971 rata cesium-137, seas pat, alas hanson,wc HLTPA 20--6 585 591 1971 rata radiocesium, lichen, alask holleman, df; lui/ 1971 HLTPA 21--5 657 666 rata radioactiv in lichns, swed mattsson, ljs HLTPA 29--1 27 41 1975 JEENA 69... 260 262 rata famphur, oxy analog, resid ivey, mc; palmer, / 1976 RHDRA 11... 487 509 rata radioactiv, sweden, nuc magi,a; snihs, jo/ 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BECTA 3---5 269 273 anam insecti residues, s dakota moore,gl; greich/ 1970 CJZOA 50--2 213 216 anam accumul stront-90, females mitchell,gj; hon/ 1972

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

bibi

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEARBECTA 19--1 2331ovca chlor hydroc residu in fat turner, jc1978BECTA 21... 116124ovca trnsplac mvmt pestic resid turner, jc1979

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CNJNA 45--3 197 202 doca selenium, hair, malnutriti hidiroglou,m; ca/ 1965

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR WIDIA 37--- 1 11 ceni sika deer, potassium defic christian, jj 1964

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR ECOLA 52--5 935 939 anim model, absorp, ret ing ele goldstein,ra; elw 1971 FLUOA 9.... 73 90 ungu pop var of fluor param, wi kay,e; tourangea/ 1976 HLTPA 29--1 43 51 fall-out pluton, swed lich holm,e; persson,r 1975

TOPIC 3. DRUG EFECTS, SEROLOGY, AND PHYSICAL RESTRAINING DEVICES

This TOPIC 3 contains referrences to three areas of activity that are related to studies of both healthy and diseased animals. Drugs are used to immobilize and tranquilize, serology is important in diagnosis and in the treatment of disorders, and physical restraining devices are used when working with animals. The reference lists will be useful to someone considering different techniques, providing efficient access to the published literature.

UNIT 3.1: DRUG EFFECTS

Drugs are used for two main purposes when working with wild ruminants: immobilization and tranquilizing. An immobilizing agent does just that and nothing more; drugs that immobilize make it impossible for an animal to move, but the animal is fully conscious and presumably receiving environmental stimuli without being able to exhibit a behavioral response. A tranquilizer reduces the level of awareness of an animal. A tranquilized animal may still be mobile, but not as excitable. Heavy doses of a tranquilizer will also immobilize an animal.

Wild animals, being alert, excitable, elusive, and very hard to handle are prime candidates for immobilizing and tranquilizing drugs during research work. The effects of drugs on the physiology and metabolism are generally not well-known. The importance of several metabolic rhythms in relation to dosage levels is being understood (see CHAPTER 7).

The number of references that follow indicate a fairly large amount of activity in this area, all confined to the last 20 years.

REFERENCES, UNIT 3.1

DRUG EFFECTS

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 31--4 686 692 cerv diazepam, capture handlin thomas, jw: robin/ 1967 NOVTA 22--- 385 400 cerv capt immob with neurolept eriksen, e 1970

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR CNJMA 26--3 57 61 od-- tranquiliz, immobilization cowan,i mct; woo/ 1962 JAVMA 147-- 1099 1101 od-- chlor hydra-halo-nitr-oxi wolff,wa; davis, 1965 JWMAA 41--2 313 315 od-- transm syr to recov imm de lovett,jw; hill,e 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR APAVD 1976- 185 194 odvi succinylcholine in deer kitchen.h 1976 CJZOA 57--4 756 767 odvi infl chem imm blood charac wesson, ja; scanl/ 1979 CJZOA 57--4 768 776 odvi infl chem imm ster hor lev wesson, ja, scanl/ 1979 COVEA 40--- 275 282 odvi anesthetization white-t de severinghaus, cw 1950 odvi trauma, secondary shock in flyger, vf; ridge/ 1962 CPSCA 3.... 236 243 JAVMA 157-5 636 640 odvi immob cap, free, etor hy c woolf,a 1970 JAVMA 167-7 574 576 odvi xylazine immob agent, capt roughton, rd 1975 JWIDA 9---4 336 341 odvi seda immobil etorph & xyla presnell,kr; pre/ 1973 JWIDA 9---4 342 348 odvi combin etorphine, xylazine presidente, pja; / 1973 JWIDA 15--4 537 541 odvi blow-gun syring, captiv de warren, rj; schau/ 1979 JWMAA 17--4 516 520 odvi flaxedil, paralysis, w-t d hall,tc; taft,eb/ 1953 JWMAA 21--2 213 220 odvi nicotine salicylate, captu crockford, ja; ha/ 1957 JWMAA 28--1 173 175 odvi mort, thor area, cap-chur thomas, jw; marbur 1964 JWMAA 29--4 889 890 odvi nicotine, field immobiliza behrend, df 1965 JWMAA 31--4 844 845 odvi device for immob trapped d white, cm 1967 JWMAA 34--1 207 odvi succin chlor, hyaluronidas allen,tj 209 1970 JWMAA 36--4 1034 1040 odvi effe immobil on blood anal seal, us; ozoga, j/ 1972 JWMAA 40--3 447 453 odvi seas var, succ chol immobi jacobsen, nk; arm/ 1976

odvi continued on the next page

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS AUTHORS	YEAR
JZAMD	52	18	18	odvi	ceel, m99, recov w/ m50-50 woolf,a	1974
PCGFA	28	500	506	odvi	<pre>succinylcho, react, physi wesson,ja,III; s/</pre>	1974
WLSBA	54	193	194	odvi	aggress toward immobi deer scanlon,pf; mira/	1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JAVMA 155-7 1080 1084 odhe new tech, restr antlerless sauer, bw; gorman/ 1969 JAVMA 171-9 987 987 odhe odvi, ketamine xylazine im richter, ag 1977 JWIDA 14--4 519 522 odhe methoxyfluran anesth, fawn trindle,bd; lewis 1978 JWMAA 25--2 202 203 odhe anesthetizin deer by arrow anderson, cf 1961 JWMAA 26--3 332 333 odhe succinylcho chloride, colo foyd, rj 1962 JWMAA 27--2 297 299 odhe effects of succinyl chlori pearson, ha; smit/ 1963 JWMAA 32--1 195 197 odhe succinyl chlor, immobiliza miller,fl 1968 JWMAA 33--4 1037 1038 odhe new dev for inject powdere liscinsky,sa; ho/ 1969 JWMAA 37--1 82 86 odhe immob, phencyc hydro chlor dean,r; hines,ww/ 1973

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDS AUTHORS	YEAR
IABLA	6	45	50	ceel	immob of marals by ditilin ushakov, kp; shol/	1971
JAVMA	169-9	888	889	ceel	yg eff, etor hydroc in dam farnsworth,rj; st	1976
JWIDA	133	258	261	ceel	immob, etorphine hydchlori magonigle,ra; st/	1977
JWMAA	233	365	366	ceel	curare, curare-like drugs post,g	1959
JWMAA	263	334	336	ceel	<pre>succinylcholine chloride flook,dr; robert/</pre>	1962
JWMAA	292	339	345	ceel	roosevelt elk, succin chlo harper,ja	1965
JWMAA	394	814	816	ceel	m99, immobili rocky mt elk coggins,vl	1975
VETRA	933	86	87	ceel	caca, ceni, immobilon,deer low,rj	1973
VE TRA	934	113	114	cee1	caca, dada, immobilon,deer chapman,d	1973
VETRA	934	114	115	ceel	immobilon in deer, scotlnd sharman,gam	1973
VETRA	945	85	86	cee1	hypersen isol pop to xylaz fletcher,j	1974
VETRA	99-21	424	424	ceel	anaesthesia in red deer whitelaw,a; fawce	1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR 917 alal hyaluronidase in immob mix haigh, jc JAVMA 175-9 916 1979 JOMAA 51--2 396 399 alal M99 etorphine, immobilizat houston, db 1970 JWMAA 25--3 326 alal narcosis, moose, nicotine rausch, ra; ritcey 1961 328 JWMAA 33--3 534 alal shiras moose, immobilizatn houston,db 537 1969 JWMAA 39--3 634 alal physiologic effect, m-99 e roussel, ye; pate/ 1975 636 alal etorp & xylaz hydroc, immo gasaway,wc, fran/ 1978 JWMAA 42--3 686 690 JZAMD 5---2 26 31 alal immobiliz of alaskan moose franzmann, aw; arn 1974 JZAMD 8---3 22 29 alal fentanyl & xylazine, captu haigh, jc, stewar/ 1977 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 28--1 49 rata newfound1, succiny1, moose bergerud, at; but/ 1964 53 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 31--4 840 842 anam succin chlor, immobilizati beale,dm; smith,a 1967 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR PIAIA 83--2 67 70 bibi immob adult bull, etorphin haugen, ao; swens/ 1976 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 40--1 174 176 ovca succ chlor, phencyc hydroc stelfox, jg; rober 1976 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JWMAA 39--1 112 117 obmo immob, marking, arct canad jonkel,cj; gray,/ 1975 ZOGAA 40... 138 142 obmo sedation of a bull muskox jones,dm 1971

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY WORDSAUTHORS	YEAR
APAVD	1975-	40	42	many	xylazine, exotic anim prac hertzog,re	1975
VTYMA	695	548	551	many	injectab anesth, wld rumin boever,wj; paluch	1974
ZOGAA	35	149	155	anim	eff tranq, anesth, wild an nouvel, j; rinjar/	1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR APAVD 1975- 16 39 ungu clin use rompun, toron zoo rapley, wa; mehren 1975 ZOOAA 1---1 231 238 ungu use of (ROCHE), tranquiliz hirst, sm; kett 1965 ZOOAA 1.... 239 247 ungu experim immobili ungulates bigalke,rc 1965 ZOGAA 33... 182 185 ungu immobil ungula catsk ga fa heck,h; douigh,f 1967

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR JAVMA 130-- 479 482 dogo nicotine, immobiliz, goats hayes,fa; jenkin/ 1957 JWMAA 27--2 292 296 new tech using cap-chur gu green,h 1963 JWMAA 33--4 1037 1038 device, inject powdered dr liscinsky,sa; ho/ 1969

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR BVJOA 131.. 545 548 dada diazepam, sedat & restrain done, sh; lees, p; / 1975 JZAMD 4---2 21 21 dada restrain 2 dee w/ xylazine klide,am; klein,1 1973 VETRA 93--1 26 27 dada immobilon in fallow deer jones, dm; manton, 1973 VETRA 93-16 454 dada immobilisation, fallow dee low, rj 455 1973 VETRA 93-16 455 dada immobilisation fallow dee lees, p; done, sh 455 1973 VETRA 94-16 362 363 dada immobilon-rompun fallow de harrington, r; wil 1974 VETRA 94-17 396 dada powd succ chlo in dart, im scanlon, pf 398 1973 VETRA 100-- 386 387 dada fentanyl & neuroleptic, im haigh, jc 1977

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR VETRA 95--3 71 71 axax tranquilising deer sutherland,c; hod 1974

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY	WORDS				AUTHORS		YEAR
JAVMA	169-9	890	893	elda	immo	obiliz	pere	davids	deer	<pre>smeller,j;</pre>	bush,/	1976

CODEN	VO-NU	BEPA	ENPA	ANIM	KEY WORDS AUTHORS	YEAR
ATRLA ATRLA ATRLA	12-32 12-32 12-32	467 471 473	470 472 475	bibo bibo bibo	narcos, chlor hydra, eth a piwowarczyk,s use of palmer gun, immobil starzynski,w eu bis hybrid, suxamethoni zaniewski,l	1967 1967 1967
ZOPOA	252	85	9 8	bibo	etorphine hydrochlo, immob kania, bf; teuchma	1975

UNIT 3.2: SEROLOGY

Serology is the science dealing with the properties and use of serum, or body fluids. Blood serum is the fluid considered most; it is important for its agents of agents of immunity and its role as an antitoxin. References to serology follow, providing access to the published literature on the wild ruminants.

REFERENCES, UNIT 3.2

SEROLOGY

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS----- YEAR AGVIA 37--2 282 284 od-- antigenic rel, epiz, bluet moore,dl; lee,vh 1972 AJVRA 35-12 1529 1531 od-- appendage-related antigen carson, ca; weisi/ 1974 AJVRA 36--9 1367 1370 od-- leptospiral antibodies in harrington,r,jr 1975 AUVJA 51--4 185 od-- doca, dosh, serolog method boulanger, p; fran 1975 189 JAVMA 132-7 289 292 od-- infect, antibody, anaplasm christensen, jf; / 1958 JAVMA 132-7 293 296 od-- serol surv, leptospi, mass reynolds, im; smit 1958 JAVMA 167-7 565 od-- surv, toxoplas-gondii anti franti,ce: conno/ 1975 568 JAVMA 175-9 911 od-- serologic profile of exoti riemann, hp; rupp/ 1979 913 JWIDA 6---4 479 482 od-- sero ev, arbovir, wis, tex issel,cj; hoff,g/ 1970 JWMAA 30--4 777 780 od-- serol stud, epiz hemorra d wilhelm, ar; train 1966 PCGFA 13... 174 od-- poisonous plants, s e dise hayes, fa; jenkins 1959 177 PIAIA 74--- 78 86 od-- incid, dis antibodie, iowa haugen, ao 1968

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR AJTHA 21--6 979 988 odvi experimental studies with issel,cj; traine/ 1972 AJTHA 23--2 242 245 odvi maternal antibod, jamestow issel,cj 1974 AJVRA 31--5 879 883 odvi serodiognosis, theileriasi gadir,fa; hidalg/ 1970 AJVRA 33-12 2545 2549 odvi elect dist, serol ch, arth sikes,d; kistner/ 1972 CNVJA 3---3 71 78 odvi cul, ser evi, lepto, ontar abdulla,pk; kars/ 1961 odvi continued on the next page

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWIDA 9---3 245 248 odvi infection in texas, califo issel,cj; hoff,g/ 1973 odvi odhe, arbovir serol, n dak hoff,gl; issel,c/ 1973 JWIDA 9---4 291 295 JWIDA 11--2 195 200 odvi serol venez eq enceph, tex smart,dl: traine/ 1975 JWIDA 12--3 322 odvi antibo toxoplasm gond, ont tizard, ir; bille/ 1976 325 odvi transm 2 strains epizootic foster, nm; breck/ 1977 JWIDA 13--1 9 16 WDABB 3---4 152 155 odvi chlamydia antibody study debbie, jg 1967 WDABB 5---4 392 397 odvi serolo surv, arbovirus, ny whitney,e; roz,a/ 1969 WDABB 6---4 295 odvi leptospiral antibodies, us shotts, eb, jr; hay 1970 298 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWIDA 12--3 427 434 odhe card agglut test, anaplasm howarth, ja; hoka/ 1976 JWIDA 13--4 429 431 odhe ceel, anam, survey, titers adrian.wj: keiss, 1977 JWIDA 14--2 193 odhe hemaglut test, epidem, lep cirone, sm; riema/ 1978 202 JWIDA 14--4 523 527 odhe stress, immune system trindle, bd; lewi/ 1978 JWIDA 15--1 25 odhe tech eval immunity brindle, bd; lewis 1979 31 JWIDA 15--3 443 446 odhe serol sympatric sheep, mul hampy, b; pence, d/ 1979 JWMAA 41--3 515 519 odhe preval reactor, lives path stauber, eh; nell/ 1977 WDABB 4---3 78 80 odhe serol surv, arbovir, calif emmons, rw 1968 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR COVEA 69--4 402 410 ceel eval anaplas rapid card ag magonigle, ra; ec/ 1979 FPARA 23--1 25 31 ceel caca, leptospiral an, wild sebek, z; kaasere/ 1976 JOMAA 17--3 250 253 ceel oservat on yellowstone elk mills, hb 1936 ceel brucell, serol, bacti, wyo thorne, et; morto/ 1978 JWIDA 14--1 74 81 JWIDA 14--4 471 478 ceel odhe, serologi study, utah merrell,cl; wrigh 1978 JWIDA 15--3 379 386 ceel eval anaplams card agglut renshaw, hw; mago/ 1979 NATUA 241-- 476 477 ceel antibody to babesia, red d latif, bma; adam, k 1973 RVTSA 23--2 133 138 ceel occur antib to babesia, lo adam,kmg; beasle/ 1977 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

JWIDA 7---2 93 95 alal surv, antibod, bovin virus thorsen, j; hender 1971 JWIDA 7---2 118 119 alal evid, arbovir activ, alber trainer, dd: hoff, 1971

661 alal rata, arbovi, serol, finlnd brummer-korvenkon 1973

AJTHA 22--5 654

CODEN	vo-nu	BEPA	ENPA	ANIM	KEY	WORDS			AUTHORS	YEAR
AJTHA	223	400	403	rata	doca	, arboviru	uses in	finla	saikku,p	1973
AVCSA	173	363	369	rata	doca	, occur an	ntib, c	hlamdi	neuvonen, e	1976

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR JOPAA 56--4 759 767 anam dosh, immun, haemonch isol allen,rw; samson/ 1970 JWIDA 11--2 157 163 anam serol surv, albert, saskat barrett.mw; chalm 1975 JWIDA 16--1 109 109 anam seroepidem survey, idaho stauber,eh; aute/ 1980 TMPRA 25--2 217 226 anam doca, detect fluor agg ant lohr,kf; ross, jp/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS------ AUTHORS------ YEAR JWIDA 9---4 306 310 bibi vaccination against pasteu heddleston,kl; we 1973 JWIDA 14--3 329 332 bibi serol surv brucell, canada choquette,lpe; b/ 1978 JWIDA 14--4 493 500 bibi serol, hemato values, colo keith,eo: ellis,/ 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca in vitro detect, homocytot hudson, rj; bandy/ 1971 CEXIA 8---2 345 354 CJCMA 36--1 69 73 ovca cell adherence react, lung hudson, rj; bandy/ 1972 DABBB 33B-8 3819 3820 ovca devel pasteurella bacterin nash, p 1973 JWIDA 6---2 105 106 ovca electrophoresis, monitorin hudson,r; kitts,/ 1970 JWIDA 7---3 171 ovca immuno globulin ressponse hudson,r; kitts,/ 1971 174 JWIDA 9---1 7 ovca electrophor, chron pneumon woolf, a; nadler, / 1973 11 JWIDA 9---1 12 17 ovca mort, clin, hema, path obs woolf,a; kradel,d 1973 JWIDA 10--2 107 110 ovca serol survey, viral infect parks, jb; england 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ovda

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

obmo

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 35--4 752 756 oram white muscle dis, br colum herbert,dm; cowan 1971

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR AJEPA 90--4 354 358 many serol evid arbovi, wild ru trainer, do; hanso 1969 AJVRA 30-11 2007 2011 many ser evi, blue tongue, n amer trainer, dd; jochi 1969 JANSA 17--1 224 236 doan ser trans, white muscl dis blinko,c; dye,wb 1958 JWIDA 12--3 322 325 mamm prev toxoplas antibod, ont tizard, ir; bille/ 1976 many prev, dist, 4 serotyp smsv smith, aw; akers,/ 1976 JWIDA 12--3 326 334 JWIDA 13--4 429 many brucello, leptospiro, colo adrian, wj; keiss, 1977 431 TMPRA 29--2 223 233 ungu preva trypanosome carriers drager,n; mehlitz 1978 VETRA 92--5 119 119 mamm antib to leptospira grippo twigg, gi; hughes/ 1973

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWIDA 15--2 339 341 dada antigen, herpes virus, alb kinyili,jh; thors 1979

UNIT 3.3: PHYSICAL RESTRAINING DEVICES

Any device used to restrain a wild ruminant has a potential effect on its physiology and metabolism. Restraint devices are necessary for some kinds of studies. A few references with descriptions of restraining devices, cages, etc. are listed. The philosophy of handling of wild ruminants in captivity is very important. I personally believe that they should be treated as naturally and with as little restraint as possible, and not as domestic cattle or sheep that are penned, tied, and herded around at will.

REFERENCES, UNIT 3.3

PHYSICAL RESTRAINING DEVICES

SERIALS

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR CJZOA 57--4 756 767 odvi blood char, inf phys res & wesson,ja,III; s/ 1979 CJZOA 57--4 768 776 odvi ster hor lev, inf phy re & wesson,ja,III: s/ 1979 JWMAA 31--2 359 361 odvi rope truss for restraining thomas,jw; robin/ 1967 JWMAA 38--4 845 847 odvi restr appar, blood samplin mautz,ww; daviso/ 1974

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

odhe

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

ceel

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR

alal

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR JWMAA 42--4 904 908 rata ef han, restr, blood param karns,pd; crichto 1978

CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR anam CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ATRLA 12-32 477 479 bibi immobilizing cage for biso krasinska,m 1967 CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovca CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR ovda CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR obmo CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR oram CODEN VO-NU BEPA ENPA ANIM KEY WORDS----- AUTHORS----- YEAR WLMOA 14--- 1 78 many pharm, physio princ, restr hartoorn,am 1965

CLOSING COMMENTS

CHAPTER 10 includes a large number of references to literature that has not yet been synthesized into ecological considerations. The factors affecting physiology and metabolism must in turn affect productivity, and productivity, in turn, affects population dynamics. The discussions in CHAPTERS 18 and 19 will relate to the literature listed here in CHAPTER 10, especially for mortality data.

Special thanks are due Dr. Susan Wade, College of Veterinary Medicine at Cornell, for sorting the long lists of references into the units in this chapter. We hope that the readily available lists for each wild ruminant species will be of value to those reviewing this literature.

> Aaron N. Moen March 20,1981

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CODEN

ACVTB Acta Veterinaria Brno AEHLA Archives of Environmental Health AESAA Annals of the Entomological Society of America AGBOB Agroborealis AGVIA Archiv fur die Gestamte Virusforschung AJEPA American Journal of Epidemiology AJHEA American Journal of Public Health AJPAA American Journal of Pathology AJTHA American Journal of Tropical Medicine and Hygiene AJVRA American Journal of Veterinary Research AMFOA American Forests AMNAA American Midland Naturalist ANYAA Annals of the New York Academy of Sciences APAVD American Association Zoo Veterinarian Annual Proceedings APMIA Acta Pathologica et Microbiologica Scandinavica APRPA Acta Parasitologica Polonica ASISA Atti della Societa Italiana delle Scienze Veterinarie ASMAC Abstracts of the Annual Meeting of the American Society for Microbiology ATMPA Annals of Tropical Medicine and Parasitology ATRLA Acta Theriologica (Poland) AUMGA Audubon Magazina AUVJA Australian Veterinary Journal AVCSA Acta Veterinaria Scandinavica (Denmark) AVSPA Acta Veteinaria Scandinavica Supplementum (Denmark) AZATA Arizona Agricultural Experiment Station Technical Bulletin BAPBA Bulletin de l'Academie Polonaise des Sciences Serie de Sciences **Biologiques** BECTA Bulletin of Environmental Contamination and Toxicology BEREA Bulletin of Entomological Research BJDEA British Journal of Dermatology BMSCB Bulletin of the Missouri Academy of Science Supplement BPURD Biological Papers of the University of Alaska Special Report BSKBA Bulletin of the University of Osaka Prefecture Series B Agriculture and Biology BVJOA British Veterinary Journal CAENA Canadian Entomologist CAFGA California Fish and Game CAFNA Canadian Field Naturalist CATRB Calcified Tissue Research CBCPA Comparative Biochemistry and Physiology CELLB Cell CEXIA Clinical and Experimental Immunology CJCMA Canadian Journal of Comparative Medicine CJMIA Canadian Journal of Microbiology

CJPEA Canadian Journal of Public Health CJZOA Canadian Journal of Zoology CNDRA Condor CNJMA Canadian Journal of Comparative Medicine and Veterinary Science CNJNA Canadian Journal of Animal Science CNRDA Canadian Journal of Research, Section D, Zoological Sciences CNREA Cancer Research CNSVA Conservationist CNVJA Canadian Veterinary Journal COVEA Cornell Veterinarian CPSCA Chesapeake Science CVTJA Ceylon Veterinary Journal DABBB Dissertation Abstracts International DKBSA Doklady (Biological Sciences, Proceedings of the Academy of Sciences of the USSR) (English translation of DOBTA) DOBTA Doklady Akademii Nauk SSSR Biological Science Section (for English translation see DKBSA) ECOLA Ecology ENAMA Entomologica Americana ESTHA Environmental Science and Technology ETBRA Entomologische Berichten (Amsterdam) EVETB Environmental Entomology FLUOA Fluoride FPARA Folia Parasitologica FPWTA Transactions of the Federal-Provincial Wildlife Conference GRBNA Great Basin Naturalist GRLEA Great Lakes Entomologist HLTPA Health Physics IABLA Izvestiya Akademii Nauk Azerbaidzhanskoi SSR, Seriya Biologicheskikh Nauk IJAHA Indian Journal of Animal Health IJLAA Indian Journal of Animal Sciences INFIB Infection and Immunity IVEJA Indian Veterinary Journal IZSPB USDA Index Catalogue of Medical and Veterinary Zoology Special , Publication JAASA Journal of the Alabama Academy of Science JANSA Journal of Animal Science JAVMA Journal of the American Veterinary Medical Association JCVPA Journal of Comparative Pathology JEBCA Journal of Entomological Society of British Columbia JEENA Journal of Economic Entomology JEMEA Journal of Experimental Medicine JEZOA Journal of Experimental Zoology JFUSA Journal of Forestry

JIDEA Journal of Investigative Dermatology JIDIA Journal of Infectious Diseases JKESA Journal of the Kansas Entomological Society JMENA Journal of Medical Entomology JNCIA Journal of the National Cancer Institute JNYEA Journal of the New York Entomological Society JOHLA Journal of Helminthology JOHYA Journal of Hygiene JOMAA Journal of Mammalogy JONUA Journal of Nutrition JOPAA Journal of Parasitology JPROA Journal of Protozoology JRMGA Journal of Range Management JRPFA Journal of Reproduction and Fertility JWIDA Journal of Wildlife Diseases JWMAA Journal of Wildlife Management JZAMD Journal of Zoo Animal Medicine JZOOA Journal of Zoology KLVBA Kongelige Veterinaer-og Landbohojskole Arsskrift LANCA Lancet MISTB Transactions Missouri Academy of Science MMLRA Mammal Review MUATA Minnesota Agricultural Experiment Station Technical Bulletin NATUA Nature (England) NATWA (Die) Naturwissenschaften NAVEA North American Veterinarian NAWTA North American Wildlife and Natural Resources Conference, Transactions of the, NCANA Naturaliste Canadien, Le NEZTA New Zealand Veterinary Journal NFGJA New York Fish and Game Journal NOSCA Northwest Science NOVDA Norsk Veterinaertidsskrift NOVTA Nordisk Veterinaermedicin NPSMD United States National Park Service Scientific Monograph Series NYCOA New York State Conservationist NZASA New Zealand Agricultural Science NZJSA New Zealand Journal of Science OUOKA Outdoor Oklahoma PAAZA Progressive Agriculture in Arizona PAHUA Parasitologia Hungarica PARAA Parasitology PAVEA Pathologia Veterinaria PCGFA Proceedings of the Southeastern Association of Game and Fish Commissioners

PHSWA Proceedings Helminthological Society of Washington Proceedings of the Iowa Academy of Science PIAIA PMASA Proceedings of the Montana Academy of Sciences PNDAA Proceedings of the North Dakota Academy of Science POASA Proceedings of the Oklahoma Academy of Science PPASA Proceedings of the Pennsylvania Academy of Science PPETA Pan-Pacific Entomologist PPPAA Pacific Science Congress Proceedings PRLBA Proceedings of the Royal Society of London B Biological Sciences PSDAA Proceedings of the South Dakota Academy of Science PSEBA Proceedings of the Society for Experimental Biology and Medicine PUNMA Proceedings of the United States National Museum PYTCA Phytochemistry PZSLA Proceedings of the Zoological Society of London RHDRA Radiological Health Data and Reports RSZOA Revue Suisse de Zoologie RVETA Refuah Veterinarith RVTSA Research in Veterinary Science SABOA Sabouraudia SAGCA Science in Agriculture SBJOD Science of Biology Journal SCIEA Science SEHEA Seminars in Hematology SENTD Southwestern Entomologist TAEPA Texas Agricultural Experiment Station Progress Report Transactions of the American Microscopical Society TAMSA Tropical and Geographical Medicine TGMEA TISAA Transactions of the Illinois State Academy of Science TMPRA Tropenmedizin und Parasitologie TNWSD Transactions of the Northeast Section, The Wildlife Society TRCIA Transactions Royal Canadian Institute TRSTA Transactions of the Royal Society of Tropical Medicine and Hygiene VETRA Veterinary Record VIWIA Virginia Wildlife VPARD Veterinary Parasitology VTPHA Veterinary Pathology VTYMA Veterinary Medicine and Small Animal Clinician WDABB Bulletin of the Wildlife Disease Association WGFBA Wyoming Game and Fish Commission Bulletin Wildlife Disease WIDIA WLMOA Wildlife Monographs WLSBA Wildlife Society Bulletin WMBAA Wildlife Management Bulletin WSCBA Wisconsin Conservation Bulletin

XAGCA USDACircular XENOB Xenobiotica

ZEVMA Zentralblatt fuer Veterinaermedizin
ZOANA Zoologischer Anzeiger
ZOGAA Zoologische Garten
ZOOAA Zoologica Africana
ZOPOA Zoologica Poloniae

ZTMPA Zeitschrift fuer Tropenmedizin und Parasitologie

LIST OF PUBLISHERS - CHAPTER TEN

acpr	Academic Press	New York	nyny
ccth	Charles C. Thomas	Springfield, IL	spi1
faog	Food & Agriculture Organization of U. N.	Rome, Italy	roit
hein	Heinemann	London, England	loen
isup	Iowa State University	Ames, IO	amia
lefe	Lea and Febiger	Philadelphia, PA	phpa
plpr	Plenam Press	New York	nyny
tcpc	Taylor County Printing Co.	Reynolds, GA	rege
wbsc	W. B. Saunders Company	Philadelphia, PA	phpa